In vitro Study for Tannic and Gallic acids as antioxidant in Diabetes Mellitus

Hattf Bazool Farhood

Chemistry Department, College Of Science, Thi-Qar University, Nassriyah, Iraq

Abstract

Antioxidants are a molecule capable of slowing or preventing the oxidation of other molecule by give up their own electrons to free radicals which start a chain of reactions that damages cells. In this study, we attempted to assessment the antioxidant activity of tannic and gallic acids *in vitro*. These acids extracted from tea specially green tea, pomegranates, persimmons, Most berries, such as cranberries, Citrus, Legumes and Chocolate. The study consist of 150 subject which divided into three groups according to type of diabetes mellitus, smoking and hypertension. Serum lipid peroxidation was promote by incubation with copper ions (copper sulphate) (1 × 10⁻⁴ M) at 37 C at 48 hr. the results of the incubation with copper ions showed that copper ions promoted lipid peroxidation which measured by malon di aldehyde (MDA) while tannic and gallic acids can decreased lipid peroxidation when they used alone or together with concentration (1 × 10⁻⁴ M) for each one.

Keywords: Diabetes Mellitus, Tannic acid, Gallic acid, Smoking,

Hypertension, Lipid Peroxidation

Introduction

Diabetes mellitus (DM) is a disease in which the body does not produce enough, or properly respond to insulin this causes sugar accumulate in the blood, often leading to chronic hyperglycemia⁽¹⁾. Hyperglycemia results in oxidative stress by generating free radicals and reactive oxygen species (ROS) and it is implicated in pathogenesis of DM. The adverse effects of smoking may results from oxidative damage to biologic substances. Such damage could result both from eigarette smoking and from the activation of phagocytic cells that generate ROS⁽²⁾, hypertension is induced by oxidative stress, in particular lipid peroxidation (measured as levels of malondialdehyde (MDA). MDA is formed as an end product of lipid peroxidation. Highly reactive free radicals and oxygen species are present in biological systems from a wide variety of sources. These free radicals may oxidize nucleic acid, proteins, lipids or DNA and can initiate a variety of disease such as diabetes ⁽³⁾⁽⁴⁾⁽⁵⁾.

Antioxidant compounds like phenolic acids, polyphenols and flavonoids scavenge free radicals and thus inhibit the oxidative mechanisms that lead to degenerative diseases. Antioxidant effect of a plant is mainly due to phenolic compounds such as flavonoids, phenolic acids and tannins⁽⁶⁾.Polyphenolic antioxidants are potent free radical terminators⁽⁷⁾.They donate hydrogen to free radical and hence, break the reaction of lipid peroxidation at the initiation step⁽⁸⁾. Polyphenolic antioxidants are a type of antioxidant containing a polyphenolic substructure which numbering over 4000 distinct species⁽⁹⁾. The main source of polyphenol antioxidants are nutritional, since they are found in a wide array of phytonutrient-bearing foods . ⁽¹⁰⁾⁽¹¹⁾⁽¹²⁾.

In this study we choose the tannic and gallic acids as a polyphenolic antioxidant, which tannic acid is a naturally polyphenolic antioxidant which distributed in tea, nettle, wood, berries. Oak wood is very rich in $tannic^{(13)}$. Tannic acid has structure as shown in figure(1) with formula($C_{34}H_{28}O_{21}$) and molecular weight: 772.57 Dalton⁽¹⁴⁾. Tannic acid has many sub names such as: Gallotanic acid, digallic acid, allotannin, tannimum. Tannic acid is a polymer

of gallic acid molecules and glucose. The anti-oxidant of tannic acid is beneficial⁽¹⁵⁾. There is many experimental evidence for the effects of tannic acid against cardiovascular disease, inflammation, diabetes and urinary tract infections⁽¹⁶⁾ and gallic acid is a polyphenolic antioxidant compound which distributed in sumac, witch hazel, tea leaves, oak, and other plants⁽¹⁷⁾. Gallic acid has structure as shown in figure(1) with formula($\mathbf{C}_7\mathbf{H}_6\mathbf{O}_5$) and molecular weight: 170.12 Dalton⁽¹⁸⁾. Gallic acid is found both free and as part of tannins. Gallic acid acts as an antioxidant and help to protect our cells against oxidative damage which was found to show cytotoxicity against cancer cells, without harming healthy cells⁽¹⁹⁾.

