Antioxidant activity of Cinnamic acid against Trichlorfon in mice

Nibal Kh. Mousa¹, Saad H. Khudier¹, Abrar N. Mohammad¹, Shahad Sh. Sabbar¹, Ishrak Ab. Ahmed²

Received: December 24,2012/ Accepted: June 16,2013

Abstract: The study was carried out to determine the toxic ,oxidant and antioxidant effects of cinnamic acid in comparison with Vitamin C towards the reduce effect of trichlorfon pesticide ,which is a chemical compound that damage hepatic cells and has mutagenic effects. The effect was studied in mice and the bioassay system depended on evaluating the enzymatic activity of two antioxidant enzymes: Catalase and Glutathione Reductase .Two concentrations of pure cinnamic acid 60 and 30 mg/ body weight were evaluated to choose the suitable concentration . In order to use it in the interaction experiments included two types of treatment pre-Trichlorfon and post — Trichlorfon to determine the mechanisms of pure cinnamic acid which showed no toxic and oxidant effects in biological system and it showed highly performance in preventing and reducing the oxidant stress influences of trichlorfon. It was increased the catalase and glutathione reductase antioxidant activity especially in dose 30mg/ body weight. The positive effect was higher when pure cinnamic acid was used as post- trichlorfon treatment and to less extent in pre —Trichlorfon treatment. The cinnamic acid can be considered as an antioxidant compound and Desmutagen in the first degree and Bioantimutagen in the second degree

Key words: Cinnamic acid, Trichlorfon, Catalase, Glutathione Reductase, Mice.

فعالية تضاد الاكسدة لحامض السيناميك على بقايا مبيد تريكلوروفان في الفئران المختبرية

نبال خليل موسى 1 ،سعد حسين خضير 1 ،ابرار ناطق محمد 1 ،شهد شكري صبار 1 ،إشراق عبد الأمير علي 2

1دائرة تكنولوجيا وبحوث البيئة والمياه - وزارة العلوم والتكنولوجيا

المركز الوطنى للرقابة والبحوث الدوائية ،وزارة الصحة

الإستلام: كانون الاول 24 -2012/القبول: حزيران 16 -2013

الخلاصة: أجريت الدراسة للكشف عن التأثير السمي المؤكسد والمضاد للأكسدة لحامض السيناميك النقي cinnamic acid ومقارنته بفيتامين C تجاه بقايا المبيد ترايكلورفون Trichlorfon والذي يعد مركبا" كيمائيا" يسبب تلف خلايا الكبد ويمتلك تأثيرا سميا" مؤكسدا"، وباستخدام نظام اللبائن في الفئران المختبرية وبالاعتماد على تقييم الفعالية الإنزيمية للمضادين للأكسدة: الكاتليز وكلوتاثايون المختزل لمجانس خلايا الكبد.استخدم تركيزين لحامض السيناميك النقي (30 ،60) ملغم لوزن الحيوان، وكل على انفراد لاختبار فعلها المؤكسد والمضاد للاكسدة وتم انتخاب التركيز الامثل للمركب والذي اعطى نتائج افضل من الحالة الطبيعية السيطرة السالبة. اجري التداخل مابين التركيز الامثل والمؤكسد للترايكلورفون Trichlorfon وبشكل معاملتين قبل وبعد العامل المؤكسد لمعرفة الالية التي يعمل بها هذا المركب في منع او تقليل الاثر التاكسدي للمبيد فقد عمل على زيادة فعالية انزيمي الكاتاليز والكلوتاثايون المختزل المضادين للاكسدة وقد

¹ Environmental-Water Research and Technology Directorate, Ministry of Science and Technology

²The National Center for Drug Control and Research, Ministry of Health

كان الفعل الاكثر ايجابية عند استعمال حامض السيناميك النقي بجرعة 30 ملغم /كغم بعد العامل المؤكسد وبدرجة اقل عند معاملة الحيوانات بحامض االسيناميك النقي قبل العامل المؤكسد و يمكن تصنيف فعل هذا المركب في نظام اللبائن كونه مضاد للاكسدة ومثبط مباشر Desmutagens بالدرجة الأولى ومثبط حيوية Bioantimutagens بالدرجة الثانية.

