Role of enzymes catalase, peroxidase and amino acid (proline) in *Raphanus sativus* and *Lepidium sativus* in exposure levels different water pollution of ion lead

Husham K. Oudah *, Mokdad M. Jawad, Eman H. Abed

Environment and Water Directorate, Ministry of Science and Technology, Baghdad, Iraq.

Abstract

Radish (*Raphanus sativus* L.) and (*Lepidium sativus*) is commonly grown in urban and suburban areas where the soil may be polluted with heavy metal such as lead. In this study, short exposure of radish and cress plantlets to (0,10,20,30,50)ppm lead in nutrient solution (three months) in growth chamber conditions elicited an antioxidative response, measured in terms of lipid peroxidation, protein and proline accumulation and peroxidase and Catalase activity. Longer exposure to lead when radish and cress was grown outdoors for 90 days in pots filled with field soil with different lead content also resulted in higher lipid peroxidation and proline accumulation and altered protein content and enzyme activity. The results also showed significant decrease antioxidant enzymes activity with the decreased oxidative stress. Also, the antioxidant enzymes activity (CAT and POD) were more activity after 30 days because of resistant for (50 ppm, 30 ppm). Besides, at high level, the response of the antioxidant enzymes activity (CAT and POD) and proline accumulation were similar to that observed in control treatment. The highest proline content (81 and 96 μmole/g) were recorded by control treatment after 30 days radish and cress, respectively. The maximum average increase in peroxidase activity (43.76 and 68.38 unit/ml) were though control level treatment after 30 days, respectively. Maximum results were found in treatments (56.25 and 56.52 unit/ml) after 30 days in radish and cress, respectively.

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al. In 71.85°, the treated with the Al concentrations of 78.3 ppm, 300 ppm, 500 ppm, 780 ppm, and 900 ppm, the accumulation of proline in the leaves increased, as shown in Table 1. The accumulated proline in the leaves was significantly higher than the control, which indicated a significant increase in proline content due to Al stress. 

The enzymatic activities, including superoxide dismutase (SOD), catalase (CAT), and ascorbate peroxidase (APX), were also significantly increased under Al stress. The activities of these enzymes are important in the metabolism of ROS and play a significant role in the regulation of photosynthesis, the synthesis of amino acids, and the synthesis of proline in plants. The increased activity of these enzymes under Al stress may be a result of the induction of these enzymes to protect the plant against the stress of Al. 

The results of the present study indicate that Al stress induces the accumulation of proline and the activity of enzymes involved in the metabolism of ROS, which helps the plant to cope with the stress of Al. The accumulation of proline and the activity of enzymes involved in the metabolism of ROS are important in the regulation of the metabolism of amino acids and in the synthesis of proline in plants. The results of the present study indicate that Al stress induces the accumulation of proline and the activity of enzymes involved in the metabolism of ROS, which helps the plant to cope with the stress of Al. The accumulation of proline and the activity of enzymes involved in the metabolism of ROS are important in the regulation of the metabolism of amino acids and in the synthesis of proline in plants.
and Certification, Ministry of Agriculture. Field experiment was conducted during 2015-2016 in silt loam soil at the research field of the Department of Biology, College of Science, Baghdad University, Baghdad, Iraq. The chemical and physical characteristics of field soil were measured in laboratory of soil department, college of agriculture, Baghdad University. Chemical fertilizers used were urea (46% N) at 200 kg ha⁻¹ and triple super phosphate (46% P₂O₅) at 100 kg ha⁻¹. All phosphorus fertilizer was applied at planting during seedbed preparation, while urea was divided into two equal amounts. The first amount was added during the land preparation prior to planting, the second was added 30 days after sowing (during the early tillering stage) and the final amount was added at panicle initiation. Seeds of radish (R. sativus, L.sativus) were sown in plug plates filled with commercial substrate and grown for 3 months in greenhouse conditions, until the 4–5-leaf stage. The uniform plantlets were selected and uprooted from the substrate with water, causing minimal damage to the roots. After washing thoroughly with running deionized water they were planted on perforated polystyrene fasteners containing Hoagland nutrient solution. The experiment was carried out in three replicates (of four plants each. The pots were kept for 3 months in a growth chamber and rotated there every day.

**Experiment**

A field experiment was conducted during 2015 in silt loam soil at the research field of Baghdad, Iraq. The chemical and physical characteristics of field soil were measured in laboratory of soil department, college of agriculture, University of Baghdad.