Figure (1): Tannic acid Gallic acid

Tannic acid and Gallic acid may have the potential to become the lead compound in the development of new types of antidiabetic pharmaceuticals that are able to reduce blood glucose levels⁽²⁰⁾. These compounds appear to aid in diabetes control and in reducing the complications associated with this disease⁽²¹⁾.

Tannic and Gallic acids were tested on pancreatic cells, which produce the hormone insulin in the presence of glucose (sugar). Small blood vessels, called

capillaries, are damaged in diabetes as a result of elevated blood sugar levels⁽²²⁾.

It is important to study the effect of tannic and gallic acids as a polyphenolic antioxidant on oxidation promoted by copper ions *in vitro* in this search.

Materials and Methods:-

1-Design of study

This study conducted at AL-Hussein Education Hospital and The Special Center of The Endocrine Glands and Diabetes in Nassriyah from January 25,2011 to July 15,2011.

There were one hundred-fifty (150) subjects, patient with Diabetes Mellitus (DM), aged (18-60) years were included in this study and classified into ten groups according to type of disease, smoking and hypertension as illustrated in the following tables below:-

Table (1):- Data of type of disease groups

| Group | Name of group | N | Age (Y) |
|-------|--------------------------|----|---------|
| T1DM | Type 1 diabetes mellitus | 50 | 18-60 |
| T2DM | Type 2 diabetes mellitus | 50 | 18-60 |

N: number of subjects

Y: years

Table (2):- Data of smoking groups

| Group | Name of group | N | Age (Y) |
|-------|-------------------------------------|----|---------|
| T1DMS | Type 1 diabetes mellitus smokers | 50 | 18-60 |
| T1DMO | Type 1 diabetes mellitus nonsmokers | 50 | 18-60 |
| T2DMS | Type 2 diabetes mellitus smokers | 50 | 18-60 |
| T2DMO | Type 2 diabetes mellitus nonsmokers | 50 | 18-60 |

T1DMO = T1DM = Type 1 diabetes mellitus

T2DMO = T2DM = Type 2 diabetes mellitus

Table (3):- Data of hypertension groups

| Group | Name of group | N | Age (Y) |
|-------|---|----|---------|
| | | | |
| T1DMP | Type 1 diabetic-hypertension patients | 50 | 18-60 |
| T1DMR | Type 1 diabetic-non hypertension patients | 50 | 18-60 |
| T2DMP | Type 2 diabetic-hypertension patients | 50 | 18-60 |
| T2DMR | Type 2 diabetic-non hypertension patients | 50 | 18-60 |

T1DMR = T1DM = Type 1 diabetes mellitus

T2DMR = T2DM = Type 2 diabetes mellitus

2- Sample Collection

One hundred-fifty subjects with fasting blood sugar (11±0.22 mmol/L) for type 1 diabetes mellitus and (16±1.7 mmol/L) for type 2 diabetes mellitus for all patients subjects. About (4 mL) of blood were withdrawn by venipuncture from each subjects and transferred into disposable tube and prepared to the next treatments.

3- Blood Treatment with Sulphate copper, tannic and Gallic acids:-

Each (2.5 mL) of blood samples were treated with several solutions which can be illustrated below:-

| | Blood | Buffer | CuSO4 | Tannic acid | Gallic acid |
|-------------|--------|--------|---------|-------------|-------------|
| Test Tube 1 | 0.5 mL | 0.5 mL | | | |
| Test Tube 2 | 0.5 mL | 0.5 mL | 0.01 mL | | |
| Test Tube 3 | 0.5 mL | 0.5 mL | 0.01 mL | 0.01 mL | |
| Test Tube 4 | 0.5 mL | 0.5 mL | 0.01 mL | | 0.01 mL |
| Test Tube 5 | 0.5 mL | 0.5 mL | 0.01 mL | 0.01 mL | 0.01 mL |

Buffer = phosphate buffer pH= 7.5, (Randox Laboratories, England)

 $CuSO_4 = (1 \times 10^{-4} \text{ M}, BDH, England)$

Tannic acid = $(1 \times 10^{-4} \text{ M}, BDH, England)$

Gallic acid = $(1 \times 10^{-4} \text{ M}, BDH, England)$

The treated samples then were incubated at 37 C for two days (48 hour). Serum malondialdehyde was measured after centrifugation of samples.