Introduction

Oxidative stress is responsible for various pathological conditions such as cancer, cardiovascular diseases, arthritis, inflammation, asthma. neurodegenerative disorders and dementia Pesticides and (1).insecticides, persistent organicpollutants that cause oxidative stress in animals, including humans. Trichlorfon -phosphate insecticide is an organo used to control cockroaches, crickets, silverfish, bedbugs, fleas, cattle grubs, flies, ticks, leaf-miners, and leafhoppers (2) as well as treating domestic animals for control of internal parasites (3). Moderate to severe symptoms of trichlorfon toxicity have been reported in farm and factory workers, military personnel-and exterminators (4).

The common human symptoms of trichlorfon toxicity include nausea, diarrhea, vomiting, abdominal cramps, headache, dizziness, eye pain, blurred vision, tears, sweating, and confusion. Severe poisoning may affect the activity of the central nervous system leading to in-coordination, slurred speech, loss of reflexes, involuntary muscle contraction and eventually paralysis of the respiratory muscles and other extremities (5). Endogenous antioxidants are essential defense weapons of the cell against free radicals and oxidative damage. However, chronic exposure to pesticides has led to the induction of oxidative stress beyond the protecting efficiency of the endogenous antioxidants. Exogenous antioxidants from natural plant sources have been widely used to counteract pesticide-induced oxidative stress(6). Phenolic acids are widespread in the plant kingdom and are known to possess a number of biological activities (7). Because the Scientifics and local tends to use the natural products specially the graces in medical and nutrition yield that made us to focus our immediately study to evaluated the hepatoprotective effect of pure cinnamic acid in one of the biosystem.

Materials and Methods

Phosphate Buffer Solution (PBS) (9).Beta-Nicotinamide Adenine Dinucleotide Phosphate Sodium Salt(NADPH+H)(1Mm) (9).Hydrogen peroxide (H₂O₂) prepared from mixing 0.112 ml from H₂O₂ (30%) with 100ml from PBS (10) . Bovine Serum Albumin (BSA)1% prepared dissolving 1g BSA in 100 ml sterile distiller water (10).Colchicine Solution prepared by1mg dissolving of colchicines tablet in one milliliter from sterile distiller water and inject mouse with 0.25 ml from colchicines solution in intraperitoneal membrane before anatomy(2.5-3)hours and it used immediately(11).Biuret solution preparing is to (10).

Doses

Two doses from the pure cinnamic acid were (CA1=60 , CA2=30) mg / body weight, vitamin C(180mg/ body weight)as comparative groups(12) and

Trichlorfon pesticide in(10 mg/body weight) as a positive control(13) and the PBS as a negative control(9).

Experimental plan

To study the oxidant effect and the antioxidant in laboratory animals: The Cinnamic acid was gulping orally by syringe 1 ml size. The Trichlorfon solution was injected Intraperitonially because it lost after (3-12) hours by urine (13,14). Fifty -five mice weighing

20-25 g were divided into three groups as in experimental plan, Figure (1): 1stgroups: we have taken perfect cinnamic acid concentration consist of (negative control (PBS) and positive control (Trichlorfon) comparative group. vitamin C. and two cinnamic acid concentration (30.60) mg / body weight).

2nd groups: pre(before)-Trichlorfon. 3rd groups: post(after) –Trichlorfon.

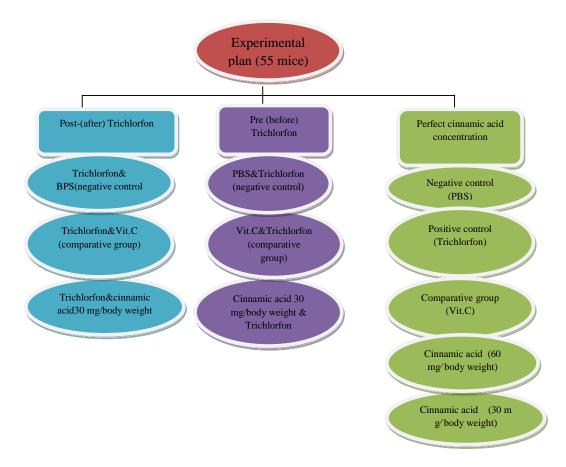


Figure (1): Experimental plan

Preparing of liver mouse serum

Weight 1 g from the liver kept in freez and cut it to very small pieces by sharp knife in 1 ml from PBS and using in the same time the mechanism pressure of hand to crush the liver tissue till be sticky solution then move the rest to the centrifuge with9,000 rpm for 20 minutes .Get the upper layer and let the remainder in the bottom of the test tube avoid the fatty layer above it and store in freezer (-20)C until evaluation (8,10).

