

COEXPRESSION OF HER2 AND P53 IN GASTRIC AND ESOPHAGEAL ADENOCARCINOMA

GINA GAMES GEORGE*
HAYDER HUSAIN IBRAHIM**
SARDAR HASSAN ARIF***
INTISAR SALIM PITY****

Submitted 1 October 2015; accepted 31 December 2015

ABSTRACT

Background and objectives: Gastric and esophageal adenocarcinoma remains deadly diseases with an on rise incidence. The recently discovered cancer-related molecular markers, such as HER2 and p53, help facilitate response to preoperative therapy and improve overall survival. This study was aimed to detect the immunoexpression of HER2 and p53 in gastric and esophageal adenocarcinoma and to determine the association of these two markers with clinicopathological parameters.

Method: The study was conducted in the Central Laboratory and Directorate of Health, Duhok-Iraq during a period from May 2009 to September 2014 on 101 gastric and esophageal adenocarcinoma cases. Using monoclonal antibodies against HER2 receptors and p53 nuclear protein, slides were stained with the fully automated immunostaining instrument, Ventana Benchmark.

Results: Total positive HER2 immunoexpression was demonstrated in 33.7% of cases with a significantly higher dense HER2 (3+) expression in esophageal adenocarcinoma compared with its gastric counterpart. p53 nuclear staining was observed in 62.4% of cases; it was significantly higher in gastric cancer than esophageal adenocarcinoma. HER2 was limited to the intestinal type whereas p53 was found to be expressed in both intestinal and diffuse types. No significant coexpression was demonstrated between HER2 and p53 in any of gastric or esophageal adenocarcinoma.

Conclusions: HER2 expression was limited to the intestinal type gastric adenocarcinoma. No significant coexpression of HER2 and p53 was demonstrated in both of gastric and esophageal adenocarcinoma.

Duhok Med J 2015; 9 (2): 60-73.

Keywords: HER2, p53, gastroesophageal adenocarcinoma.

Gastric and esophageal (gastroesophageal) adenocarcinomas form a substantial number of cancer cases with a dramatic rising incidence over the past 20 years, particularly among young adults¹⁻⁷. The overall 5-year survival rates are up to 27% with no significant change over the past 40 years despite advances in surgical treatment and chemotherapy^{2,4,5,6,8}. Discovery of new molecular markers and novel pharmacogenetic traits helped improve patients care, fostered hope and applied new directions of cure⁹⁻¹². Markers

* Lecturer in Pathology, Department of Pathology, College of Medicine, University of Duhok, Iraq.

** Assistant professor in Surgery, Department of Surgery, College of Medicine, University of Duhok, Duhok, Iraq.

*** Lecturer in Surgery, Department of Surgery College of Medicine, University of Duhok, Duhok, Iraq.

**** Professor in Histopathology, Department of Pathology, College of Medicine, University of Duhok, Duhok, Iraq.

Correspondence: Professor Intisar Salim Pity, Pathology department,
E.mail: intisarsalimpity@gmail.com Mobile:07504788000

of interest include those contributed in growth regulation like human epidermal growth factor receptor-2 (HER2) and those involved in apoptosis and cell cycle control, like p53⁸⁻¹². Given the advantages of both HER2 and p53 as responders for target therapy and predictors for better outcome in breast cancer, such properties facilitated numerous studies that are intriguing in identifying those markers in gastric adenocarcinoma. On the other hand although overexpression of HER2 and p53 in gastroesophageal adenocarcinomas may increase their dismal outcome with a resistance to the conventional chemotherapy, many ongoing studies conducted in this field are intriguing in that patients get benefit from specific therapy targeting these molecular markers, both for prognostic and therapeutic purposes^{9,12}.

The current study may provide an insight on the immunohistochemical expression of HER2 and p53 immunomarkers in gastric and esophageal adenocarcinoma in Kurdistan-Iraq. To the best of our knowledge, no previous study was conducted in the same field to evaluate the coexpression of HER2 and p53 in adenocarcinoma of stomach and esophagus in this particular area of Iraq.

MATERIALS AND METHODS

The study was conducted in the Central Laboratory/Directorate of Health, Duhok-Iraq. Specimens were retrieved from histopathologic laboratories in Duhok Region during the period from May 2009 to September 2014. Paraffin embedded, pretreatment (endoscopic biopsy or

gastrectomy) specimens were available for 101 patients with newly diagnosed gastric (n=63) and esophageal (n=38) adenocarcinoma. Information pertaining to the patient's age at presentation, gender and type of operation were obtained from patient's request forms.

Four micron-thick tissue sections were taken from the tumor, processed and embedded in paraffin wax, then stained again with Hematoxylin and Eosin (H&E) stains to confirm the diagnosis of adenocarcinoma and for grading purposes. Tumors were classified into 2 types, intestinal and diffuse (signet ring). The intestinal-type adenocarcinomas were further graded, according to the modified WHO classification system, into low grades (well and moderately differentiated) and high grades (poor and undifferentiated). Pathological staging, applied only on gastrectomy specimens (n= 43), was done according to the pathologic TNM tired staging system from I-IV based on the microscopic examination of the primary tumor within organ wall, all available lymph nodes, omentum and any associated structure if available¹³.