4- Determination of serum malondialdehyde (MDA):-

Determination of serum malondialdehyde level which consider as a lipid peroxidation marker were preformed according to the method of Fong *et al.* 1973⁽²³⁾. In this method MDA reacts with thiobarbituric acid (TBA). In shaking water bath for 90 min at 60° C to developed a colored complex MDA(TBA)₂ which measured at 532 nm after cooling and centrifugation for 10 min at 600 g.

5- Statistical Analysis

Statistical analysis was carried out by One way ANOVA-test was used to compare parameters in different groups. P-values ($p \le 0.05$) were considered statistically significant. The results were expressed as mean \pm standard deviations (mean \pm SD) by using SPSS version 10.0.

Results and Discussion:-

Free radicals and reactive oxygen attack the normal cells to damage them and lead to formed in the body⁽²⁴⁾. Lipid peroxidation is one of several processes which is initiated by free radicals activities on lipids of cell membranes. Polyphenolic antioxidants are potent free radicals and hence, break the reaction of lipid peroxidation at the initiation step⁽²⁵⁾, therefore, in this study malondialdehyde (MDA) (an end product of lipid peroxidation) to evaluate free radicals generation and catalyzed lipid peroxidation by used copper sulphate and estimate the role of phenolic compounds such as tannic and gallic acids as antioxidants.

According to type of diabetes mellitus table(4) shows a significant (p \leq 0.05) increase in serum MDA levels in (B+C) T1DM in comparison with (B, B+C+T, B+C+G, B+C+T+G) T1DM and a significant increase (P \leq 0.05) increase in serum MDA levels in (B+C)T2DM in comparison with (B, B+C+T, B+C+G, B+C+T+G)T2DM but a significant decrease in serum MDA concentrations in (B, B+C, B+C+T, B+C+G, B+C+T+G) T1DM in comparison with (B, B+C, B+C+T, B+C+G, B+C+T+G) T1DM

Table(4):- Serum MDA levels of (B, B+C, B+C+T, B+C+G, B+C+T+G) T1DM and T2DM

| GROUP | N | | ROUP N | N | MDA concentra | tion (nmol/L)* |
|---------|------|------|-------------------------------|---------------------------|---------------|----------------|
| | | | mean | ± SD | | |
| | T1DM | T2DM | T ₁ DM | T2DM | | |
| В | 10 | 10 | 64.61 ± 0.63 ^a | 66.78 ± 0.22^{a} | | |
| B+C | 10 | 10 | 84.74± 0.10 ^b | 89.14± 0.56 ^b | | |
| B+C+T | 10 | 10 | 65.22 ± 0.27^{c} | 68.33 ± 0.13^{c} | | |
| B+C+G | 10 | 10 | $68.35 \pm 0.08^{\mathrm{d}}$ | 70.63 ± 0.23^{d} | | |
| B+C+T+G | 10 | 10 | $64.92 \pm 0.59^{\mathrm{e}}$ | 66.99 ± 0.06 ^e | | |

^{*} Each value represents mean \pm SD values with non identical superscript (a, b or c...etc.) were considered significantly differences (P \leq 0.05).

T1DM= Type 1 Diabetes Mellitus

T2DM= Type2 Diabetes Mellitus

B=buffer

B+C=buffer+ CuSO4

B+C+T=buffer+ CuSO4+tannic

B+C+G=buffer+ CuSO4+gallic

B+C+T+G=buffer+ CuSO4+tannic+gallic

Araya M, et al (2002)⁽²⁶⁾found that copper ions act as promoter of lipid peroxidation this finding matched with our results. It can be shown that the concentrations of serum MDA has been decreased in case using tannic and gallic acids alone or together. Many reporter⁽²⁷⁾⁽²⁸⁾⁽²⁹⁾ provide that tannic and gallic acids are a free radical scavenger against toxic effects of active oxygen

which decrease lipid peroxidation and MDA concentrations which similar to our finding.

