

Protective Effect of Alcoholic extract of Black Current in Male Reproductive System of Methionine Overload Rats

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Summary

This study was undertaken to investigate the deleterious- effect of methionine overload on male reproductive system of adult male rats. It was also aim to studying the beneficial protective effect of alcoholic extract of black current. Eighteen adult male rats (175-200 gm) were used, animals were randomly divided into three groups (6/group) and were treated daily for 42 days as follows: 1-Group C (control); 2- group I(GI), the rats in this group were orally intubated by gavages needle DL-methionine (100 mg / Kg BW.) 3- animals in group II(GII) were administered orally alcoholic extract of black current (60 mg /Kg BW.) in addition to methionine. At the end of the experiment blood samples were collected for measuring serum testosterone (T) hormone and follicular stimulating hormone (FSH) concentrations. As well as histological section, for testis was taken for histopathological study the results revealed that oral intubation of 100mg /kg BW. of methionine for 42 days caused significant decrease - in serum testosterone (T) hormone and follicular stimulating hormone (FSH) concentrations in groups (GI and GII) comparing to control ,while oral intubation of alcoholic- extract of black current (GII) caused significant elevation in the concentration of these two hormones comparing to methionine treated group (GI). Furthermore, methionine intubation caused significant decrease in diameter and thickness of seminiferous tubules with cellular degeneration of Sertoli cells, while black current intubation caused significant increase in the previous criteria and regeneration of Sertoli cells. In conclusion, this study pointed to the deleterious effect of methionine overload and assures protective effect of alcoholic extract of black current on adult male rats reproductive system.

Keywords: Methionine, testosterone, FSH, male reproduction, black current.

التأثير الوقائي للمستخلص الكحولي للزبيب الأسود في الجهاز التناسلي الذكري للجرذان المعاملة بفرط الميثيونين

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الخلاصة

صممت هذه الدراسة لمعرفة التأثير الضار لفرط الميثيونين على الجهاز التناسلي الذكري للجرذان، بالإضافة الى معرفة التأثير الوقائي للمستخلص الكحولي للزبيب الأسود. تم استخدام ثمانية عشر من ذكور الجرذان البالغة وقسمت عشوائياً الى ثلاثة مجاميع متساوية وعولمت كالاتي لمدة 42 يوم 1-مجموعة السيطرة (C) جرعت محلول الداروىء 2؛- مجموعة المعاملة الأولى (GI) جرعت الجرذان في هذه المجموعة 100 ملغم /كغم من وزن الجسم ميثيونين بأستخدام اللي المعدي 3- جرعت حيوانات المجموعة الثانية (GII) 60ملغم /كغم من وزن الجسم بالمستخلص الكحولي بالإضافة الى 150 ملغم/كغم من الجسم من الميثيونين . جمعت عينات الدم في نهاية التجربة لقياس تركيز هرموني التستوستيرون وهرمون محفز الجريب في مصل الدم، فضلاً عن أخذ مقاطع نسيجية من الخصية لدراسة التغيرات النسيجية المرضية. أظهرت النتائج حدوث انخفاض معنوي في تركيز هرموني التستوستيرون و محفز الجريب في المجاميع المعاملة بفرط الميثيونين (GI و GII) مقارنة مع مجموعة السيطرة، في حين أدى التجريع الفموي للمستخلص الكحولي للزبيب الأسود (GII الى حصول ارتفاع معنوي في هذين الهرمونين مقارنة مع مجموعة (GI) . فضلاً عن ذلك فقد تسبب فرط الميثيونين الى حصول انخفاض معنوي في سمك وقطر النبيبات المنوية مع حدوث تنكسات خلوية في خلايا سرتولي مقارنة مع المجموعة المجرعة بالمستخلص الكحولي للزبيب الأسود الذي تسبب في حدوث ارتفاع معنوي في سمك وقطر النبيبات المنوية مع انكفاء التنكس الحاصل في خلايا سرتولي. نستنتج من هذه الدراسة وجود التأثير الضار لفرط الميثيونين والذي يمكن الوقاية منه بأستخدام المستخلص الكحولي للزبيب الأسود في الجرذان البالغة.