The immunohistochemical technique applied was streptavidin-biotin system, using monoclonal antibodies manufactured by Ventana Corporation (Ventana, Rocklin, Calif), the chromogen used was 3-3'-diaminobenzidine tetrahydrochloride (DAB) and a standard DAB detection kit (Ventana) was used according to instructions supplied by the manufacturer's (Ventana) and as described previously by Pity et al and Pity and Baizeed^{14,15}. Representative tissue sections from the

tumor (without necrosis and little mesenchymal tissue) were selected from the paraffin blocks. Three μm tissue sections were cut with a manual microtome and mounted on poly-L-lysine-coated slides. Sections were placed in oven at 56-60 C° overnight, then stained with a fully automated immunostaining instrument; Ventana Benchmark (Ventana Medical System Inc., Cell Margue, Ventana, Rocklin, Calif.) where deparaffinization, dehydration, antigen retrieval in addition to the application of primary and secondary antibodies were achieved. The primary antibodies used included monoclonal antibodies for HER2 (REF-790-2991, Ventana, USA) and for p53 (REF-760-2542, Ventana, USA). Positive controls (strongly positive breast carcinoma for both HER2 and p53) and negative controls (using the same procedure without primary antibodies) were used with each run. Sections were counterstained with Mayer's hematoxylin, dehydrated through graded alcohols to xylen and then mounted with DPX solution and coverslipped.

Positive p53 protein expression was defined as clear nuclear immunostaining in more than 10% of tumor cells¹⁶. HER2 staining was evaluated as described by Hofmann et al who addressed 4-graded scales [grade 0 referring to tumors without detectable staining or membrane staining of less than 10%, grade 1+ pertaining to weak staining of greater than 10% of tumor cells, grade 2+ which is defined as weak to moderate staining of the entire cell membrane (thin ring) in more than 10% of tumor cells and grade 3+ reflecting

moderate to strong staining of the entire cell membrane (thick ring) in more than 10% of tumor cells]¹⁷. Both (0 and 1+) grade scales are considered negative while grade 2+ and 3+ indicate staining¹⁸.

Statistically, the collected data were organized and tabulated, and descriptive statistics were used to summarize demographic variables. Chi square and Fisher exact tests were used for testing associations between categorical tumor parameters, and differences at the level of $p \leq 0.05$ were considered as statistically significant.

RESULTS

Patient's ages ranged between 28-90 years (mean: 61.7 years). Sixty six patients were males and 35 were females. Gastric specimens (n= 63) included 43 (68.3%) gastrectomy specimens and 20 (31.7%) endoscopic biopsies. All esophageal specimens (n=38) were endoscopic biopsies from the lower esophagus. Histologically, 49 (77.8%) gastric cases were intestinal type and the remaining 14 (22.2%) cases were diffuse type adenocarcinoma whereas all esophageal cancers were intestinal type adenocarcinoma. Thus the total (gastric and esophageal) intestinal type adenocarcinoma formed 87 cases; of these, 41 (47.1%) cases were low-grades and 46 (52.9%) were high-grades. Of the 43 gastrectomy specimens, the tumor (T) status comprised 2 (4.7%) T1, 10 (23.2%) T2, 26 (60.5%) T3 and 5 (11.6%) T4. The lymph node (N) status formed 11 (25.6%) N0, 21 (48.8%) N1, 8 (18.6%) N2 and 3 (7%) N3.

HER2 and p53 Expression

Positive HER2 membranous expression (scores +2 and +3) was demonstrated in 34 (33.7%) cases while p53 nuclear staining was observed in 63 (62.4%) cases. No significant association of any marker was

observed with age and gender. The highest frequency of positive {20.8% of HER2 and 36.6% p53} cases was observed among 60-69 year age group (Figure 1), and there was trend toward male gender (Figure 2).

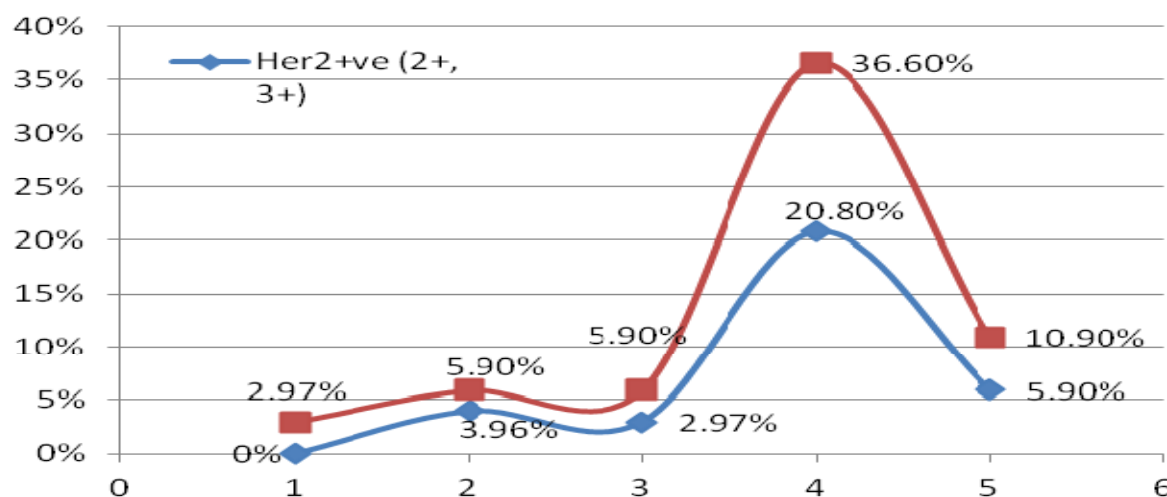


Figure 1. Age distribution for positive HER2 and p53 in gastroesophageal adenocarcinoma cases (Fisher exact test used, $p=0.5$ for HER2 and 0.1 for p53).