Evaluation the Glutathione Reductase

To evaluate the activity of enzyme in volume 1ml,contains interaction mixture: 0.1 ml from BSA 1%., 0.4 ml from oxidation Glutathione, 0.4 ml from 1mM NADPH+H.,0.1ml from the enzyme extraction then read the interaction mixture in spectrophotometer in 340 nm and recorded the reading every 30 second for half an hour.

The Glutathione Reductase (GR) evaluation based on moving hydrogen of NADPH₂ to the oxidation Glutathione then measure the interaction by spectrophotometer: the continuous of interaction led to the reduction of absorbance as a result to change the oxidized GSH to reduced GSH. The unit of enzyme evaluation based on the reduction of the amount of absorbance through one minute under the standard interaction (18, 19).

Evaluation the Catalase

One milliliter of phosphate buffer and 0.4 ml water were added to 0.1 ml of 'BSA', the reaction was started by adding 0.5 ml H $_2$ O $_2$, the mixture was incubated at 37°C for 1 minute the reaction was stopped by adding 2 ml of dichromate: acetic acid reagent and kept in a boiling water bath for 15 minutes.

The mixture was cooled and absorbance was read at 570 nm. (15).

Evaluation the amount of protein

2ml of sterilize distilled water was added to the first tube (blank) then we add 2ml standard solution of protein to the second tube (standard) added 0.2ml from the test (liver mouse extract) and 1.8ml sterilize distilled water to the third tube (test sample) and then added 5ml Biuret solution to the three tubes and mixed well, then put in water bath at 37°C for 10min. Cooled and read absorbanent at 50nm and calculated amount of protein(9,10) as:

Protein amount = test absorbance X---(gm / 100ml) SAP
SCP=Standard concentrate of protein
SAP=Standard absorbance of protein

Statistical analysis

The statistical analysis is done to get the means ±SE and test the different significant among the means by using Duncan test (20) then differences among the means in interaction experiments were compared between the Vit. C, cinnamic extract and the trichlorfon by using T- test (21).

Results and Discussion

Antioxidant enzymes results for 7 days Catalase enzyme (CAT for 7 days)

The results in figure (1) showed lowing in activity of catalase when treated with trichlorfon, the treatment with comparative group showed that there was statistically significant of specific activity comparison with negative and positive treatment with p value(p≤0.05).Among the means-in

interaction experiments were compared between the Vit. C, cinnamic extract and the trichlorfon by using T- test (21).

The cinnamic acid (60mg/body significantly weight) showed comparative with positive treatment and comparative group while there were no significant with negative treatment. So the cinnamic acid concentration (30mg/body weight) showed no significant comparison with comparative group.

Glutathione Reductase(GR for7 days)

The positive treatment (Trichlorfon) led to low value activity with significant for GR The comparative group gave significant when comparative with negative and positive treatment. Results in figure (2) and table(1)of gulping mice with cinnamic acid concentration(CA1=60mg/body weight) showed significant in comparing with other treatment, while gulping the mice with concentration (CA2=30mg/body weight) of cinnamic acid showed elevating in significantly and significant when compared with the comparative group ($p \le 0.05$).

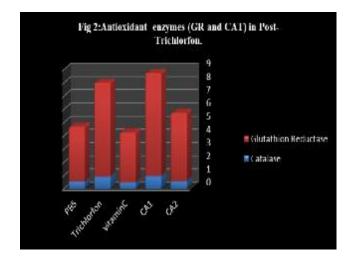


Figure (2): Antioxidant enzymes (GR and CAT) in Post Trichlorfon

Table (1): Antioxidant activity of GR and CAT in mice

| Treatment | Negative control PBS | Positive control Trichlorofon | Vitamine C | Cinnamic acid (60mg/body weight) | Cinnamic acid(30 mg/body weight) | | |
|--------------------------|----------------------------|-------------------------------------|---------------|---|---|--|--|
| Mean ±SE (U/ mg protein) | | | | | | | |
| Catalase | 0.53±0.13 | 0.91±0.3 | 0.16±0.92 | 0.956±0.82 | 0.594±0.21 | | |
| Glutathione reductase | 4.132±0.24 | 7.09±0.05 | 3.75±0.42 | 7.77±0.64 | 5.126±0.29 | | |