الكلمات المفتاحية: الزبيب الاسود، الجرذان، فرط الميثيونين، الجهاز التناسلي الذكري.

Introduction

Methionine is an essential for maintaining proper growth and development in mammals and domestic animals, such as pigs and chicken ,contributes to better production efficacy(1). Lynch and Strain, (2) found that excess L-methionine intake induce the production of oxidative metabolites and cause defect in the enzymatic antioxidant defense system due to induction of homocysteine (Hcy). Hcy is an endogenous sulfur-containing amino acid found during the metabolism of methionine and is not obtained from diet (3).

Hyperhomocysteinemia (HHcy), is a state of elevated plasma homocysteine level (4). Methionine is believed to exert several toxic effects, it has been implicated in retrospective and prospective studies, as a risk factor for atherosclerosis which includes venous thrombosis (5, 6 and 7), coronary artery disease (8), myocardial infarction, stroke (9 and10), primary hypertension (11), rheumatoid arthritis and osteoporosis (12) as well as hypertrophy and fatty change in the liver (13). HHcy exerts its toxicity by increasing H₂O₂ production and promoting lipid peroxidation (LPO) (14 and 15). Besides, HHcy induced endothelial dysfunction through the attenuating nitric oxide (NO) bioavailability and injury involves oxidative damage (16 and17).

Reactive oxygen species (ROS) including oxygen ions, free radicals and peroxides are generated by sperm and seminal leukocytes within sperm produce infertility (18 and19) by damaging the sperm membrane or by alter sperm DNA (20).Today, grape seed extract use to treatment of a range of health problem related to free radicals damage including aging, DNA damage, cancer and heart disease (21). Shi *et al.*, (22) pointed that polyphenol in different grape species potentially inhibited reactive oxygen species .Also procyanidins (PCOS) found in grape juice are highly antioxidant effects than vitamin E and C(23) . Recently, it has been shown that natural honey (24) and *Nigella sativa* (25) caused almost protection against methionine - induced hyper homocysteinemia in rats.

Current interest is focused upon modulating the methionine over load through natural preventive strategies. The study aims to investigate the effect of induced methionine over load on some testicular functions in adult male rats and treated by using black current.

Materials and Methods

Seventy percent of alcoholic extract of black current *Vitis Venifera L.* belong to family Vitaceae was prepared according to the procedure described by Harbom (26). Eighteen adult male rats weighed (175-200gm) were randomly divided into three groups (6/group) and were treated daily for 42 days as follows: Group C; rats in this group were administered daily buffer solution 0.1M, PH 7 by oral intubation using gavage needle and served as control group; group I (GI) the rats in this group were orally by gastric gavage tube DL.methionine (100 mg / Kg BW.) (27). Diluting in buffer 0.1M, PH7 and animals in groupII (GII) were administered orally alcoholic extract of black current (60mg /Kg BW.) (28) in addition to methionine as described in GI, using gastric gavage tube .

At the end of the experiment blood samples were drawn via cardiac puncture technique and serum was collected by centrifugation and frozen at -20 until analysis. Testosterone (T) hormone and follicular stimulating hormone (FSH) concentrations was measured in serum by radioimmunoassay (RIA) kits (Beckman-France).The testis was excised and a portion of it was immersion fixed in 10% neutral buffered formalin .Sequential five sections of testis were prepared and stained with hematoxylin and eosin (H&E) to evaluate the histological changes of seminiferous tubules (29). The results were analyzed using one-way analysis of variance (ANOVA) .The level of statistic was set at P<0.05(30).

Results and Dissuasion

Table (1) the mean value of testosterone concentration and FSH in control and treated groups .The results showed a general tend for testosterone values to decrease in two treated

groups (GI and GII) as compared to control group .This decrement reached statistical significance ($P<0.05$) after 42days of experiment where the mean values of serum testosterone concentration in GI and Gil after 42days of experiment are 2.68 ± 0.05 and 2.91 ± 0.06 respectively, as compared to control. From other side oral intubation of alcoholic extract of black current seeds (GII) caused significant elevation of this hormone comparing to methionine treated group (GI). A significant decrease ($P<0.05$) in serum FSH concentration in methionine treated group (GI) were observed at the end of the experiment comparing to control group .While, oral intubation of alcoholic extract of black current with methionine (GII) superesed the decrease of FSH concentration ($P>0.05$) compared with GI group.