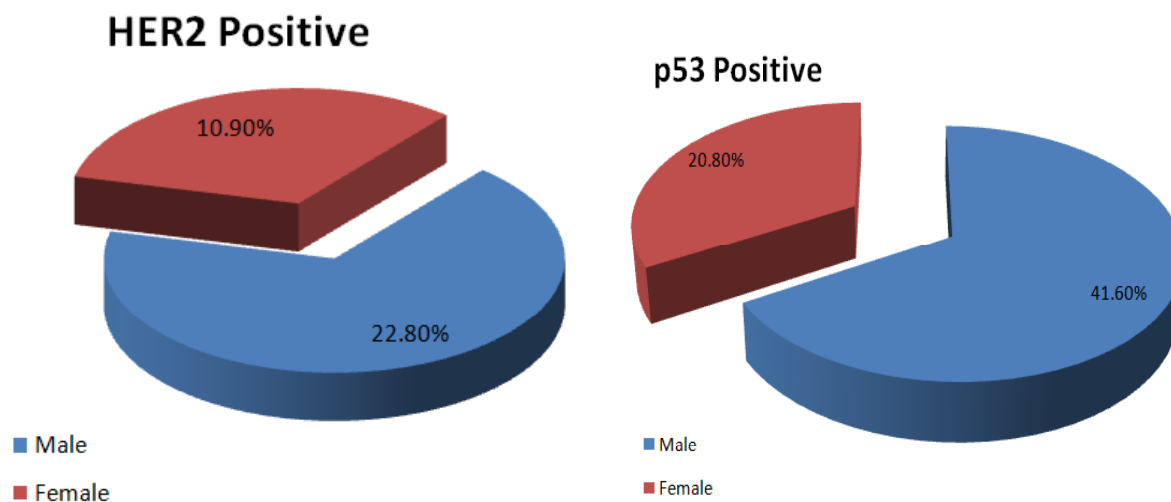


Figure 2. Gender distribution and positive HER2 ($p=0.06$) and p53 ($p=0.073$). χ^2 used.

Forty-five gastric and 22 esophageal cancers showed a negative HER2 (0/1+) expression. The remaining positive cases showed a significantly higher dense HER2 (3+) expression among esophageal compared with the gastric cases but lighter density HER2 (2+) was demonstrated only

in GC where it was identified in 10.9% of cases (Table 1).

Much higher p53 nuclear expression was demonstrated in gastric 46 (45.6%) compared with the esophageal adenocarcinoma 17 (16.8%), $p=0.06$.

Table 1. HER2 scores and site of adenocarcinoma.

	Gastric cancer	Esophageal cancer	Total
HER2			
Negative HER2 (0/+1)	45 (44.6%)	22 (21.8%)	67 (66.4%)
Positive HER2 (+2)	11 (10.9%)	0 (0%)	11 (10.9)
Positive HER2 (+3)	7 (6.9%)	16 (15.8%)	23 (22.8%)
Total*	63 (62.4%)	38 (37.6%)	101 (100%)
P53			
P53 +ve	46 (45.6%)	17 (16.8%)	63 (62.4%)
P53 -ve	17 (16.9%)	21 (20.8%)	38 (37.6%)
Total**	63 (62.4%)	38 (37.6%)	101 (100%)

*: X^2 , $p = 0.05$, **: X^2 , $p = 0.08$

Regarding the histologic type, as shown in figure 3 HER2 immunoexpression was demonstrated only in the intestinal adenocarcinoma. It was completely absent in the diffuse type. In contrast, p53 expression was more obvious in the diffuse

(47.5%) than the intestinal type adenocarcinoma (14.9%), but the difference didn't reach the level of significant ($p = 0.36$).

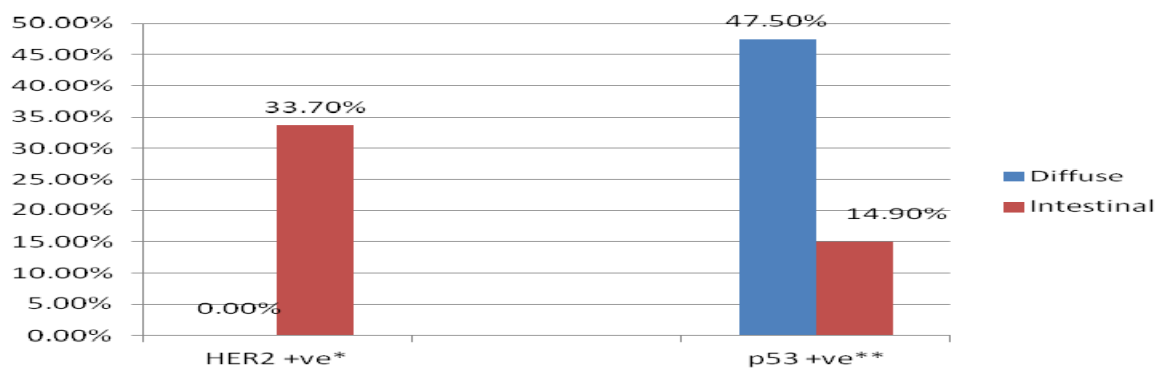
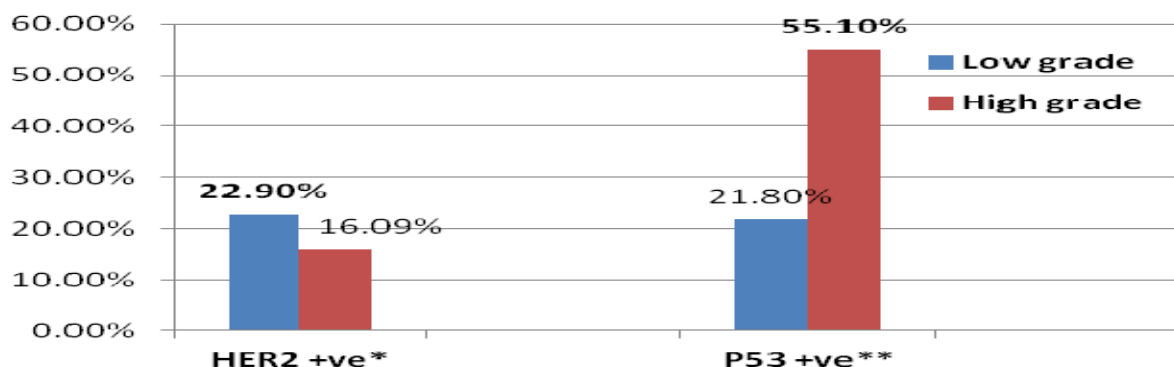