Antioxidant and activity of GR and Catalase in mice treated with Trichlorfon comparative with positive control

Catalase enzyme (CAT)

The treatment in figure (3) showed that Vit. C after trichlorfon increasing activity in comparison with control treatment.

When gulping with concentration (30mg /body weight) of cinnamic acid after trichlorfon showed increasing in specific activity when compared with both control and Vit. C treatment with p value ($p \le 0.05$).

Glutathione reductase enzyme (GR)

The treatment in figure (3) and table (2) showed that treating with Vit. C after trichlorfon increasing in specific activity of glutathione reductase when comparison with control treatment. When gulping with (30 mg/body weight) of cinnamic acid after Trichlorfon treatment showed increasing in specific activity when compared with both control and Vit.C treatment with p value ($p \le 0.05$).

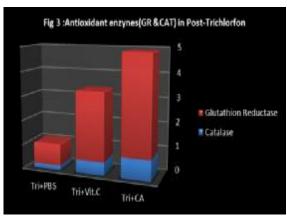


Figure (3): Antioxidant enzymes (GR and CAT) in Post Trichlorfon

Table (2): Antioxidant enzymes (GR&CAT)in post-Trichlorfon

| Treatment | Trichlorfon | Trichlorfon | Trichlorfon | | | | |
|--------------------------|-------------|-------------------|-------------------|--|--|--|--|
| | and | and | and | | | | |
| Test | Phosphate | vitamin C. | Cinnamic acid (30 | | | | |
| | Buffer | | mg/body weight) | | | | |
| | solution | | | | | | |
| Mean ±SE (U/ mg protein) | | | | | | | |
| Catalase | 0.193±0.025 | 0.534 ± 0.034 | 0.846 ± 0.003 | | | | |
| Glutathione | 0.803 ± | 2.905 ± 0.081 | 5.085 ± 0.085 | | | | |
| Reductase | 0.032 | | | | | | |

Enzymatic antioxidants

Mammalian cells are equipped with antioxidant enzymes to minimize the damage resulting cellular from intracellular ROS (reaction oxygen species) or pesticide-induced oxidative stress (22). Catalase and Glutathione reductase work in a coherent manner to inactivate the ROS generated Catalase is responsible breakdown of H₂O₂ to water and oxygen, protecting the cell from the damaging action of H₂O₂ and the hydroxyl radical. Glutathione reductase catalyzes the reaction of hydroperoxides with reduced glutathione to form glutathione disulfide (GSSG) and the reduction product of the hydroperoxides. Organophosphates dichlorvos (trichlorfon) was reported to decrease the activity of these enzymes of mice blood (23) and mice brain (24,25).

Role of cinnamic acid

The antioxidant activity of cinnamic acid and its derivatives are well known in literature (8,29). Among of cinnamic acid derivatives caffeic acid has been reported to exhibit good antioxidant and free radical scavenging activities (29). activities of The antioxidant different cinnamic acid derivatives have been reported earlier, and pcoumeric acid bearing a structural similarity to CA possesses moderate antioxidant activity (29).The antioxidant activity of cinnamic acid able to act as antioxidant as in the following ways: Phenolic hydroxyl groups are good hydrogen donors (30). Hydrogen donating antioxidants can react with reactive oxygen and reactive nitrogen species (31,32) and breaks the cycle of generation of new radicals (33). Following interaction with the initial reactive species a radical form of the antioxidant was produced and had a greater chemical stability than the initial radical (33, 34). Interaction of phenol hydroxyl groups with π -electrons of benzene ring gave molecules with special properties, the ability to generate free radicals where stabilized by delocalization (31). Formation of these long-lived free radicals is able to modify radical-mediated oxidation processes (33).

