

Anti-tumor Activity Of Virulent Newcastle Virus With *Urtica pilulifera* On Mammary Adenocarcinoma In mice

الفعالية المضادة للورم لفايروس نيوكاسل الضاري ونبات القريص على سرطانة الغدة اللبنية في الفئران

Abdul Ameer Oda Ismail *

*Clinical Pathology Department .College of Pharmacy, Kerbala University .

Abstract :

The anti-tumor activity of the Virulent Newcastle Virus (VNV) ($LD_{50}10^9$) and water extract of *Urtica pilulifera* leaves (U.P) were evaluated against the murine mammary adenocarcinoma transplanted subcutaneously in mice . The activity was assessed using growth inhibition of tumor volume, relative tumor volume and histopathological examination. Intratumoral injection of (VNV) and *Urtica pilulifera* crude extract was given different doses resulted in a prominent suppression of tumor statistically orally with significant with 82%, the relative tumor volume was also reduce and the suppression of tumor size in this group more than others. Histopathological examination show massive necrosis associated with fibrosis and lymphocytic with macrophagic infiltration.

الخلاصة

تم تقييم الفعالية المضادة للسرطان لكل من فايروس نيوكاسل الضاري (نصف الجرعة القاتلة له $LD_{50}10^9$) والمستخلص المائي لأوراق القريص ضد سرطانة الغدة اللبنية الفأري المغروس تحت الجلد في الفئران. وقد رت هذه الفعالية من خلال تثبيط نمو حجم الورم وحجم الورم النسبي والفحص النسيجي المرضي. ان الحقن المتعدد لفايروس نيوكاسل الضاري داخل كتلة الورم والتجريع المتكرر لمستخلص اوراق نبات القريص ادى الى تثبيط نمو حجم الورم بنسبة 82% وبشكل مهم احصائي و كذلك ضمور حجم الورم النسبي وان تثبيط الورم في هذه المجموعة كان اكبر من التثبيط الحاصل في بقية المجاميع. أما نسيجيا، فقد تم ملاحظة منطقة واسعة من التخر بالاضافة الى ارتشاح الخلايا اللمفاوية والبلاعم الكبيرة مع ترسب النسيج الليفي.

Introduction

The idea of using viruses in tumor therapy is based on observation of remission of human cancers following natural viral infections such as mumps or measles(1). Newcastle Disease Virus (NDV) has been suggested as a very promising agent for tumor therapy(2) because this avian virus in mammalian species including man has interesting anti-neoplastic (3) as well as immune stimulation(4) properties, it can be oncolytic (5) and it can activate host immune cells to produce cytokines (6) and to become cytotoxic against tumor cells(7). Herbal remedies are widely used for the treatment and prevention of various diseases and often contain highly active pharmacological compounds (8). Urticaceae was reported as one of the effective medical plants, these plants are being consumed without any report of adverse effects (9). The *U. pilulifera* L. is commonly used as a remedy for diabetes mellitus (10), anti-tumor activity (11), and anti oxidant effect(12).

According to the previous studies on the NDV and plant extract, it was of interest to estimate its anti-cancer effect on mammary adenocarcinoma transplanted in mice.

Material And Methods

Newcastle Disease Virus(NDV).

The virulent lytic strain NDV (Iraqi strain) was obtained from department of Pathology and Poultry Disease, College of Vet. Med. Baghdad University. A stock of infectious virus was propagated in embryonated chicken eggs (9-11 days) harvested from the allantoic fluid, purified from debris by cold centrifugation (3000 rpm, 30 min, 4°C)(13).

NDV was quantified by the hemagglutination and hemagglutination inhibition test (14). The measurement of Embryonated Leathal Dose 50% (ELD₅₀) of Virus was conducted according to the Karber method (15).

Plant(U.pilulifera)

The leaves of U. p were obtained from Iraqi Center of Cancer and Medical Cytogenetics.

Extraction and separation

Twenty-five grams gram of U. p powder were exhaustively extracted with 300 ml of Distille water, the mixture was put in water bath (50°C, 24 hours), filtered, centrifuged at (4000 rpm, 30min) and the extract concentrated by rotary evaporatour (16).

Tumor cells

The cell line of murine mammary adenocarcinoma (AM3) were obtained from Iraqi Center of Cancer and Medical Cytogenetics. Transplantation of tumor cells subcutaneously according to (3).

Laboratory animals.

Female balb \C mice (22-26gm) (8-12 weeks) were procured from Iraqi Center of Cancer and Medical Cytogenetics, in March 2004. They were housed in boxes in a control environment (temp. 25°C) with standard laboratory diet and water. The animals were classified in to six groups each of them contain five mice. All of them were injected subcutaneously by (0.25ml) suspension of tumor cells. When the tumor nodule growth S\C and reached about (6-12 mm), all animals were subjected to different treatments as follows.