Figure 3. HER2 and p53 immunoexpression and histologic type of adenocarcinoma. (Fisher exact test, $p = 0.36$).

No statistical significance could be demonstrated between any marker and tumor grade. There was a trend for HER2 toward low grade tumors and p53 toward high grade cancers (Figure 4).



* $p = 0.22$, ** $p = 0.09$ for p53.

Figure 4. HER2 and p53 immunoexpression and tumor grade.

Considering TNM staging, no significant association was demonstrated between any of HER2 or p53 and T-status despite a trend toward T3 (Figure 5).

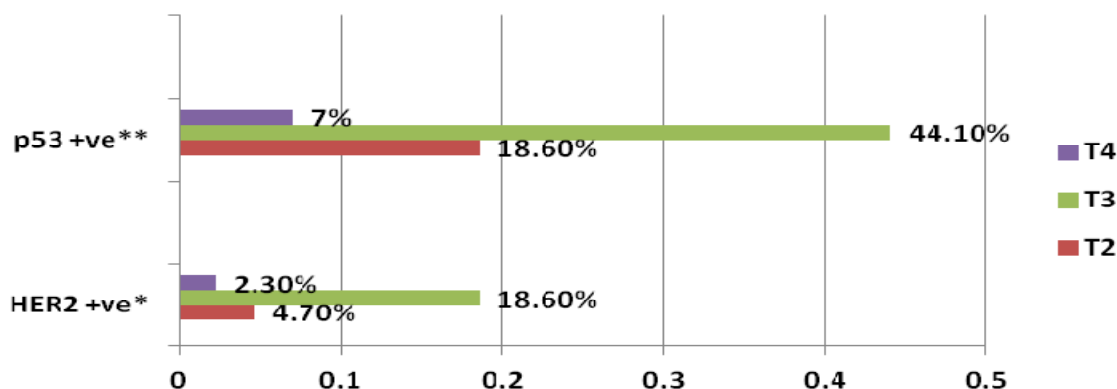


Figure 5. HER2 and p53 expression and T-status.
Fisher exact test used, *: $p = 0.93$; **: $p = 0.14$

As shown in figure 6, no statistical differences were observed between any of HER2 or p53 and N-status despite a trend for both markers toward N1.

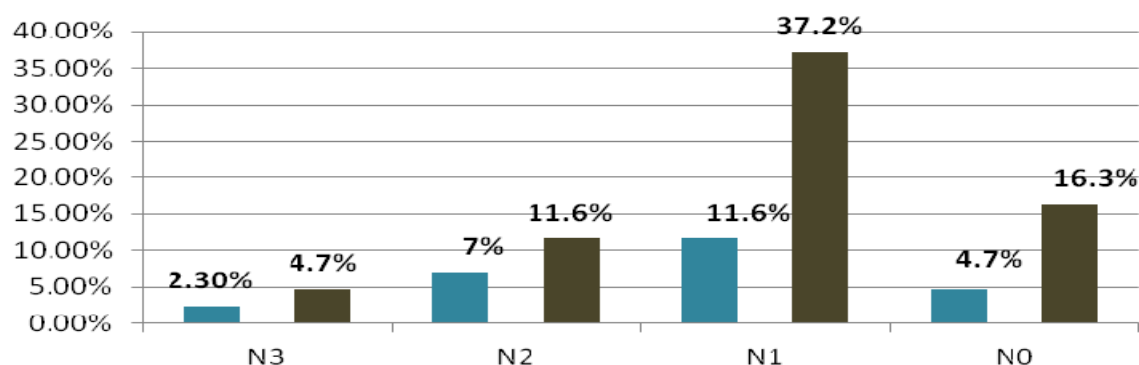


Figure 6. Distribution of gastric cancer cases, according to the the nodal (N) status (n= 43).
Fisher exact test used; *: $p = 0.8$; **: $p = 0.87$.

Coexpression of HER2 and p53

Among gastric cancer cases, 19.1% of cases illustrated coexpression of both markers and 53.9% showed negative HER2/positive p53. Lack of both markers was observed in 17.5% of cases, and the remainders (9.5%) showed positive HER2/negative p53. No significant association was found between presence and absence of the 2 markers (Figure 7).

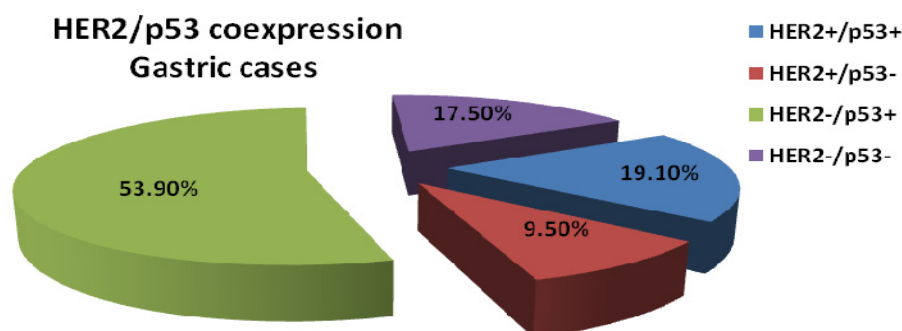


Figure 7. HER2/p53 coexpression in gastric adenocarcinoma, Fisher exact test used, $p = 0.67$.