Tables (5) and (6) ,respectively, show the studied group according to smoking. Table(5) showed a significant increase ($p \le 0.05$) in serum MDA in group (B+C)T1DMS in comparison with (B, B+C+T, B+C+G, B+C+T+G) T1DMS and a significant increase ($p \le 0.05$) increase in serum MDA levels in (B+C) T1DMO in comparison with (B, B+C+T, B+C+G, B+C+T+G)T1DMO. Also a significant decrease($p \le 0.05$)(B,B+C,B+C+T,B+C+G,B+C+T+G)T1DMO in comparison with (B, B+C, B+C+T, B+C+G, B+C+T+G) T1DMS.

Table(5):- Serum MDA levels of (B, B+C, B+C+T, B+C+G, B+C+T+G) T1DMS and T1DMO

| GROUP | N | | MDA concentra | tion (nmol/L)* |
|---------|-------|-------|--------------------------|-------------------------------|
| | | | mean | ± SD |
| | T1DMS | T1DMO | T1DMS | T1DMO |
| В | 10 | 10 | 79.32 ± 0.11^{a} | 64.61 ± 0.63 ^a |
| B+C | 10 | 10 | 97.67± 0.31 ^b | 84.74± 0.10 ^b |
| B+C+T | 10 | 10 | 83.18 ± 0.58^{c} | 65.22 ± 0.27^{c} |
| B+C+G | 10 | 10 | 87.39 ± 0.29^{d} | $68.35 \pm 0.08^{\mathrm{d}}$ |
| B+C+T+G | 10 | 10 | 80.02 ± 0.43^{e} | 64.92 ± 0.59^{e} |

^{*} Each value represents mean \pm SD values with non identical superscript (a, b or c...etc.) were considered significantly differences (P \leq 0.05).

T1DMS= Type 1 Diabetes Mellitus smokers

T1DMO= Type 1 Diabetes Mellitus non smokers= T1DM

B=buffer

B+C=buffer+ CuSO4

B+C+T=buffer+ CuSO4+tannic

B+C+G=buffer+ CuSO4+gallic

B+C+T+G=buffer+ CuSO4+tannic+gallic

Table(6):- Serum MDA levels of (B, B+C, B+C+T, B+C+G, B+C+T+G) T2DMS and T2DMO

| GROUP | N | | JP N MDA concentrat | tion (nmol/L)* | |
|---------|-------|-------|---------------------------|--------------------------|--|
| | | | mean | ± SD | |
| | T2DMS | T2DMO | T2DMS | T2DMO | |
| В | 10 | 10 | 89.26 ± 0.48^a | 66.78 ± 0.22^{a} | |
| B+C | 10 | 10 | 112.50± 0.19 ^b | 89.14± 0.56 ^b | |
| B+C+T | 10 | 10 | 97.74 ± 0.65^{c} | 68.33 ± 0.13^{c} | |
| B+C+G | 10 | 10 | 95.19 ± 0.12^{d} | 70.63 ± 0.23^{d} | |
| B+C+T+G | 10 | 10 | 90.35 ± 0.73 ^e | 66.99 ± 0.06^{e} | |

^{*} Each value represents mean \pm SD values with non identical superscript (a, b or c...etc.) were considered significantly differences (P \leq 0.05).

T2DMS= Type 2 Diabetes Mellitus smokers

T2DMO= Type 2 Diabetes Mellitus non smokers =T2DM

B=buffer

B+C=buffer+ CuSO4

B+C+T=buffer+ CuSO4+tannic

B+C+G=buffer+ CuSO4+gallic

B+C+T+G=buffer+ CuSO4+tannic+gallic

While table (6) show a significant (p \leq 0.05) decrease in the concentration of serum MDA in groups (B, B+C+T, B+C+G, B+C+T+G) T2DMS in comparison with (B+C)T2DMS and a significant increase (p \leq 0.05) in serum MDA levels in (B+C) T2DMO in comparison with (B, B+C+T, B+C+G, B+C+T+G)T2DMO.

Also a significant decrease $(p \le 0.05)(B,B+C,B+C+T,B+C+G,B+C+T+G)$ T2DMO in comparison with (B,B+C,B+C+T,B+C+G,B+C+T+G) T2DMS.