Group I: The mice were treated orally with (1ml) U. pilulifera four doses and injection of the NDV (0.1ml, LD₅₀ 10⁹) directly in the tumor (Intratumoral) (I.T) four doses, three days intervals between doses.

Group II: Similarly treated as in the group I, but injection of the virus was through peritoneum (Intraperitoneally).

Group III: The mice were treated with (1ml) U.P orally only four doses (three days intervals between doses).

Group IV: The mice were treated with (1ml) U.P orally and injection of allantoic fluid (virus-free) directly in the tumor (I.T). four doses, three days intervals between doses. This group is considered a positive control group for group I. (C+VE).

Group V: Similarly treated as in the group IV but injection of allantoic fluid was done Intraperitoneally. Positive control group for group II.

Group VI: The mice injected with tumor cells only without treatment. A negative control group to all groups (C-VE).

Determination of tumor volume(T.V).

The tumor volume of treated mice and tumor bearing mice was estimated according to Grote, *etal* (17). Tumor volume = $A \cdot B^2 \cdot 2 = \text{mm}^3$

A = Length B = Wide

Determination of Growth inhibition(G.I).

The measurement of G.I% was conducted according to Phuangsab, *etal* (18).

$$G.I\% = \frac{T.V \text{ in untreated group} - T.V \text{ in treated group}}{T.V \text{ in untreated group}} \times 100$$

Determination of Relative Tumor Volume (R.T.V)

The determination of R.T.V. was followed according to Phuangsab ,*etal*(18).

$$R.T.V.\% = \frac{T.V(\text{day X})}{T.V(\text{day 0})} \times 100$$

Pathological study.

Postmortem examination was done on dead mice. Tissue samples were taken from tumor mass and subjected to routine histopathological study, consisted of fixation and dehydration . Five to six microns-thick sections were stained with haematoxylin & eosine staining and examined with light microscope at various magnifications(19).

Statistical analysis:

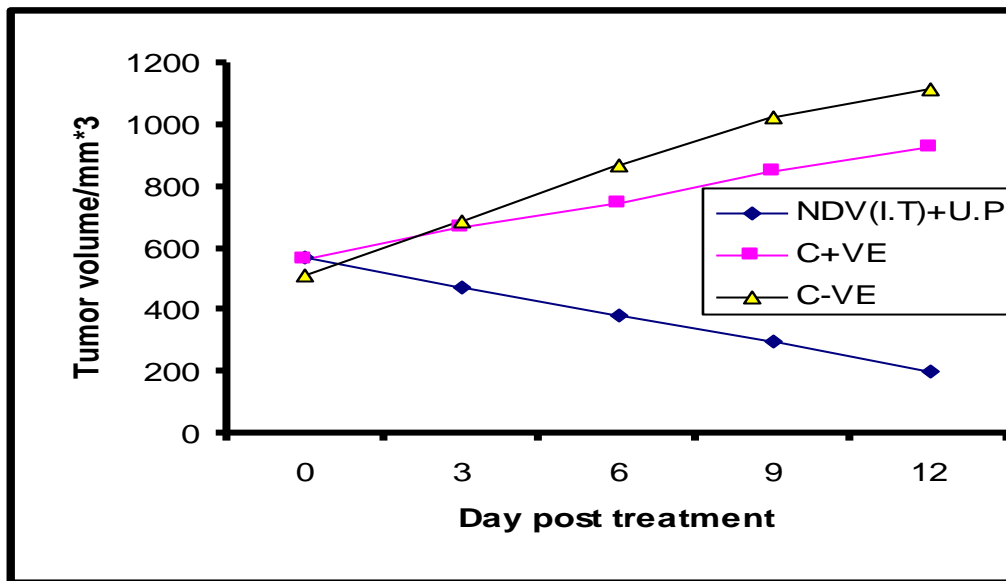
Programe of SAS(2001) to analysis the result of study by Least Significant Differences(LDS)was used in this study.