The effect of oral intubation of alcoholic extract of black current and methionine to adult male rats on mean value of diameter of seminiferous tubules were clarified in table (2). A significant decrease ($P<0.05$) in the mean value of this parameter in group GI (192.16 ± 2.50) was observed as compared to control (210.63 ± 1.67).Moreover, the value of diameter of seminiferous tubules tend to increase significantly ($P<0.05$) in group Gil (204.78 ± 2.96) comparing to GI and the value tend to normalize that of the control . Depending on the results clarified in table (2), there was significant decrease ($P<0.05$) in thickness of seminiferous tubules in GI(72.0 ± 1.47) and GII(80.20 ± 1.59)comparing to control group (86.51 ± 2.05) while oral intubation of alcoholic extract of black current caused significant elevation in thickness of seminiferous tubules comparing to methionine treated group.

Histological section of rat testis treated with (100mg/kg BW.) of methionine for 42 days showed acute cellular degeneration in the Sertoli cells with low spermatogenesis and presence of protenious materials in the lumen of seminiferous tubules (figure-1) comparing to normal section of testis in control group (figure-2). Oral intubation of 60mg/kg BW. of alcoholic extract of black current concurrently with 100mg/kg BW. Methionine caused regression of pathological lesions and black current preserved the histological structure of testis with regeneration of testicular tissue (figure-3).

Table (1): Effect of alcoholic extract of *Black Current* and methionine overload on serum testosterone (T), follicular stimulating hormone (F.S.H) in adult male rats.

Group	(C) Control	(GI) Methionine (100mg/kg. BW)	(GII) (Methionine 150mg/kg.BW +black Current 60mg/kg.BW)
Parameters			
Testosterone (ng/ml)	3.51 ± 0.06 A	2.68 ± 0.05 B	2.91 ± 0.06 C
F.S.H (I U/ml)	1.77 ± 0.03 A	1.52 ± 0.02 B	1.69 ± 0.03 A

Values are expressed as mean ± SE; n= 6 / group.

Different capital letters denote between groups differences, $P< 0.05$ Vs. control.

Table (2): Effect of alcoholic extract of *Black Current* and methionine overload on diameter and thickness of seminiferous tubules in adult male rats.

Group	(C) Control	(GI) Methionine (100mg/kg. BW)	(GII) (Methionine 150mg/kg.BW +black Current 60mg/kg.BW)
Parameters			
Diameter (µM)	210.63 ± 1.67 A	192.16 ± 2.50 B	204.78 ± 2.96 A
Thickness (µM)	86.51 ± 2.05 A	72.00 ± 1.47 B	80.20 ± 1.59 C

Values are expressed 1 as mean ± SE; n= 6 / group. Different capital letters denote between groups differences, $P< 0.05$ vs. control.

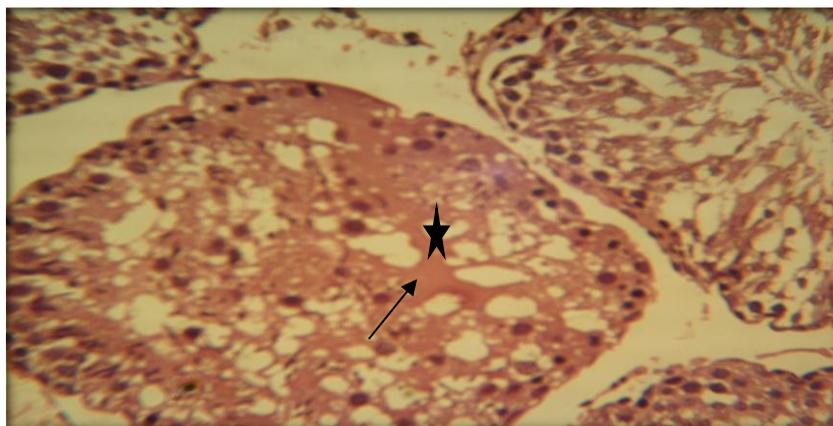


Figure (1): Histological section of rat testis treated with methionine overload (GI).Show degeneration in Sertoli cell (→) and proteinaceous material in lumen of seminiferous tubule (★) (H&E X40).