Antioxidant capacity of phenolic

compounds is also attributed to ability ions chelate metal involved free production of radicals (34). However, phenolic compounds can acts as pro-oxidants by chelating matale in manner that maintains or increas activity or by reducing catalytic metals, thus increasing their ability to form free radicals (34). Hydrophobic benzenoid rings and hydrogen bonding potential of phenolic hydroxyl groups interact with protein and gave cinnamic acid capacity to inhibit some enzymes involved in radical generation (33, 34).

Conclusion

-Cinnamic acid dose (30mg/body weight) has no oxidation effects on mice liver antioxidant enzymes (GR,CAT).

-The toxic effect which reflected from trichlorfon removed and protection in post- Trichlorfon clearly more than pre-Trichlorfon.

-Because Post-trichlorfon treatment was the best results which reflect cinnamic acid dose(30 mg / body weight) Desmutagenes at first degree due to cinnamic acid ability to stop mutagen toxicity before it damage genetic material DNA and mean

cinnamic acid work outside of cell and

References

- 1. Hekimi S; Lapointe, J. and Wen, Y(2011). Taking a "good" look at free radicals in the aging process. Trends In Cell Biology.;21 (10):569-76.
- Karami-Mohajeri, S. and Abdollahi , M. (2011). Toxic effects of organophosphate, arbamate, and organochlorine pesticides on cellular metabolism of lipids, proteins, and carbohydrates:

 A comprehensive review. Hu Exp Toxicol; 30 (9):1119-1140
- Eghbal MA, Pennefather PS, O'Brien PJ. (2004).H₂S cytotoxicity mechanism involves reactive oxygen species formation formation and mitochondrial depolarization .Toxicology;203:69-76.
- Possamai FP, Fortunato JJ, Feier G, Agostinho FR, Quevedo J, Wilhelm Filho D, Dal-Pizzol F.(2007).Oxidative stress after acute and sub-chronic malathion intoxication in Wistar rats. *Environ Toxicol Pharmacol*; 23(2):198-204.
- López- Arrieta J and Schneider L (2008). "Metrifonate for Alzheimer's disease". In López- Arrieta, Jess. Cochrane Database of Systematic Reviews (1).
- Staples, RE. and Goulding, EH.(1979).
 Dipterex teratogenicity in the rat, hamster, and mouse when given by gavage.
 Environ Health Perspect; 30:105-13.
- 7. Mehrdad, M.,Mesripour,M.and Raja,A. (2010).Effect of cinnamon extract on the number of spermatocyte and spermatozoa on cells in mice.. Iranian Journal of Medical and Aromatic Plants; 26(1 (47));83-9
- 8. Mousa, N.; Labeeb, Ah., Iman, I. and Ishrak, A. (2011).Biochemical, Hepatoprotective Effects of pure cinnamic acid Against Cyclophosphamide in white mice, Journal of Madenat Alelem College ,3(2)45-65.
- Taubert, D.; Breitenbach, T.; Lazar, A; Censare k, P.; Harlfinger, S. and Berkels, R. (2003).
 Reaction rate constants of superoxide scavenging by plant antioxidants. Free Radic Biol Med; 35:1599-607.
- Al-Zubaidi, L.(2009). Antioxidantal, antimutagenic of pure curcumin against the mutant carbon tetrachloride and its role in mice embryogenesis, Ph.D., Genetic

called extracellular.