Results

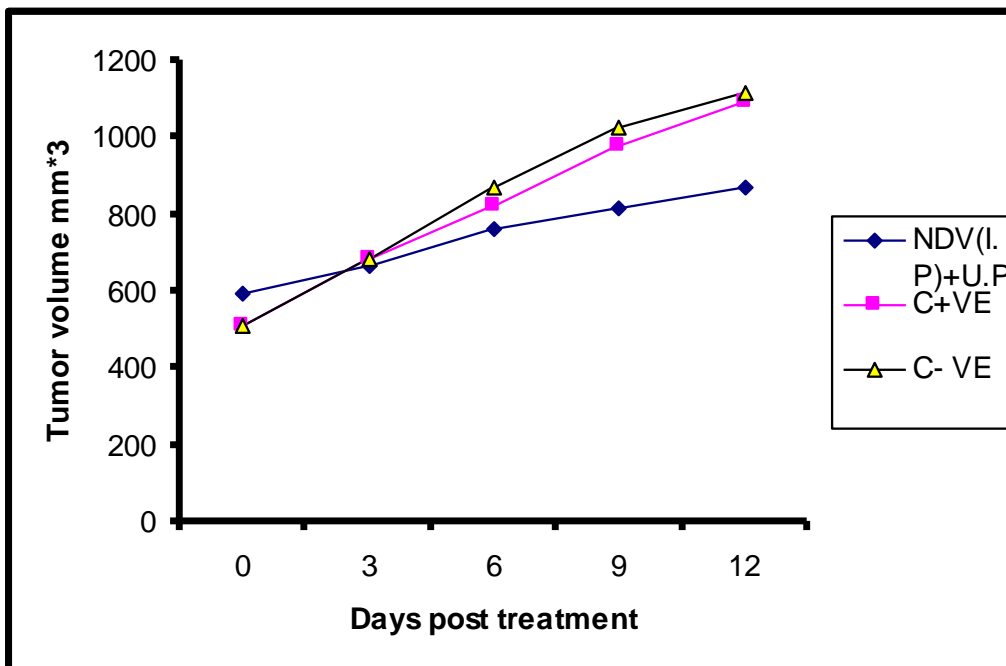
Tumor volume.

The first group which was treated with U.pilulifera orally and injected with virulence newcastle virus directly in the tumor(I.T,resulted inhibition of tumor growth after three days from first dose.This inhibition of tumor growth continued after four doses (12) days from start of treatment.The percent of growth inhibition at the end of treatment was (82%),statistically significant ($P<0.0001$) compared with negative group control , and (78%) statistically significant ($P<0.0001$) compared with positive group control.(Fig.1)

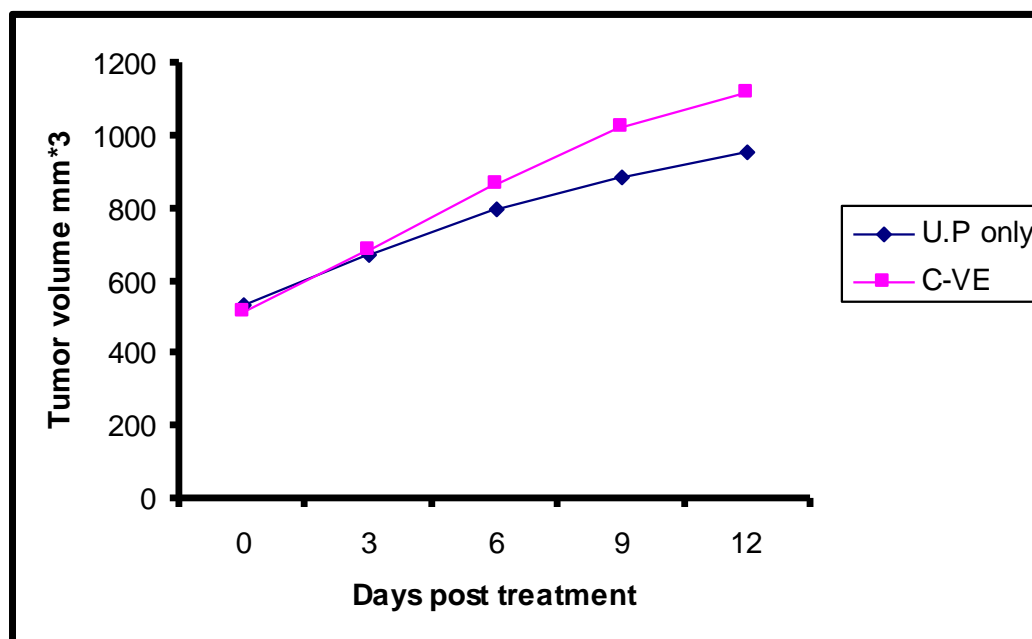
The second group which was injected by the virus intraperitonealy, showed an inhibition of tumor growth after first dose injected and the percent of growth inhibition at the end of treatment was (22%), statistically significant ($P<0.01$) compared with the negative control group ,and (20%) compared with the positive control group.(Fig.2). The third group which was treated with U.pilulifera orally only, showed inhibition of tumor growth and the percent of growth inhibition at the end of treatment was(14%) compared with the negative control group (Fig.3). The growth inhibition in the first group was more than statistically significant ($P<0.0001$) compared with second &third groups.