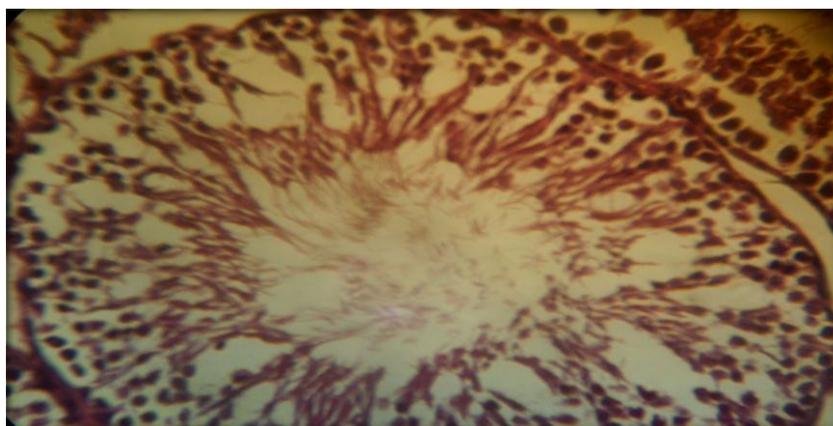


Figure (2): Histological section of testis of rat from control group .Show normal structure of seminiferous tubule (H&E X40).



Figure (3): Histological section of rat testis treated with methionine overload and alcoholic extract of black current (GII).Show regression of pathological lesion (→) (H&E X40).

Evidence now suggests that ROS — mediated oxidative damage to sperm is a significant contributing to pathology of 30-80% of male infertility (31, 32, 33 and 34). Inductions of

HHcy and oxidative stress after methionine overload have well documented (35, 36, 37 and 38). Accordingly, it can be hypothesized that oxidative stress and HHcy induced after methionine overload may be responsible for such detrimental damage of male rat reproductive system. The toxic accumulation of homocysteine may cause reproductive dysfunction and oxidative stress within the testis (39 and 40) ; abnormal spermatogenesis and male infertility (41).Furthermore, epididymal sperm concentration , percentage of progressive sperm motility and plasma testosterone concentration were significantly decreased in Hcy — treated rats (40), It has been well documented that inflammatory cytokines ,such as those induced by HHcy ,have been associated with impaired sperm parameters and male fertility(42).Furthermore, nitric oxide (NO) signaling are involved in penile erection ,spermatogenesis ,dynamic of blood tests barrier ,sperm motility , capacitation ,acrosomal reaction and fertilization (43 and 44) .Thus any alteration of NO bioavailability by HHcy may have direct consequences on male reproductive functions. The major methyl donor, SAM stimulated human chorionic gonadotropin (hCG) - mediated testosterone synthesis in purified rat leydig cells in vitro, whereas, SAH produced due to methionine overload (methylation defect) had opposite effect (45).

The suspected decrease in number of leydig cells, which is responsible for synthesis and secretion of testosterone after methionine overload necrosis and atrophy of Sertoli cells, may lead to decrease in testosterone concentration (46). Selective suppression of FSH in this experiment could result from action of methionine overload on induction of hepatic cytochrom P450 enzymes and expression of substantial level of non-steriodogenic P450 inciting local cell land tissues damage (47) leading to inactivation of cell responsible for secretion of hormone including FSH hormone. Overall, this scenario is consistent with histological picture of methionine overload group exhibited cellular degeneration of Sertoli cells and proteineous materials in lumen of seminiferous tubules.