On the other hand, all esophageal adenocarcinoma showed either absent HER2, p53 or both markers. No coexpression of both markers was demonstrated among esophageal cases. The difference was statistically significant (Figure 8).

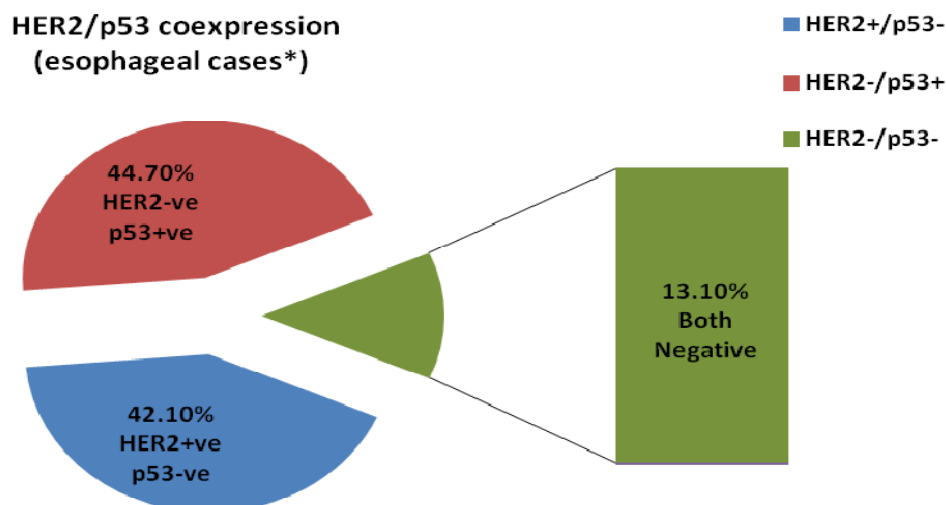


Figure 8. HER2/p53 coexpression in esophageal adenocarcinoma, *Fisher exact test, $p = 0.034$.

DISCUSSION

Overall, this study demonstrated a relatively high HER2 positivity (33.7%) in gastric and esophageal adenocarcinoma compared with what was previously reported by Kunz et al and Kataoka et al among Americans and Japanese with gastric cancer (8.5% to 10.3%)^{19,20}. By a sequence analysis done in 122 centers out of 24 countries with gastric and esophageal cancers, the Trastuzumab anti-HER2 target therapy for Gastric Cancer (ToGA) trial demonstrated 10.4% to 22.1% HER2 positivity⁹. Barros-Silva et al, in their study among Portugal's population, observed a wider HER2 positivity range (5.2%-22.6%) but still lower than the current frequency²¹. The relatively higher expression of HER2 among our series is probably the result of combining HER2 (2+ and 3+) immunoexpression despite the fact that there is a high concordance between IHC and in situ hybridization for HER2 2+ demonstrated by Hofmann et al

and Dowsett et al in their study of HER2 in both gastric and breast cancers^{17,22}.

It is worth mentioning that HER2-positivity differed by its staining intensity among our series. A strikingly high HER2 (3+) expression was observed among esophageal cancers compared with its gastric counterpart. In contrast, HER2 (2+) was completely absent among esophageal cancer cases. Tanner et al also demonstrated higher HER2 positivity in gastroesophageal junction cancer than GC (24% versus 12%) on their study among Finland population²³. However, different results have been observed by many other studies^{9,16,18}. The technique applied for HER2 detection with a further subcategorization of HER2 (2+) into positive or negative according to the in situ hybridization reading may influence the detection rate ranges^{17,21,22}.

Concerning p53 nuclear immunoexpression, in the course of this experiment we faced an exciting finding

that it was strikingly high in gastric adenocarcinoma compared with its esophageal counterpart (45.6% versus 16.8%). Wide p53 range rates for both gastric and esophageal adenocarcinoma have been documented in the literature (19% to 90%)²⁷⁻³². The conflicting data demonstrated by different studies probably reside in the differences of geographic populations studied with heterogeneous socioeconomic sample sets studied and dietary habits as well as differences in their personal behaviors². However, variation in the methods applied for evaluation of mutated p53 and the anti-p53 antibodies used may also contribute to such wide range rates³¹.

In the current study, HER2 positivity was limited to the intestinal adenocarcinoma in both gastric and esophageal tumors; it was completely negative in the diffuse type. Interestingly, this finding is in agreement with numerous prior studies conducted in this field among American and European populations^{9,17}. In addition to the fact that mutated HER2 is already low in the diffuse type, lack of HER2 immunoexpression among our diffuse cases is at least partly related to the small sample size of the present study. In contrary to HER2 expression, p53 nuclear expression was found to be expressed in both intestinal and diffuse adenocarcinoma with no significant difference between the two. This finding is comparable to what was observed by Pinto *et al* among Portugal's population³⁰. However, contradictory findings were observed by Zheng *et al* and Lee *et al* among Japanese and Koreans where nuclear p53

overexpression was much more frequent in intestinal than the diffuse type adenocarcinoma^{24,34}.