Fig(1).The inhibition of tumor growth after treatment by multiple injection of NDV(I.T)with U.P (orally).



Fig(2). Anti-tumor activity of NDV(I.P) with U.P(orally).

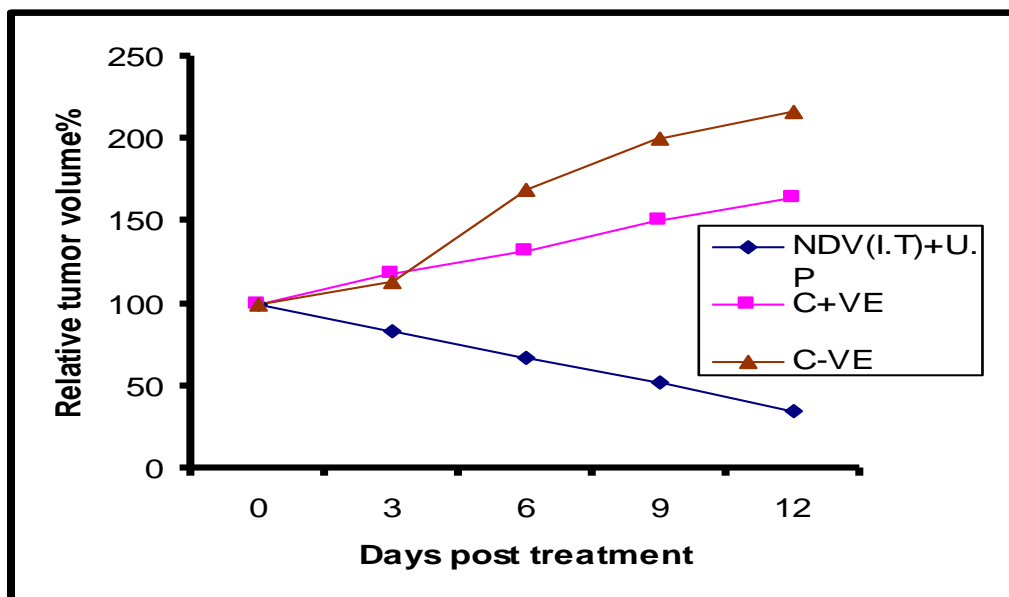


Fig(3). Anti-tumor activity of the *U.pilulifera*(orally).

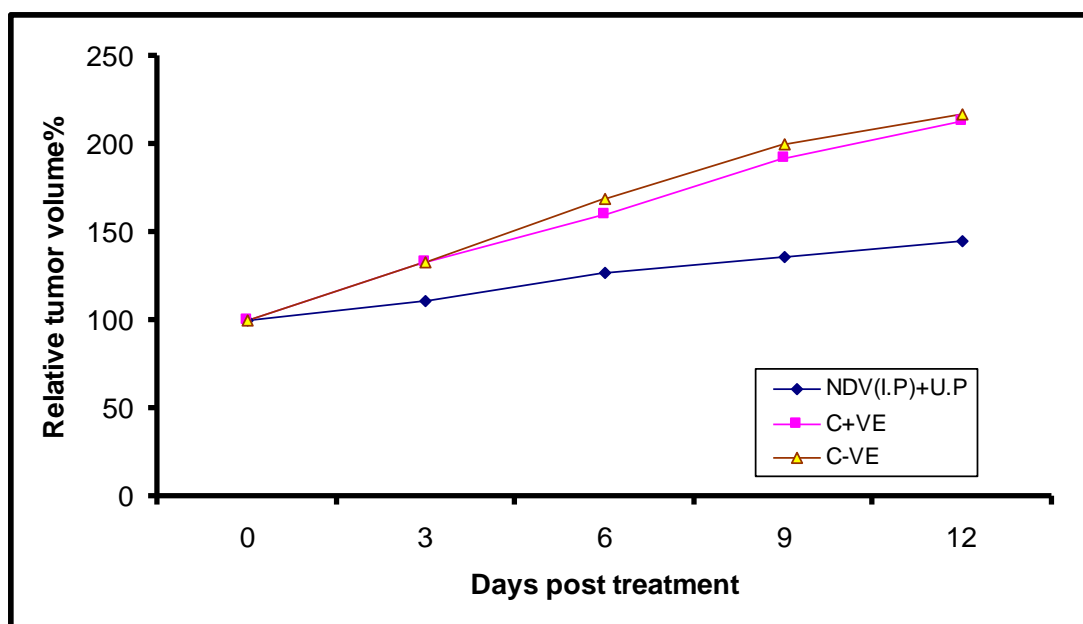
Relative tumor volume(R.T.V)