It has been reported⁽³⁰⁾⁽³¹⁾⁽³²⁾ that cigarette smoking consider a major risk factor for development of hyperglycemia which causes free radical production and oxidative stress which result increase lipid peroxidation.

Yagi.k ,et al (2002)⁽³³⁾ found that increase lipid peroxidation in smokers supports of hypothesis that smoking increase free radical-mediated oxidative damage of lipid, where this result is similar to our results.

Our previous analysis showed the smoking-diabetic patients have the highest concentration of MDA. Cigarette smoking has already copper ions⁽³⁴⁾ and the incubation of samples with addition of copper ions (copper sulphate) as a promoter of lipid peroxidation which result this increase in MDA levels⁽³⁵⁾.

In this research we focused on the antioxidant activity of tannic and gallic acids these acids showed a strong and a substantial activity as an antioxidants by their donation of electron to reactive oxygen species (ROS) which initiated lipid peroxidation and this active belong to tannic and gallic acids may be potentially useful for their structures⁽³⁶⁾.

According to hypertension- diabetic groups table(7) shows a significant (p \leq 0.05) increase in serum MDA levels in (B+C) T1DMH in comparison with (B, B+C+T, B+C+G, B+C+T+G) T1DMH and a significant increase (P \leq 0.05) increase in serum MDA levels in (B+C)T1DMR in comparison with (B, B+C+T, B+C+G, B+C+T+G)T1DMR but a significant decrease in serum MDA concentrations in (B, B+C, B+C+T, B+C+G, B+C+T+G)T1DMR in comparison with (B, B+C, B+C+T, B+C+G, B+C+T+G) T1DMH.

Table(7):- Serum MDA levels of (B, B+C, B+C+T, B+C+G, B+C+T+G)
T1DMH and T1DMR

| GROUP | N | | MDA concentra | tion (nmol/L)* |
|---------|-------|-------|--------------------------|-------------------------------|
| | | | mean | ± SD |
| | T1DMH | T1DMR | T ₁ DMH | T1DMR |
| В | 10 | 10 | 71.15 ± 0.95^{a} | 64.61 ± 0.63 ^a |
| B+C | 10 | 10 | 92.33± 0.21 ^b | 84.74± 0.10 ^b |
| B+C+T | 10 | 10 | 77.24 ± 0.62° | 65.22 ± 0.27^{c} |
| B+C+G | 10 | 10 | 80.21 ± 0.81^{d} | $68.35 \pm 0.08^{\mathrm{d}}$ |
| B+C+T+G | 10 | 10 | $74.64 \pm 0.56s^{e}$ | 64.92 ± 0.59^{e} |

^{*} Each value represents mean \pm SD values with non identical superscript (a, b or c ...etc.) were considered significantly differences (P \leq 0.05).

T1DMH= Type 1 Diabetes Mellitus with hypertension

T1DMR= Type 1 Diabetes Mellitus non hypertension

B=buffer

B+C=buffer+ CuSO4

B+C+T=buffer+ CuSO4+tannic

B+C+G=buffer+ CuSO4+gallic

B+C+T+G=buffer+ CuSO4+tannic+gallic

Also a significant (p \leq 0.05) decrease in the concentration of serum MDA in groups (B, B+C+T, B+C+G, B+C+T+G) T2DMH in comparison with (B+C)T2DMH and a significant increase (p \leq 0.05) in serum MDA levels in (B+C) T2DMR in comparison with (B, B+C+T, B+C+G, B+C+T+G)T2DMR

and a significant increase (p \leq 0.05) (B, B+C, B+C+T, B+C+G, B+C+T+G) T2DMH in comparison with (B, B+C, B+C+T, B+C+G, B+C+T+G) T2DMR. As illustrated in table (8).