The results also showed that the alcoholic extract of *Black Current* caused significant correction to male function through elevation of testosterone and FSH concentration in addition to increase thickness and diameter of seminiferous tubules. Several studies have reported that the levels of ROS within semen can be reduced by augmented the scavenging capacity of seminal plasma using oral antioxidant supplementation like a astaxanthine (48) ,combination of vitamins E and C (49) zinc and selenium (50) . Antioxidant supplementation like folate which possessed the capacity to increase the enzymatic efficacy of the Mther and cystathionin glutathione B-synthase enzyme responsible for removing of Hcy from circulation (51) has beneficial effect (antioxidant) on spermatogenesis and enhancement sperm quality (50).Accordingly, supplementation of grape seed extract may restore antioxidant activity of the body, repair the case of oxidative stress and improve seminiferous tubule function. The beneficial effect of grape seeds against toxic effect of methionine may be attributed to the its active compound like resveratrol and proanthocyanidin (52) .These compound are considered as antioxidant which remove the free radicals produced after methionine overload and restoring male reproductive function . Increase in FSH and testosterone concentration in animal received grape seeds extract, may increase testicular weight and volume concomitant with increasing diameter of seminiferous tubules (53 and 54). On the other hand, testosterone supports spermatogenesis and increase maturation; facilitating round to elongated spermatid and may lead to increase lumen and diameter of seminiferous tubules (55 and 56).

Vitamin E is one of the major membranes protective against ROS and lipid peroxidation and improved sperm viability in cattle under oxidative stress (57).As well as, vitamin E as antioxidant can prevent adverse effect of homocystein on testosterone level, epididymal sperm count and motility in male rats (40). Antioxidant compound of grape seeds extract may increase plasma vitamin E (58), which is play important role in decreasing the toxicity of methionine and returning the diameter of seminiferous tubules near to normal.

References

- 1-Toue, S.; Kodama, R.; Amao, M.; Kawamata, Y.; Kimura, T. and Sakai, R. (2006). Screening of toxicity biomarkers for methionine excess in rats. *J Nutr.*, 136: 1716-1721.
- 2-Lynch, SM. and Strain, JJ.(1989). Increased hepatic lipid peroxidation with methionine toxicity in the rats. *Free Radic Res.*, 5:221-226.
- 3-Ueland, PM. (1982). Pharmacological and biochemical aspect of S-adenosylhomocysteine a S-adenosylhomocysteine hydrolase. *Pharmacol. Rev.*, 34:223-253.
- 4- Bostom, AG. and Culleton, BF. (1999). Hyperhomocysteinemia in chronic renal disease. *J. Am. Soc. Nephrol.*, 10: 891- 900.
- 5-Den Heijer, M. and Keijzer, MB.(2001). Hyperhomocysteinemia as a risk factor for venous thrombosis. *Clin. Chem. Lab. Med.*, 39: 710-713.