The first group, show that reduction in (R.T.V) was observed after three days from the start of the treatment, this reduction were continuous to end of the experiment(four doses,12 days)from the start of the treatment. The percent of R.T.V was (35%)statistically significant($P < 0.0001$)compared with the tumor size before the start of the treatment(fig.4).That means (65%) of tumor size was regression. The second group which was injected by virus intraperitonealy show an increase in R.T.V about(45%)at the end of experiment compared with the tumor size before the start of the treatment , but this increase was less than the increase which occurs in the negative (117%) and positive (113%) control groups compared with tumor size before the start of the treatment(fig.5), that means the tumor size in the treated group was less than about(72%)from the tumor size in untreated groups at the end of experiment.

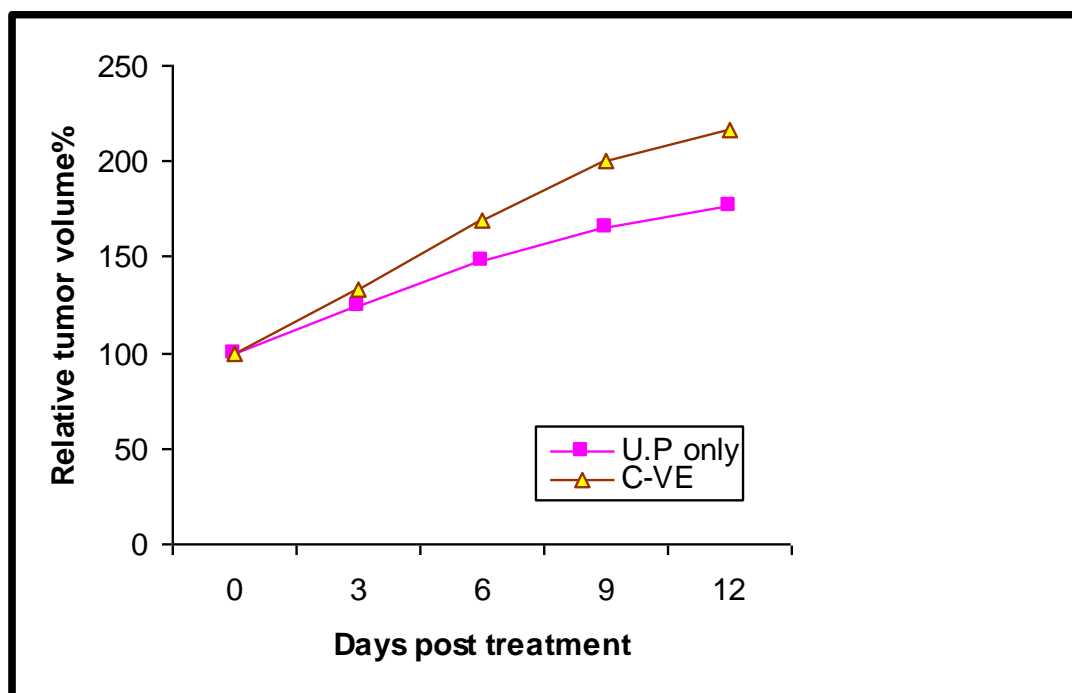
The third group which was treated with *Urtica pilulifera* only show an increase in the R.T.V (77%) at the end of experiment compared with the size of tumor at the start of experiment ,but the tumor size in this group was less than about (40%) from the tumor size in untreated group at the end of experiment(fig.6).



Fig(4).Effect of treatment by NDV(I.T)and U.P on R.T.V.