As far as the grade is concerned, we failed to demonstrate any significant association between any of HER2 and p53 positivity and grading despite a trend toward low grade tumor for HER2 and toward high grade for p53. Divergent results have been demonstrated by many other studies conducted among different geographic populations 24-26 although similar results have been reported in a study done by Gleeson *et al* on gastroesophageal adenocarcinoma³⁵.

The already high T3 and N1 frequencies among our series may explain the predominance of HER2 and p53 positivity among both categories. However, the differences didn't reach the level of significance. Similarly, Kataoka *et al* denied any correlation between pathological stage and HER2 overexpression²⁰.

Moreover, despite a trend toward elderly and male gender neither HER2 nor p53 immunoexpression was significantly associated with any of age groups or gender in the current study. Honda *et al*, in their study among Japanese, have observed a significantly lower p53-immunoreactivity among young patients 36. However, our finding is in sharp contrast to the negative correlation between HER2 with both age and gender that was previously reported by Chen *et al* among Chinese population and the remarkable predominance of mutated p53 among the Romanian male gender^{16,37}.

Another important finding in this study is that no significant difference or association between both markers was demonstrated. Cases lacking HER2 with or without p53 among GC cases were more frequent than those expressing both markers together and all esophageal cancers were negative for at least one marker if not both. Such observation is in sharp contrast to what was previously described by Kataoka et al who suggested a possible role of p53 perturbation in the development of HER2-positive gastric cancer²⁰.

Moreover, the information obtained in this experiment provided a clinicopathological analysis of only 63 patients with GC and 38 patients with esophageal cancer, which is relatively small sample size. As well, we did not perform an in situ hybridization recommended to determine the real HER2 status particularly in equivocal cases, i.e. IHC (2+). These might subject our data to selection bias. More ad hoc-designed studies are needed to clarify these aspects and to ascertain whether HER2 and p53 immunoexpression really reflects their gene mutations and whether alterations of the genes themselves or their pathways have an intercommunicating role in gastric and esophageal adenocarcinoma.

REFERENCES

1. Anderson WF, Camargo MC, Fraumeni JF Jr, Correa P, Rosenberg PS, Rabkin CS Age-specific trends in incidence of noncardia gastric cancer in US adults. *JAMA* 2010; 303:1723-8.
2. American Cancer Society. Cancer Facts and Figures. Atlanta, Ga: American Cancer Society 2012. <http://www.cancer.org/acs/groups>.
3. Balbuena L, Casson AG. Physical activity, obesity and risk for esophageal adenocarcinoma. *Future Oncol* 2009; 5(7):1051-63.
4. Cunningham D, Allum WH, Stenning SP, Thompson JN, Van de Velde CJ, Nicolson M, et al. Perioperative chemotherapy versus surgery alone for resectable gastroesophageal cancer. *N Engl J Med* 2006; 355(1):11-20.
5. Norsett KG, Lagreida A, Midelfart H, Yadetie F, Erlandsen SE, Falkmer S, et al . Gene expression based classification of gastric carcinoma. *Cancer Lett* 2004; 227–37.
6. Kamangar F, Dores GM, Anderson WF. Patterns of cancer incidence, mortality, and prevalence across five continents: defining priorities to reduce cancer disparities in different geographic regions of the world. *J Clin Oncol*; 2006;24(14):2137-50
7. Holmes RS, Vaughan TL. Epidemiology and pathogenesis of esophageal cancer. *Semin Radiat Oncol* 2007; 17:2- 9.
8. Ruschoff J, Hanna W, Bilous M, Hofmann M, Osamura . HER2 testing in gastric cancer: a practical approach. *Mod Pathol* 2012 ; 25:637-50.
9. Bang YJ, Van Cutsem E, Feyereislova A, Chung HC, Shen L, Sawaki A, et al. Trastuzumab in combination with chemotherapy versus chemotherapy alone for treatment of HER2-positive advanced gastric or gastro-oesophageal junction cancer (ToGA): a phase 3, open-label, randomized