Table(7):- Serum MDA levels of (B, B+C, B+C+T, B+C+G, B+C+T+G)
T2DMH and T2DMR

| GROUP | N | | MDA concentra | tion (nmol/L)* |
|---------|-------|-------|---------------------------|-------------------------------|
| | | | mean | ± SD |
| | T2DMH | T2DMR | T2DMH | T2DMR |
| В | 10 | 10 | 79.51 ± 0.32^{a} | 66.78 ± 0.22^{a} |
| B+C | 10 | 10 | 108.47± 0.49 ^b | 89.14± 0.56 ^b |
| B+C+T | 10 | 10 | 89.88 ± 0.67^{c} | 68.33 ± 0.13^{c} |
| B+C+G | 10 | 10 | 90.14 ± 0.54 ^d | $70.63 \pm 0.23^{ m d}$ |
| B+C+T+G | 10 | 10 | $85.20 \pm 0.24^{\rm e}$ | $66.99 \pm 0.06^{\mathrm{e}}$ |

^{*} Each value represents mean \pm SD values with non identical superscript (a, b or c...etc.) were considered significantly differences ($P \le 0.05$).

T2DMH= Type 2 Diabetes Mellitus with hypertension

T2DMR= Type 2 Diabetes Mellitus non hypertension=T2DM

B=buffer

B+C=buffer+ CuSO4

B+C+T=buffer+ CuSO4+tannic

B+C+G=buffer+ CuSO4+gallic

B+C+T+G=buffer+ CuSO4+tannic+gallic

A preponderance of studies in the past indicates that hypertension induced by oxidative stress which caused by reactive oxygen species and consequently lipid peroxidation increase, also many researches⁽³⁷⁾⁽³⁸⁾⁽³⁹⁾ proved that hypertension seems to be linked to increase some trace elements such as copper, in other words, hypertension induced the increase of lipid peroxidation and copper ions which can be promoted *in vitro* by incubation with copper ions (copper sulphate).

Observable, the results in tables (6)(7) that tannic and gallic acids have a fundamental role as antioxidant and they are capable of minimizing lipid peroxidation marker MDA⁽⁴⁰⁾. Furthermore, the high inhibition of these two acids for copper mediated lipid peroxidation due to their structure which may be the essential factor for their antioxidant property⁽⁴¹⁾⁽⁴²⁾⁽⁴³⁾.

References:-

- 1- Baynes, JW;" Role of oxidative stress and chromium in development of complications in diabetes"; *Diabetes*.**40**: 406–412; 2001.
- 2- Stadler, N., Lindner, RA., Davi,es MJ;"Direct detection and quantification of transition metal ions in human atherosclerotic plaques: evidence for the presence of elevated levels of iron and copper"; *Arterioscler Thromb Vasc Biol.* **24**: 949–954;2004.
- 3- Fernandez-Real, JM., Lopez-Bermejo, A., Ricar, W;"Cross-talk between iron metabolism and diabetes"; *Diabetes* **51**:2348–2354; 2008.
- 4- Giugliano, D., Ceriello, A., Paolisso, G.; "Diabetes mellitus, smoking, and cardiovascular disease: which role for oxidative stress"; Metabolism. **44**:363-8;2005.
- 5- Wolfe, J. T., Ross, D. & Cohen, G. M.;" A role for metals and free radicals in the induction of apoptosis in thymocytes"; FEBS Lett. **352**:58-62;2004.

- 6-Bjelakovic G, et al. "Mortality in randomized trials of antioxidant supplements for primary and secondary prevention: systematic review and meta-analysis". JAMA **297** (8): 42–57; 2007.
- 7-Vertuani S, Angusti A, Manfredini S ."The antioxidants and proantioxidants network: an overview". Curr Pharm Des**10** (14): 1677–94; 2004.
- 8-Imlay J. "Pathways of oxidative damage". Annu Rev Microbiol **57**: 395–418;2003.
- 9-Halliwell, B. "Are polyphenols antioxidants or pro-oxidants? What do we learn from cell culture and in vivo studies?". *Archives of Biochemistry and Biophysics* **476** (2): 107–112;2008.
- 10- Ji LL. Exercise and oxidative stress: Role of the cellular antioxidant systems. In: Holloszy JO, Ed. Exercise Sport Science Reviews. Baltimore, MD: Williams & Wilkins, pp135–166; 2005.
- 11-Warner D, Sheng H, Batinić-Haberle I. "Oxidants, antioxidants and the ischemic brain". *J Exp Biol* **207** (Pt 18): 3221–31; 2004.
- 12- Sorensen, A.-D. M.; Haahr, A.-M.; Becker, E. M.; Skibsted, L. H.; Bergenståhl, B.; Nilsson, L.; Jacobsen, C. "Interactions between iron, phenolic compounds, emulsifiers, and pH in omega-3-enriched oil-in-water emulsions" *J. Agric. Food Chem.* **56**:1740-1750;2008.
- 13-Thoss, V., Baird, M. S., Lock, M. A. & Courty, P. V. "Quantifying the phenolic content of freshwaters using simple assays with different underlying reaction mechanisms". *J. Environ. Monit.* **4**:270-275; 2002.
- 14- McGee, Harold. "On Food and Cooking. Simon & Schuster". New York, NY. pg 714: 2004.
- 15- S. M. Fiuza. "Phenolic acid derivatives with potential anticancer properties—a structure—activity relationship study. Part 1: Methyl, propyl and octyl esters of caffeic and gallic acids". Elsevier;2004.