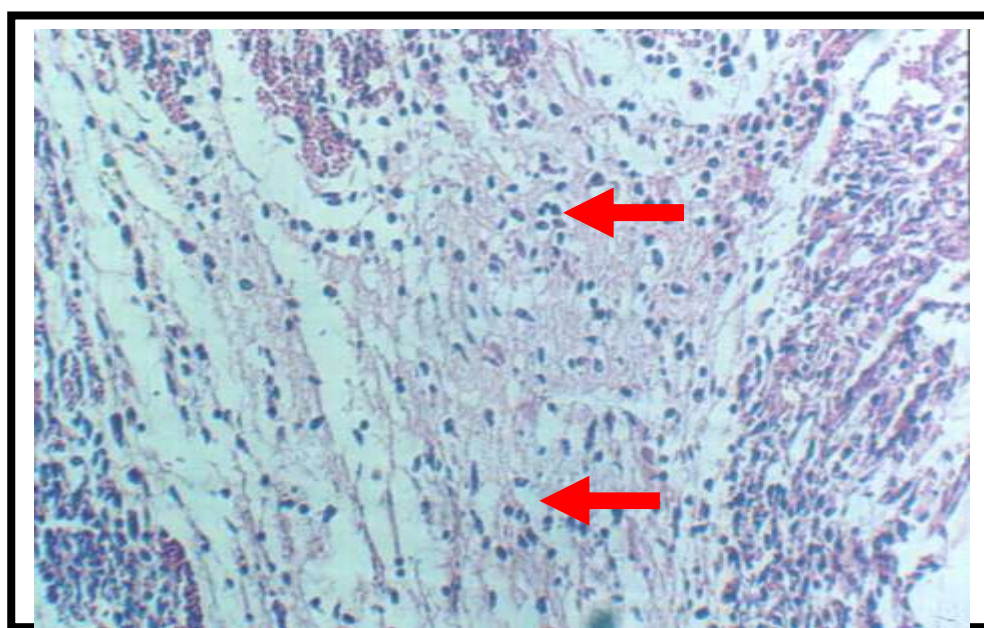
Fig(5).Effect of treatment byNDV(I.P)with U.P on R.T.V.



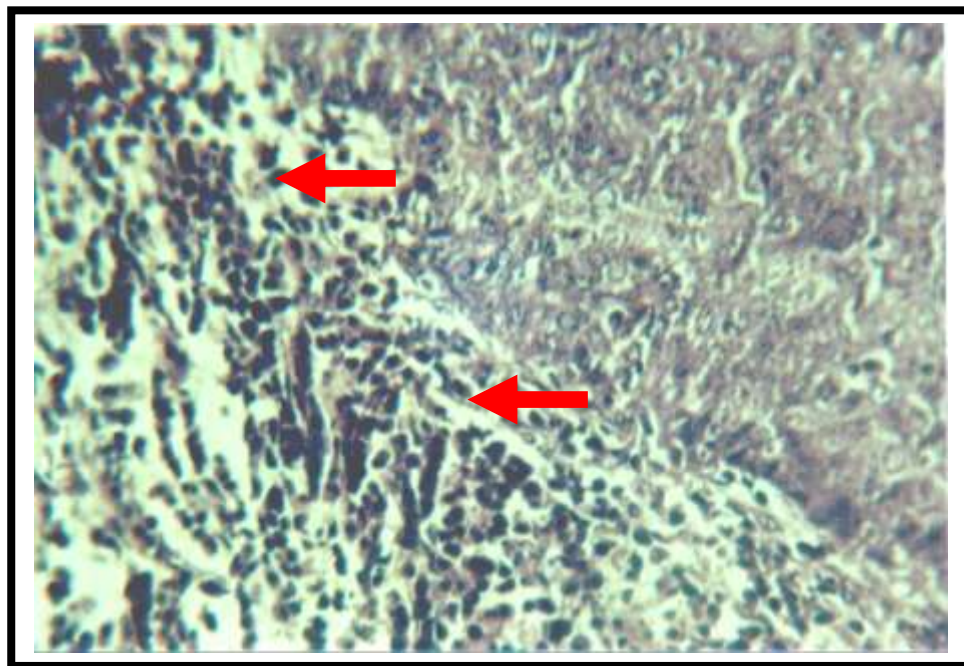
Fig(6).Effect of U.P only on R.T.V.

Histopathological examination.

The histopathological effects of treating tumors with intratumoral injection of virulence NDV with *U.pilulifera* (orally), show wide area of massive necrosis of tumor cells associated with infiltration of inflammatory cells such as neutrophils, plasma cells, lymphocytes and macrophages ,in addition fibrous connective tissue replacement the necrotic area with vacillation in cytoplasm of tumor cells like apoptosis(fig.7). The histopathological effects of intraperitoneally treatment by virulence NDV with *U.pilulifera*(orally), necrosis in some area of tumor associated with infiltration of lymphocytes and macrophages was observed . There are present area of tumor cells without necrosis(fig.8).



Fig(7).Massive area of necrosis with inflammatory cells and fibrous connective tissue.(H&E 200 X)



Fig(8).Area of necrosis with inflammatory cells and area of tumor without necrosis(H&E 200X).

Discussion

In this study, virulence NDV (Iraqi strain) had a pronounced anti-tumor effect when it was given by both local (intratumoral) and systemic (intraperitoneal) routes. Virulent strain was selected for the study since it has been previously given to human and animals by a variety of routes and had demonstrated a good safety profile at the dose used (3,20). A dose of 1×10^9 ELD₅₀ was administered I.T in the first group lead to inhibition of tumor growth and relative tumor volume. Phuangsab *et al* (18) administered NDV viral oncolysate preparation to twelve mice with subcutaneous xenografts of epidermoid carcinoma were intratumoral treatment, the virus caused an average of (81 %) growth inhibition and reduction in relative tumor volumes for the mice treated with NDV were also significantly.