- controlled trial. *Lancet* 2010; 376: 687–97.
10. Hu B, El Hajj N, Sittler S, Lammert N, Barnes R, Meloni-Ehrig A. Gastric cancer: Classification, histology and application of molecular pathology. *J Gastrointest Oncol* 2012; 3(3): 251–61
 11. Gibson MK, Abraham SC, Wu T-H, Burtress B, Heitmiller RF, Heath E, et al. Epidermal Growth Factor Receptor, p53 Mutation, and Pathological Response Predict Survival in Patients with Locally Advanced Esophageal Cancer Treated with Preoperative Chemoradiotherapy. *Clin Cancer Res* 2003 ; 9, 6461-8.
 12. Joerger AC, Ang HC, Fersht AR . Structural basis for understanding oncogenic p53 mutations and designing rescue drugs. *Proc Natl Acad Sci USA* 2006; 103: 15056–61
 13. Edge SBB DR, Compton CC, Fritz AG, Greene FL, Trotti A, editors . *AJCC Cancer Staging Manual*. 7th ed. New York: Springer; 2010
 14. Pity IS, Arif SH, Hadji DA. Angiogenesis, p53 and Bcl2 in Colorectal Carcinoma. *IJOART* 2013 ; 2(3):1-6.
 15. Pity IS, Baizeed AM . Identification of *Helicobacter Pylori* in Gastric Biopsies of Patients with Chronic Gastritis, Histopathological, Immunohistochemical Study. *Duhok Med J* 2011 ; 5(1):69-77
 16. Daniela Lazar, Sorina Taban, I. Sporea, Alis Dema, Mariora Cornianu, Elena Lazar, et al. The immunohistochemical expression of the p53-protein in gastric carcinomas. Correlation with clinicopathological factors and survival of patients. *RM Morphol Embryol* 2010; 51(2):249–57.
 17. Hofmann M, Stoss O, Shi D, Büttner R, van de Vijver M, Kim W, et al Assessment of a HER2 scoring system for gastric cancer: results from a validation study. *Histopathol* 2008; 52(7): 797–805
 18. Park DI, Yun JW, Park JH. HER-2/neu amplification is an independent prognostic factor in gastric cancer. *Dig Dis Sci* 2006 ; 51(8):1371-9
 19. Kunz PL, Mojtahed A, Fisher GA, Ford JM, Chang DT, Balise RR, et al. HER2 expression in gastric and gastroesophageal junction adenocarcinoma in a US population: clinicopathologic analysis with proposed approach to HER2 assessment. *Appl Immunohistochem Mol Morphol* 2012 ; 8(1):13–24
 20. Kataoka Y, Okabe H, Yoshizawa A, Minamiguchi S, Yoshimura K, Haga H, et al. HER2 expression and its clinicopathological features in resectable gastric cancer. *Gastric Canc* 2012 ; 8(1):84–93
 21. Barros-Silva, JD, Leitao, D, Afonso, L, Vieira, J, Dinis- Riberio, M, Fragoso, M, et al . Association of ERBB2 gene status with histopathological parameters and disease- specific survival in gastric carcinoma patients. *Br J Cancer* 2009 10;100:487-93.
 22. Dowsett M, Bartlett J, Ellis IO, Salter J, Hills M, Mallon E, et al. Correlation between immunohistochemistry (Hercep Test) and fluorescence in situ hybridization (FISH) for HER-2 in 426 breast carcinomas from 37

- centres. *Journal of Pathology* 2003 ; 199(4):418–423
23. Tanner M, Hollmen M, Junttila TT. Amplification of HER-2 in gastric carcinoma: association with topoisomerase IIa gene amplification, intestinal type, poor prognosis and sensitivity to trastuzumab. *Ann Oncol* 2005 ; 16:273-8.
 24. Tafe LJ, Janjigian YY, Zaidinski M, Hedvat CV, Hameed MR, Tang LH, Hicks JB, Shah MA, Barbashina V. Human epidermal growth factor receptor 2 testing in gastroesophageal cancer: correlation between immunohistochemistry and fluorescence in situ hybridization. *Arch Pathol Lab Med* 2011 ; 8(11):1460–5.
 25. Yu GZ, Chen Y, Wang JJ. Overexpression of Grb2/HER2 signaling in Chinese gastric cancer: their relationship with clinicopathological parameters and prognostic significance. *J Cancer Res Clin Oncol* 2009 ; 8(10):1331–9.
 26. Zhan GH. Multicenter study on HER-2/neu gene amplification and protein expression in patients with gastric cancer. *Chin J Dig* 2006 ; 8(10):657–60.
 27. Kim R, Clarke MR, Melhem MF. Expression of p53, PCNA, and C-erbB-2 in Barrett's metaplasia and adenocarcinoma. *Dig Dis Sci* 1997 ; 42:2453–62
 28. Wang Jy, Lin Sr, Hsieh Js, Hsu Ch, Huang Ys, Huang Tj. Mutations of p53 gene in gastric carcinoma in Taiwan. *Anticancer Res* 2001 ; 21(1B):513–20.
 29. Casson AG, Evans SC, Gillis A, Porter GA, Veugelers P, Darnton SJ, et al. Clinical implications of p53 tumor suppressor gene mutation and protein expression in esophageal adenocarcinomas: results of a ten-year prospective study. *J Thorac Cardiovasc Surg* 2003 ; 125: 1121-31
 30. Pinto-De-Sousa J, Silva F, David L, Leitato D, Deixas M, Pimenta A, et al. Clinicopathological significance and survival influence of p53 protein expression in gastric carcinoma. *Histopathol* 2004 ; 44(4):323–31.
 31. Ismail Hm, Moneer M, El-Baradie M, Khordhid O, Touny A. Clinicopathologic and prognostic significance of overexpression of her-2/Neu and p53 oncoproteins in gastric carcinoma using tissue microarray. *J Egypt Natl Cancer Inst* 2007 ; 19(2):147–57.
 32. Ji F, JIN X, Jiao Ch, Xu Qw, Wang Zw, Chen YI. Fat 10 level in human gastric cancer and its relation with mutant p53 level, lymph node metastasis and TNM staging. *World J Gastroenterol* 2009 ; 15(18):2228–33
 33. Zheng H, Takahashi H, Murai Y, Cui Z, Nomoto K, Miwa S, et al. Pathobiological characteristics of intestinal and diffuse-type gastric carcinoma in Japan: An immunostaining study on the tissue Microarray. *J Clin Pathol* 2007 ; 60: 273-7.
 34. Lee KE, Lee HJ, Kim YH, Yu HJ, Yang HK, Kim WH, et al. Prognostic significance of p53, nm23, PCNA and c-erbB-2 in gastric cancer. *Jpn J Clin Oncol* 2003 ; 33 (4): 173-9.

35. Gleeson CM, Sloan JM, McManus DT, Maxwell P, Arthur K, McGuigan JA, *et al.* Comparison of p53 and DNA content abnormalities in adenocarcinoma of the oesophagus and gastric cardia. *Br J Cancer* 1998 ; 77: 277 – 86.
36. Honda T, Tamura G, Endoh Y, Nishizuka S, Kawata S, Motoyama T, *et al.* Expression of tumor suppressor and tumorrelated proteins in differentiated carcinoma, undifferentiated carcinoma with tubular component and pure undifferentiated carcinoma of the stomach. *Jpn J Clin Oncol* 2005; 35(10): 580–6
37. Chen B, Luo RC, Cui F, Qian XY . Association of HER-2/Neu expression with prognosis of gastric cancer. *Nan Fang Yi Ke Da Xue Xue Bao* 2006 ; 26(3):344-7.