- 16- Riedl, K. M. & Hagerman, A. E. Tannin-protein complexes as radical scavengers and radical sinks. *J. Agric. Food Chem.* **49**:4917-4923;2001.
- 17- R. Puupponen-Pimiä, L. Nohynek, C. Meier, M. Kähkönen, M. Heinonen, A. Hopia & K.-M. Oksman-Caldentey, "Antimicrobial properties of phenolic compounds from berries" *Journal of Applied Microbiology* 90(4) pp494;2001.
- 18-Okuda, T., Yoshida, T. & Hatano, T." Hydrolysable tannins and related polyphenols". *Fortschr. Chem. Org. Naturst.* 66:1-117;2005.
- 19- Ling-Ling Yang, Chih-Ying Lee, Kun-Ying Yen "Induction of apoptosis by hydrolyzable tannins from Eugenia jambos L. on human leukemia cells" Cancer Letters 157;2000.
- 20- Lu Y., Foo LY. "Antioxidant and radical scavenging activities of polyphenols from apple pomace". *Food Chem*; **68**: 81-85;2000.
- 21- Crozier A., Burns J., Aziz A.A., Stewart A.J., Rabiasz H.S., Jenkins G.I., Edwards C.A., Lean MEJ. "Antioxidant flavonols from fruits, vegetables and beverages: measurements and bioavailability". Biol Res 33: 79-88;2000.
- 22- Tapiero H., Tew K.D., Ba G.N., Mathe G. "Polyphenols: do they play a role in the prevention of human pathologies?" *Biomed Pharmacotherapy*. **56** 2-7 ;2002.
- 23- Fong, K.L., McCay, P.B., and Poyer, J.L.; J. Biol. Chem., 248: 7792; 1973.
- 24- Harman, D. "Role of free radicals in aging and disease"; Annals of New York Academy of Sciences. **673**:126-141;2002.
- 25- West, I.C. "Radicals and oxidative stress in diabetes" *Diabetic Med* .17:171–180;2000.
- 26- Araya M, González M, Olivares M, Uauy U. "Biological effects of chronic copper exposure. In: MASSARO E (ed) Copper Pharmacology and Toxicology". Totowa, New Jersey: Humana; 2002.

- 27- Barbosa, D. S. "Green tea polyphenolic compounds and human health" *J. Verbraucherschutz Lebensmittelsicherheit* . 7: 407-413; 2007.
- 28-Jittawan, K.; Sirithon, S. "Phenolic contents and antioxidant activities of bitter gourd (Momordica charantia L.) leaf, stem and fruit fraction extracts in vitro" *Food Chem.* **110:** 881-890; 2008.
- 29-Strlic, M.; Radovic, T.; Kolar, J.; Pihlar, B. "Anti- and prooxidative properties of gallic acid in Fenton-type systems" *J. Agric. Food Chem.* **50**: 6313-6317; 2002.
- 30- Trummer H, Habermann H, Haas J, Pummer K. "The impact of cigarette smoking on human semen parameters and hormones". 17:1554–1559;2002.
- 31- Sohal R. "Role of oxidative stress and protein oxidation in the aging process". *Free Radic Biol Med* **33** (1): 37–44;2002.
- 32- David E. Laaksonen, Leo Niskanen, Kari Punnonen, Kristiina Nyyssönen.