Cytolysis strains of NDV were selectively replicate in and rapidly kill adverse group of human tumor cells (21). Oncogenic transformation increase the sensitivity of malignant tumor cells to NDV cytolysis (22). Stodji *et al* (23) suggested that this selectivity is based upon a cancer-specific down regulation of the interferone pathway within the tumor cells that leaves these cells open to infection with certain types of lytic viruses.

The inhibition of tumor size and the histopathological changes (necrosis and inflammatory cells) in the group treated with NDV (I.T) were occurred more than in that group treated with NDV (I.P), there are numerous possibilities explain the mechanism of this such as a- binding of virus to normal tissues or cells. b-inactivation of virus before it reaches the tumor site or c-regrowth of virus resistant tumor cells after the initial challenge (18).

In addition, *Urtica pilulifera* herb extract showed a good antioxidant activity, this support the anti-cancer effect of the plant extract (11). Also, Seeram *et al* (24) found that the flavonoids, especially these contain C-7 and C-4 hydroxyl groups act as anti-mutagen and anti-malignant agent and the phytochemical studies showed that U.P contains the same flavonoid (25). Mak *et al* (26) explained the mode of action of genistein glycosides as anti-tumor drug. *U. pilulifera* contains isoflavone genistein glycoside and tyrosine kinase which inhibits proliferation of prostate cancer cell through the induction of apoptosis (27).

The increase of inflammatory cells (lymphocytes and macrophages) in necrotic area of tumor is due to the fact that these cells are the first one to arrive at sites of infection, where they can

release chemokines and proteases that can in turn recruit both specific and nonspecific immune effector cells (28) . They can also release toxic granules against neighboring cells, suggesting their potential anti-tumor activity(29).

References

- 1- Sinkovics,J.G.and Horvath ,J.C. (1993).New development in the Virus therapy of cancer: A historical review . Inter virology (1993),36:193-214.
- 2-Pecora,A.;Rizvi,N.;Cohen,G.I.;Meropol,N.J.;Starman,D.; Mar Shall,J.L.; Goldberg,S.;Gross,P.;O'neil,J.D.;Groene, W.S.; Roberts , M.S.; Rabin, H .; Batmat, M.K . and W.S.; Roberts , M.S.; Rabin , H .; Batma t, M.K. and Lorence , R. M. Phase 1 trail of intravenous Administration of PV701 an oncolytic virus ,In patient with advanced solid cancer . Journal of clinical Oncology, (2002) ,20(9):2251 – 2266
- 3-AL-Shamaery,A.M. Study the effect of immune stimulation On the transplanted tumor cells in white mice. Athesis of Master.Vet.College.University of Baghdad(2003) .
- 4- Quade,G..Complementary and alternative medicine statement for Health professional Newcastle disease virus. National cancer Institute . (2004). Pp:1-55.
- 5 - Sinkovics ,J.G. and Horvath,J.C. Newcastle disease virus . Brief History of its oncolytic strains. Journal of Clinical Virology ,(2000)16(1):1-15.
- 6- Zorn,V.;Dallmann,I. and Grosse,J. Induction of cytokines and cytotoxicity against tumor cells by Newcastle disease virus . Cancer Biother(1994).9(3):225-235.
- 7- Krin , D.H. and Mc Cormick,F.. Replicating viruses as selective cancer therapeutics .Molecular Medicine Today(1996),:519-527.
- 8-Saad,B.;Azaiz,H and Said.D. Traditional and perspectives of Arab herbal medicine :Areview Evid Based Complement Alternat .Med(2005).;2:475-9.
- 9-Ali-Shtayeh, M.S.; Yaniv, Z and Mahajna, J. Ethnobotanical survey in the palestinian area: A classification of the healing potential of medicinal plants .J.Ethnopharmacol(2000)...73:221-232.
- 10-Kavalali,G.H.;Tuncel,S.Goksel and Hatemi,H.H. HypoGlycemic activity of Urtica pilulifera in streptozotocin Diabetic rats.J.Ethnopharmacol. (2003)..84:241-245
- 11- Abdel-kader,M.;Mahmoud,A.H.;Motawa,H.M.;Wahba,H.E. And Ebrahim, A.Y. Anti-tumor activity of Urtica pilulifera on Ehrlich Ascites Carcinoma in mice. Asian Journal of Biochemistry(2007) ..2(6):375-385.
- 12-Mahmoud,A.H.;Matawa,H.M.;Wahba,H.E.andEbrahim,A.Y. Study of some antioxidant parameters in miceliver affected with Urtica pilulifera. Asian Journal of Biochemistr. (2007) . .1(1):67-74.
- 13- Alexander,D.J.. Newcastle disease virus and other paramyxo virus in :Isolation and identification of avian pathogens, (4th ed). Edited by, M.W.;Pearson,J.E. and Reid,W.M. American Association of Avian Pathologists .U.S.A(1998).Pp.156-163.
- 14- Hanson,R.P.. Newcastle disease .In: Isolation and Identification of avian pathogen (2nd ed.) edited by Hitchner ,S.B.; Domermuth ,C.H.;Purchase ,H.G. andWilliam,J.E. (1980) Pp.63-66.
- 15- Allan ,W.H.;Lancaster,J.E.and Toth,B..Newcastle disease vaccines.Their Production and Use.FAO.Italy(1978),Pp.1-9.
- 16-Sakai,Y.;Nagase,H.;Ose,T.;Yamada,A.;Hibi,M.andYamada, F. Antimutagenecity of extracts from crude drugsIn chinese medicine.Mutat.Res(1986)..1741-4.
- 17-Grote,D.;;Russell,S.J.; Cornu,T.I.; Cattaneo,R.;Vile, R.;Poland,G. A. and Fielding,A.K . Live attenuated measles virus induce