پوخته

لټنډرینا HER2 و p53 ل په نجه شپړا گلاندی یا گه دئ و بوریکا خوارنئ

پېښه کی و نارمانچ: په نجه شپړا گلاندی یا گه دئ و بوریکا خوارنئ د مینیت ټیک ژ نه خوشیښ کوژه ک دگه ل زیده بوونا ریژا توشبوونی. نیشاندهرین گریډای په نجه شپړیڼه ټوین نوی هاتینه دیارکرن و هکی HER2 و p53 د هاریکارن بو بله زکرنه به رسفدانی بو چاره سه ریا به ری نشته رگه ری و ب گشتی ژیانئ باشر لټدکه ن. نارمانچا فئ ټه کولینئ دیارکرنه خونیشاندانا پاریزکرنئ یا HER2 و p53 ل په نجه شپړا گلاندی یا گه دئ و بوریکا خوارنئ و ده ستنیشانکرنه هه ټبه ندیی دناقه را هه ربووا دگه ل پیڅه رین کلینیکی و تاقیگه هی.

ریکین ټه کولینئ: ټه ټه کولینه هاته ټه نجامدان ل تاقیگه ها مه لبه ندیا ریڅه به ریا ساخله میا دهوکی - عیراق دناقه را گولانا ۲۰۰۹ ټی و ټیلونا ۲۰۱۴ لسه ر ۱۰۱ حاله تین په نجه شپړا گلاندیا گه دئ و بوریکا خوارنئ بکارټینانا دژله شین ټیک جور دئ و ده رگرین HER2 و پروتینی ناوه کی یی p53 سلاید هاتنه رهنگکرن ب ټامیری فول ټوتوماتیکی رهنگکرنئ (Ventana Benchmark).

ټه نجام: سه رجه می خونیشاندانا پوزه تیډ یا HER2 ب پیڅانا (۲+ / ۲+) هاته دیارکرن ل ۲۲,۷٪ ژ حاله تان و بشپړه کی به رچاډ ۲+ پتر بوو ل په نجه شپړا بوریکا خوارنئ ببه راوردی دگه ل په نجه شپړا گه دئ، و به روڅاڅی ۲+ هه ر هیچ دیارنه بوو ل په نجه شپړا بوریکا خوارنئ. رهنگدانه ټه یا p53 هاته دیتن ل ۶۲,۴٪ ژ حاله تان و بشپړه کی به رچاډ پتر بوو ل په نجه شپړا گه دئ ژ په نجه شپړا بوریکا خوارنئ.

ده رټه نجام: HER2 ب تنی ل جورئ رویڅیکی هاته دیتن ل پ53 لهه ربوو جورین رویڅیکی و به ربه لاف هاته دیتن. چ خونیشاندانین بهه فا یین HER2 و p53 نه هاتنه دیتن لهه ربوو په نجه شپړین گه دئ و بوریکا خوارنئ.

الخلاصة

معاينة HER2 و p53 في السرطان الغدي للمعدة والمرئ

الخلفية وأهداف البحث: تبقى السرطانات الغدية للمعدة والمرئ أمراض مميتة مع ارتفاع نسبة الإصابة بها، أما العلامات الجينية المتعلقة بالأورام أو المكتشفة حديثاً مثل HER2 و p53 تساعد على تسهيل كل من الاستجابة للعلاج قبل الجراحة وتحسين البقاء على قيد الحياة بشكل عام.

تم تصميم هذه الدراسة لتقييم التعبير المناعي لكل من HER2 و p53 في السرطانات الغدية للمعدة والمرئ ولإيجاد الارتباط بين HER2 و p53 مع الأنماط السريرية.

طرق البحث: أجريت الدراسة في المختبر المركزي للمديرية العامة لصحة محافظة دهوك، وتم البحث على عينات نسيجية (منظارية أو مستئصلة جراحياً) مأخوذة من مائة وواحد شخصاً مصاباً بسرطان الغدي للمعدة والمرئ، كما تم صبغ الشرائح بجهاز الصبغ الكيميائي المناعي الآلي (Ventana Benchmark) باستخدام أجسام مضادة أحادية السلالة لمستقبلات HER2 و p53.

النتائج: أظهرت النتائج الإيجابية للمعاينة الكيميائية المناعية لـ (HER2+2/+3) بنسبة ٢٣.٧% لدى المرضى المصابين بسرطان الغدي للمعدة والمرئ مع ارتفاع ملحوظ لـ (HER2+3) لدى المصابين بأورام المرئ مقارنة بنظائرها في المعدة، والعكس صحيح بالنسبة للمعاينة المناعية (p53) في ٦٢.٤% من الحالات، مع ارتفاع ملحوظ في الأورام الغدية للمعدة مقارنة بالمرئ. وفيما يتعلق بالنوع النسيجي فقد كان HER2 مقتصرًا على النوع المعوي لسرطان المعدة في حين ظهر p53 في كلا النوعين المعوي والمنتشر على الرغم من تغلبه باتجاه النوع المعوي.

الاستنتاج: اقتصرت معاينة الـ HER2 على النوع النسيجي المعوي لسرطان الغدي للمعدة، ولم يكن هناك أي ربط بين HER2 و p53 في هذه الأورام ولاسيما أورام المرئ.