 " The Metabolic Syndrome and Smoking in Relation to Hypogonadism in Middle-Aged Men: A Prospective Cohort Study". 90: 712-719: 2005.
- 33-Yagi, K.;"Assay for serum lipid peroxide level and its clinical significance";In: Yagi K, ed;"Lipid Peroxides in Biology and Medicine";New York, NY: *Academic Press*; 2002.
- 34- Chow, CK.; "Cigarette smoking and oxidative damage in the lung"; Ann N Y Acad Sci. **686**:289-298;2003
- 35- Giugliano, D., Ceriello, A., Paolisso, G.; "Diabetes mellitus, smoking, and cardiovascular disease: which role for oxidative stress"; Metabolism. **44**:363-8;2005.
- 36- Fazary, A. E.; Ju, Y. H. "Non aqueous solution studies on the protonation equilibria of some phenolic acids" *J. Solution Chem.* **37**:1305 -1319;2008.
- 37- Kruszynska YT, Yu JG, Olefsky JM, Sobel BE. "Effects of troglitazone on blood concentrations of plasminogen activator inhibitor 1 in patients with diabetes and in lean and obese normal subjects". *Diabetes* . **49**: 633–639;2000.

- 38- Mogensen CE. "Microalbuminuria, blood pressure and diabetic renal disease: origin and development of ideas". **42**: 263–285;2009.
- 39- Haffner FM, Greenberg AS, Weston WM, Chen H, Williams K, Freed MI. "Effect of rosiglitazone treatment on non traditional markers of cardiovascular disease in patients with diabetes mellitus. Circulation" . **106**: 679–684;2002.
 - 40- Demmig-Adams B, Adams W. "Antioxidants in photosynthesis and human nutrition". *Science (journal)***298** (5601): 2149–53;2004.
 - 41- Olivares M, Pizarro F, Depablo S, Araya M, Uauy R. "Iron, zinc and copper: Contents in common Chilean foods and daily intakes in Santiago City, Chile". **20**: 205-212; 2004.
 - 42-Abdullakasim P, Songchitsomboon S, Techagumpuch M, Balee N, Swatsitang P, Sungpuag P. "Antioxidant capacity, total phenolics and sugar content of selected Thai health beverages" .*Int J Food Sci Nutr*. Feb;**58**(1):77-85;2007.
 - 43-Sabu MC, Smitha K, Kutan R. "Anti-diabetic activity of green tea polyphenols and their role in reducing oxidative stress in experimental diabetes". *J Etnopharmacol* **83**:109–116; 2002.

دراسة مختبريه لحامضي التانيك والكاليك كمضادات أكسدة في داء السكري

هتاف بازول فرهود قسم الكيمياء - كلية العلوم - جامعة ذي قار الناصرية - العراق

الخلاصة

مضادات الأكسدة هي جزيئات قادرة على إبطاء أو منع أكسدة جزيئه أخرى من خلال منحها لإلكترونها إلى الجذور الحرة والتي تبدأ بسلسلة من التفاعلات التي تحطم الخلايا.حاولنا في هذه الدراسة تقييم الفعالية المضادة للأكسدة لحامضي التانيك والكاليك. استخلصت هذه الحوامض من الشاي خاصة الشاي الاخضر ،الرمان ، ثمر البرسيمون ،التوت على الاغلب التوت البري ،الليمون، البقوليات والكاكاو. تضمنت هذه الدراسة 150 شخصاً قسموا إلى ثلاث مجاميع بالاعتماد على نوع داء السكري ، التدخين و ارتفاع ضغط الدم .حيث تم تعزيز الأكسدة الفوقية للدهون بحضن مصل المرضى مع ايونات النحاس (كبريتات النحاس 1×10^{-4} مولاري) لمدة عند 37 م . أظهرت نتائج حضن المصل مع أيونات النحاس بأن ايونات النحاس سوف يزيد من الأكسدة الفوقية للدهون (MDA) عند استخدامهما لوحدهما أو كليهما وبتركيز (1×10^{-4} مولاري) لكل منهما .

الكلمات المفتاحية :- داء السكرى، حامض التانيك، حامض الكاليك، التدخين، ارتفاع ضغط الدم، الأكسدة الفوقية للدهون