- 6-Rowan, E.; Dickinson, H.; Stephens, S.; Ballard, C.; Kalaeia, R. and Anne, KR. (2007). Homocystein and post-stroke cognitive decline. *Aging.*, 36:339-343.
- 7-Hillenbrand, R.; Hillenbrand, A.; Liewald, F. and Zimmermann, J. (2008). Hyperhomocysteinemia and current carotid stenosis . *Cardiovascular Disorder*, 8:1-9.
- 8-Boushey, CL.; Beresford, SA.; Omenn, GS. and Motulsky, AG.(1995). A quantitative assessment of plasma homocysteine as a risk factor for vascular disease .Probable benefits of increasing folic acid intake. *JAMA.*, 274:1049-1057.
- 9-Lentz, SR. and Haynes, WG. (2004). Homocysteine: Is it a clinically important cardiovascular risk factor. *Clin. J. Med.*, 71(9):729-734.
- 10-Haim, M.; Tanne, D.; Goldbourt, U.; Doolman, R.; Boyko, V.; Brunner, D.; Sela, B.A. and Beha, S. (2007). Serum homocystein and long-term risk of myocardial infarction and sudden death in patients with coronary heart disease. *Cardi.*, 107:52-56.
- 11- Oveckin, A.; Tygi, N. ; Lominadze, D. ; Steed, M. ; Moshal, K. and Tygi, S. (2006). 3-Deazadenosine mitigates arterial remodeling and hypertension in hyperhomocysteinemic mice. *AM. J. Physiol. Lung Cell Mol. Physiol.*, 29(5): 1905-1911.
- 12-Sahi, A.; Pan, X.; Paul, R. ; Malladi, P.; Kahli, R. and Whitinton ,F. (2006). Roles of phosphatidylinositol 3-kinase and osteopontin in steatosis and aminotransferase release by hepatocytes treated with methionine - choline deficient medium. *Am. J. Physiol. Gastro. Intest. Physiol. Liver Physiol.*, 291:55-62.
- 13-Adinolfi, LB.; Ingrosso, D.; Cesaro, C. and Cimmino, A. (2005). Hyperhomocysteinemia and the MTHFR C6771 polymorphism promote steatosis and fibrosis in chronic hepatitis C patients. *Hepatology* ,41: 995-1003.
- 14-Faraci, F.M.(2003). Hyperhomocysteinemia : a million ways to lose control *Arterioscler Thromb Vase Biol.*, 23:371-373.
- 15-Loscaizo, J. (2003). Oxidant stress .a key determinant of atherothrombosis. *Biochem. Soc. Trans.*, 31:1059-1061.
- 16-Huang, RS.; Hsu, Y.; Lin, H. and Yang, FL. (2001). Folate depletion and elevated plasma homocysteine promote oxidative stress in rat liver. *J. Nutr.*, 13:33-38.
- 17-Ungvari, Z.; Csiszar, A.; Edwaed, JG.; Kaminski, PM; Wolin, MS. ;Kaley, G. and Koller, A. (2003). Increase superoxide production in coronary arteries in hyperhomocysteinemia : role of tumor necrosis factor —alpha ,NADPH oxidase and inducible nitric oxide synthase. *Arterioscler. Thromb. Vase. Biol.*, 23:418-424.
- 18-Shen, HM. and Ong, CN. (2000). Detection of oxidative DNA damage in human sperm and its association with sperm function and male infertility. *Free Rad Biol. Med.*, 28: 529-539.
- 19-Agarwal, A.; Saleh, RA. and Bedaiwy, MA. (2003). Role of reactive oxygen species in the pathophysiology of human reproduction. *Fertil. Steril.*, 79 (4): 829-843.
- 20-Xu, DX.; Shen, HM.; Zhu, QX.; Chua, L.; Wang, QN.; Chia, SE. and Ong, CN.(2003). The associations among semen quality, oxidative DNA damage in human spermatozoa and concentrations of cadmium ,lead and selenium in seminal plasma. *Mutat. Res.*, 543: 155-163.