- regression of human lymphoma xenografts in Immuno-deficient mice. *Blood*, (2001) 97(12):3746-3754.
- 18-Phuangsab, A.; Lorence, R.M.; Reichard, K.W.; Peoples, M.E. and Waltser, R.J.. Newcastle disease virus therapy of human tumor Xenografts : anti tumor effects of local or systemic administration. *Cancer Letters*, (2001) 172:27-36.
- 19-Culling, C.F.A.; Allison, D.T. and Barr, W.T. Cellular Pathology techniques. 4th. ed., London Butter Worth (1985). 155-163.
- 20-Ismail, A.O. Immunopathological therapy of murine mammary adenocarcinoma transplanted subcutaneously in mice. Athesis of PhD. Vet. College. Univ. Baghdad (2005).
- 21-Reichard, K.W.; Lorence, R.M.; Cascino, C.J.; Peebles, M. E. and Walter, R.T. Newcastle disease virus selectively Kills human tumor cells. *J. Surg. Res.* (1992). 52:448-453.
- 22 - Lorence, R. M.; Reichard, K. W.; Katubig, B. B.; Reyes, H.M.; Phuangab, A.; Sasseti, M.D.; Walter, R.J. and Peoples, M.E. Complete regression of Human fibrosarcoma Xenografts in athymic mice after local Newcastle disease Virus therapy. *Cancer Res* 1994). 54:6017-6021.
- 23- Stojdl, D.F.; Lichty, B.; Knowles, S.; Marius, R.; Atkins, H. Sonenberg, N. and Bell, J.C.. Exploiting tumor –specific defects in the Interferon pathway with a previously unknown oncolytic virus . *Nature Medicine* , (2000) 6 (7) : 821-825.
- 24-Seeram, N.P.; Adams, L.S.; Henning, S.M.; Niu, Y.; Zhang, Y.; Nair, M.G. and Heber, D. In vitro antiproliferative, apoptotic and antioxidant tannins extract are enhanced in combination with other polyphenols as found in pomegranate juice. *J. Nut. Biochem.* (2005). 16:360-367.
- 25-Gao, H.; Kuroyanagi, M.; Wu, L.; Kawahara, N.; Yasuno, T and Nakamura, Y. Anti-tumor promoting constituents from *Dioscorea L.* in JB6 mouse epidermal cells. *Biol. Pharm. Bull.*, (2002). 25:1241-1243.
- 26- Mak, P.; Leung, Y.K.; Tang, W.Y.; Harwood, C and Ho, S.M .Apigenin suppresses cancer cell growth through ERbeta¹. *Neoplasia*, (2006). 8:896-904.
- 27-Lunyin, Y.U.; George, L.B. and Jin-Rong, Z. Genstein and diadzein downregulate prostate androgen-regulated transcript-1 (part-1) gene expression induced by dihydrotestosterone in human prostate LNCaP cancer cells *J. Nut.*, (2003). 133:89-392.
- 28- Coussen, L.M. and Werb, Z. Information and cancer. *Nature*. (2002). 420:860-867.
- 29-DiCarlo, E.; Froni, G.; Lollini, P.; Colombo, M.P.; Modesti, A. and Musiani, P. The intriguing role of poly morpho-Nuclear neutrophils in antitumor reactions. *Blood* (2001, 97: 339-345.