- Engineering and Biotechnology Institute for Postgraduate Studies, Baghdad, Iraq.
- 11. Jwad, B. M. (2010). Toxicopathological and Mutagenic effects of fluoride given in drinking water to male mice.MS.c.; University of Baghdad College of Veterinary Medicine.
- 12. Al-Kinani, Eb. (2005).Vitamins-Rols (E,C,A) in mendingImmunology and Cytogenetic effects of Etopside druge in white mice.M. Sc.Dep.Biology, Ibn-Hautham College , Baghdad University ,Baghdad, Iraq.
- 13. Occupational-Health-Services.(1991).MSDS for trichlorfon. Syracuse, NJ: OHS.
- 14. Staples RE, Goulding EH.(1979). Dipterex teratogenicity in the rat, hamster, and mouse when given by gavage. Environ Health Perspect; 30:105-13.
- Bano, M. and Bhatt, DK.(2007).
 Neuroprote -ctive role of a novel combination of certain antioxidants on pesticide induced toxicity in cerebrum of mice. Res J Agri Bio Sci;3:664-9.
- 16. Takimoto, CH.; Calvo, E Pazdur, R.; Wagman, LD.; Camphausen, K A.; Hoskins, WJ.(2008).."Principles of Oncologic Pharmacotherapy"in (Eds) Cancer Management: AMultidisciplinary Approach.11 ed.
- 17. Irshad, M. and Chaudhuri, PS. 2002). Oxidant antioxidant system: Role and significance in human body. Indian J Exp Biol;40:1233-9.
- 18. Sinha AK. Colorimetric assay of catalase. Anal Biochem 1972;47: 389-94.
- 19. Sahoo, A.; Samanta, L. and Chainy-GB.(2000). Mediation of oxidative stress in HCH-induced neurotoxicity in rat. Arch Environ Contam Toxicol;39:7-12.
- 20. Duncan, D. B. (1955). Multiple ranges and multiple F- test biometrics 11: 1-42.
- Steel, R. G. D. and Torrie, J. H.(1980).Principle and Procedure of Statistics.(2nd ed). McGraw Hill: New York
- 22. Sahoo, A.; Samanta L and Chainy GB. (2000). Mediation of oxidative stress in HCH- induced neurotoxicity in rat. Arch Environ Contam Toxicol; 39:7-12.
- 23. Fridovich I.(1986) Biological effects of the superoxide radicals. Arch Biochem Biophys;287:1-11.

- 24. Singh,R. and Sharma,P.(2011) Hepatoprotective effect of curcumin on lindane-induced oxidative stress in male Wistar rats. Toxicol Int;18:124-9.
- Sharma P. and Singh R.(2010). Protective role of curcumin on lindane induced reproductive toxicity in male Wistar rats. Bull Environ Contam Toxicol: 84:378-84.
- 26. O'Grady, JG.; Schalm, SW. and Williams, R.(1993). "Acute Liver Failure redefining the Syndromoes." Lancet . 342(8866)273-5.
- 27. Valentão, P.; Fernandes, E.; Carvalho, F.; Andrade, P.B.; Seabra, R.M. and Bastos, M. L. (2003). 'Hydroxylradical and hypochlorous acid scavenging activity of small centaury (*Centaurium erythraea*) infusion. A comparative study with green tea (*Camellia sinensis*)". Phytomedicine. 10, 517-522.
- 28. Valentão, P.; Fernandes, E.; Carvalho, F.; Andrade, P.B.; Seabra, R.M.; Bastos, M.L. (2002). "Studies on the antioxidant activity of *Lippia citriodora* infusion: scavenging effect on superoxide radical, hydroxyl radical and hypochlorous acid" *Biol. Pharm. Bull.*, 25, 1324-1327.
- 29. Heim, K. E.; Tagliaferro, A. R. and Bobilya, D.J.(2002). "Flavonoid antioxidants: chemistry, metabolism and structure activity relationships." J. Nutrit. Biochem., 13, 572-584.
- 30. Choi, H.R.; Choi, J.S.; Han, Y.N.; Bae, S.J. and Chung, H.Y.(2002). Peroxynitrite scavenging activity of herb extracts. *Phytother. Res*, 16, 364-367.
- 31. Payá, M.; Halliwell, B. and Hoult, J.R.S. (1992) "Interactions of a series of coumarins with reactive oxygen species. Scavenging of superoxide, hypochlorous-acid and hydroxyl radicals". *Biochem. Pharmacol.*, 44, 205-214.
- 32. Parr, A. J. and Bolwell, J.P.(2002). Phenols in the plant and in man. The potential for possible nutritional enhancement of the diet by modifying the phenols content or profile. *J. Sci. Food Agric.*, 80,985-1012.
- 33. Yang, C.S.; Landau, J.M.; Huang, M.-T. and Newmark, H. L. (2001).Inhibition of carcinogenesis by dietarypolyphenolic compounds. *Annu. Rev. Nutr.*, 21,381-406.
- 34. Croft, K.D. (1998). The chemistry and biological effects of flavonoids and phenolic acids. *Ann. N. Y. Acad. Sci.*, 854, 435-442.