- 21-Moller, P.; Loft, S.; Alfthan, G. and Freese, R. (2004). Oxidative DNA damage in circulating mononuclear blood cells after ingestion of black current juice or anthocyanin —rich drink. *Mutat. Res.*, 551: 119-126.
- 22-Shi, J.; Yu, J.; Pophorly, JE. and Kaakuda, Y.(2003). Polyphenolic in grape seeds-biochemistry and functionally. *J. Medicinal Foods*, 50(21):6217-6221.
- 23-De Castillo, ML. ; Dobson, G.; Brennan, R. and Gordon S. (2004). Fatty acid content and juice in black current (*Ribes nigrum* L.) genotypes. *J. Agric. Food Chem.*, 52: 948-952.
- 24-El-Saleh, SC. (2006). Protection by natural honey against hyperhomocysteinemia in rats. *Vascular Disease Prevention*, 3 (4):313-318.
- 25-El-Saleh, SC. (2006). Protection by *Nigella sativa* (Black Seed) against hyperhomocysteinemia in rats. *Vascular Disease Prevention*, 3 (1):73-78.
- 26-Harborn, JB. (1984). Method of Extraction and Isolation, *Phytochemical Methods*. 2nd Ed. London .New York. Chapman and Hall.
- 27-Seshadri, N. and Robinson , K. (2000). Homocysteine, B vitamin and coronary artery disease. *Med. Clin. North AM.*, 84 (1):215-237.
- 28-Al-Zubiady, AA. (2007). Comparative study between the prophylactics effects of aqueous extract of black current (*Vitis vinifera* L.) and vitamin E on some biological parameters related with heart diseases in oxidative stressed rats. MSc. Thesis, College of Veterinary Medicine, University of Baghdad.
- 29-Luna, LG.(1980).Manual of Histology Staining .Methods of Armed Forces. Institute of Pathology 3rd Ed. McGraw - Hill Book Company, New York and London.
- 30-Steel, RG. and Torries JH. (1980). Principal and Procedures of Statistics. Abiometrical approach 2nd Ed. McGraw - Hill Book Company. New York USA.
- 31-Agarwal, A.; Prabakaran, S. and Allamaneni, S .(2006).What an Andriologist / urologist should know about free radicals and why? *Urolo.*, 67:2-8.
- 32-Smith, R.; Kaune, H.; Parodi, D.; Madariaga, M.; Rios, R.; Morales, I. and Castro, A. (2006). Increased sperm DNA damage in patient with varicocele: relation with seminal oxidative stress. *Hum. Reprod.*, 21:986-993.
- 33-Smith, GR.; Kaune, GH.; Parodi, CD. ; Madariaga, AM.; Morales, DI.; Rios, SR. and Castro, GA. (2007) .Extent sperm DNA damage in spermatozoa from men examined for infertility. Relationship with oxidative stress. *Rev. Med. Chil.*, 135:279-286.
- 34-Ishikawa,T. ;Fujioka, H.; Ishimura,T.;Takenake, A. and Fujisawa, M. (2007). Increased testicular 8-hydroxy -2-deoxyguanosine in patient with varicocele. *B. J. U. Inter.*, 100: 863-866.
- 35-Lee, HC.; Jeong, YM.; Lee, SH.; Cha ,KY.; Song ,SH.; Kim, NK.; Lee, KW. and Lee, S. (2006). Association study of four polymorphisms in three folaterelated enzyme gene with non-obstructive male infertility. *Hum. Reprod.*, 21:3162-3170.
- 36- Zhou-Cun, A.; Yang, Y.; Zhang, SZ. ; LI, N. and Zhang, W. (2007). Single nucleotide polymorphism C677T in the methylenetetrahydrofolate reductase gene might be a genetic risk factor for infertility for Chinese men with azoospermia or sever oligozoospermia. *Asian J. Androl.*, 9:57-62.
- 37-Ansari, MN. and Bhandari, U.(2008). Protective effect of *Embelia ribes* Burm on methionin-induced hyperhomocysteinemia and oxidative stress in rat brain. *Indian J. Experim. Biol.*, 46:521-527.
- 38-Kapoor, P.; Ansari, MN. and Bhandari, U.(2008). Modulatory effect of curcumin on methionin-induced hyperlipidemia and hyperhomocysteinemia in albino rats. *Indian J. Experi. Biol.*, 46:534-540.
- 39-Forges, T.; Monnier-Barbarino, P.; Alberto, JM.; Gueant —Rodriguez, RM.; Daval, JL. and Gueant, JL. (2007). Impact of folate and homocysteine metabolism on human reproductive heath . *Hum. Reprod Update.*, 13: 225-238.

- 40- Somnez, M.; Yuce, A. and Turk, G. (2007). The protective effect of melatonin and vitamin E on antioxidant enzyme activities and epididymal sperm characteristics of homocysteine treated male rats. *Reprod Toxicol.*, 23:226-231.
- 41-Kelly, TL.; Li, E. and Trasler, JM. (2003). 5-aza-2'-deoxycytidine induces in murine spermatogenesis and pregnancy outcome. *J. Androl.*, 24: 822-830.
- 42-Aubry, F.; Habasgue, C. and Satie, AP.(2000).Expression and regulation of CCchemokine monocyte chemoattractant protein -1 in rat testicular cell in primary culture .B. 10 *Reprod.*, 62:1427-1435.
- 43-Herrero, MB. ; De Lamirande, E. and Ganong, C. (2003). Nitric oxide is a signaling molecule in spermatozoa. *Curr. Phann. Des.*, 9: 419-425.
- 44-Lee, NP and Cheng, CY.(2004). Nitric oxide /nitric oxide synthase spermatogenesis, and tight junction dynamics. *Biol. Reprod.*, 70 : 267-276.
- 45- Papadopoulos, V.; Kamtchouing, P. and Drosdowsky, MA. (1987). Effect of the transmethylation inhibitor S-adenosyl-homocysteine and of the methyl donor Sadenosyl-methionine on rat Leydig cell function in vitro. *J. Steriod Bioche.*, 26:93-98.
- 46-Castilla-Cortazar, I.; Garcia, M. and Quiroga, J. (2000). Insulin - likes growth factor - I reverts testicular atrophy in rats with advanced cirrhosis. *Hepatol.*, 31:592-600.
- 47-Moran, FM.; Ford, JJ.; Corbin, CJ.; Mapes, SM.; Njar, VC.; Brodie, AM. and Conley, AJ. (2002). Regulation of microsomal P450 , redox partner proteins, and steriodogenesis in the developing testes of the neonatal pig. *Endocrinol.*, 143: 3361-3369.
- 48-Comhaire , FH.; El Garem, Y.; Mahmuod, A. and Eertmans, F. (2005). Combined conventional/antioxidant Astaxanthin ' treatment for male infertility: a double blind, randomized trial. *Asian J. Androl.*, 7:257-262.
- 49-Greco, E.; Iacobelli , M.; Rienzi, L.; Ubaldi ,F.; Ferro, S. and Tesarik, J. (2005). Reduction of the incidence of sperm DNA fragmentation by oral antioxidant treatment . *J. Androl.*, 26:349-353.
- 50-Menezo, YJ.; Hazout, A.; Panteix, G.; Robert, F.; Roll, J.;Cohen-Bacrie, P.; Chapuis, F.; Clement, P. and Benkhalifa, M. (2007). Antioxidant to reduce sperm DNA fragmentation: an unexpected adverse effect .*Reprod Biomed.*, 14:418-421.
- 51- Matthews. (2002). Methyl enetetra hydrofolate reductase :a common human polymorphism and its biochemical implications. *Chem. Rec.*, 2:4-12.
- 52-Shao , ZH.; Vanden Hoek, TL. and Li, CQ.(2004). Synergistic effect of scutellaria biacalensis and grape seed proanthocyanidins on scavenging reactive oxygen species in vitro. *AM. J. Chinese Med.*, 32(1):89-95.
- 53-Arston, M.; Weinbauer, GF.; Schlatt, S.; Shahab, M. and Nieschlag, E.(1999). F.S.H and testosterone, alone or in combination, initiate testicular growth and increase the number of spermatogonia and Sertoli cells in a juvenile non-human primate (*Macaca mulatta*). *J. Endocrinol.*, 136:235-239.
- 54-Walczak-Jedrzejowska, R.; Kula, K.; Oszukowska, E.; Marchlewska, K.; Kula, W. and Slowikowska-Hilczer, J. (2011). Testosterone and oestradiol in concert protect seminiferous tubule maturation against inhibition by GnRH-antagonist. *Inter. J. Androl.*, 1365-1378.
- 55-McLachlan, RI; O' Donnell, L.; Meachem, SJ.; Stanton, PG.; de Kretser, DM.; Pratis , K. and Robertson, DM. (2002). Identification of specific sites of hormonal regulation in spermatogenesis in rats, monkey and man. *Recent Progress in Hormoners Res.*, 57:149-179.
- 56-Haywood, M.; Spaliviero, J.; Jimenez, M.; King, NJ.; Handelsman, DJ. and Allan, CM. (2003). Sertoli and germ cell development in hypogonadal (hpg) mice expressing transgenic follicular-stimulating hormone alone or in combination with testosterone. *Endo crinol.*, 144:509-517.
- 57-Bansal, AK. and Bilaspuri, GS. (2009). Antioxidant effect of vitamin E on motility, viability and lipid peroxidation of cattle spermatozoa under oxidative stress. *Anim. Sci. Reprod.*, 27(1):5-14.
- 58-Bagchi, D.; Garga, A.; Krohn, RL. ; Bagchi, M.; Tran, MX. and Stohs, SJ. (1997). Oxygen free radical scavenging abilities of vitamin C, E and grape seed proanthocyanidin extract in vitro. *Res. Commun. Mol. Pathol. Pharmacol.*, 95:179-189.