**Proline determination**

Free Proline content of R. sativus and L. leaves was determined following the method of [15]. Samples of 0.5 g of fresh weight of leaves from each treatment were homogenized in sulphosalicylic (3% w/v H₂O), then centrifuged at 3000 rpm for 5 minutes. Samples of 2 ml from the supernatant were added to 2 ml of each of ninhydrin and glacial acetic acid and incubated at 100°C for 1 hour in water bath. The reaction was arrested in an iced bath and the chromophore was extracted with 4 ml toluene and its absorbance at 520 nm was determined in spectrophotometer (Varian Australia Pty Ltd). Proline concentration was determined using a calibration standard curve prepared with authentic Proline and calculated its amount on fresh weight basis using the following formula.

\[ \mu \text{moles of Proline/g of fresh material} = \left( \frac{\text{μg Proline/ml} \times \text{ml toluene}}{115.5 \, \mu \text{g/μmoles}} \right) \left( \frac{\text{g sample (0.5)/5}}{\text{DF}} \right) \]

**Catalase Activity (CAT)**

**Determination of Catalase (CAT) Activity**

The activity of catalase was determined according to [16] in 3ml of reaction solution, which contained: 2ml of phosphate buffer pH 7.0 and 0.3ml of hydrogen peroxide solution (3%) in a test tube, then 0.2ml of extract containing enzyme (supernatant) was added. The blank was composed from: 2.3 ml of phosphate buffer pH 7.0 and 0.2 ml of extract containing enzyme (supernatant). After 1min the absorbance was measured at wave length of 240 nm using UV-Vis spectrophotometer for activity test and blank tubes. The activity of catalase was calculated as shown below:

\[ \text{Enzyme activity (unit/ml)} = \frac{\Delta A/\text{min} \times \text{dilution factor (DF)} \times \text{reaction volume (V)}}{\text{Extinction Coefficient (EC)} \times \text{Volume of enzyme (EV)}} \]

\[ \Delta A/\text{min} = \text{absorbance of test tube/minute} \]

\[ \Delta A/\text{min} = \text{absorbance of blank tube/minute} \]

\[ \text{DF} = \text{dilution of supernatant} \]

\[ \text{V} = \text{Volume of reaction} \]

\[ \text{EC} = \text{40} \]

\[ \text{EV} = \text{Volume of enzyme (0.2 ml)} \]

**Peroxidase activity (POD)**

**Determination of peroxidase activity (POD)**

1.5 ml of hydrogen peroxide solution (1.7mM) was mixed with 1.5ml of 4-Aminoantipyrine reagent (2.5mM), then 0.1ml of extract containing enzyme (supernatant) was added to the absorbance which measured at wave length of 510 nm using UV-Vis spectrophotometer. The activity of peroxidase was calculated as shown in below equation:

\[ \text{Enzyme activity (unit/ml)} = \frac{\Delta A/\text{min} \times \text{reaction volume (V)} \times D}{6.58 \times \text{EV}} \]

Where:

\[ \Delta A/\text{min} = \text{The change in absorbance at 510nm/minute} \]
RV= Total volume of reaction mixture (3ml)
D = Enzyme dilution factor
6.58 = Extinction coefficient
EV = Volume of enzyme sample (0.1ml) [17, 18].

Result and discussion

Effect of lead on proline content of leaves of two R. sativus and L. sativus under field condition. Results presented in Figure-1 exhibited that average of proline content was for increased by lead concentration. Exposure of plants to 50ppm water stress led to increased proline content of leaves by 29 and 54 % after 30 days after sowing, respectively. Average of proline content was different among lead concentration treatment. The highest proline content (81 and 96 μmole/g) were recorded by control treatment after 30days, while treatments other recorded the lowest proline content (11.00 and 16.00 μmole/g),after 30 days R. sativus and L. sativus, respectively. However, normal and with adding lead concentration, differential response in terms of leaves proline content has been observed. Minimum proline content (11.00μmole/g) was found in R. sativus after 30 days.

Figure 1- Proline concentration. (μmoles/g) in leaves tissue after 30 days from add different of lead concentration.

Effect of lead on Catalase enzymes activity of leaves.

Figure-2 that, showed a decrease in the Catalase production under control (47.01 and 86.18 unit/ml) after 30 days, respectively. Maximum results were found in control treatments (56.25 and 56.52 unit/ml) after 30 days, respectively. Also, there were significantly increases the Catalase content in control treatment (without lead) interaction with lead concentration after 30 days. Although this minimum was much less that (16.87 unit/ml) when R. sativus and L. sativus treated with lead concentration after 30 days, respectively. Data presented in Figure-2 show the effect lead concentration stress on Catalase activity after 30days of R. sativus and L. sativus plants grown field capacity. Results presented exhibited that average of Catalase activity was for increased by water deficit stress. Exposure of plants to 50 ppm lead led to increased Catalase activity in leaves by (39.65 and 39.38%) after 30 days after R. sativus and L. sativus, respectively.
Figure 2- Catalase enzymes activity (unit/ml) in leaves tissue after 30 days from add different of lead concentration.

Effect of lead on peroxidase enzymes activity of leaves.

The peroxidase activity Figure-3 at 30 days maximum average increase by (21.47% and 46.09 %) in peroxidase activity R. sativus and L. sativus, respectively. The activity of this antioxidant enzyme again increased within lead concentration treatments. The maximum average increase in peroxidase activity (43.76 and 68.38 unit/ml) were though control level treatment, while minimum (24.07 and 27.35 unit/ml) were recorded in lead concentration treatments after 30 days, respectively.

Figure 3- Peroxidase enzymes activity (unit/ml) in leaves tissue after 30days from add different of lead concentration.

Peroxidase are widely distributed in living organisms including microorganisms, plants and animals. POD is mainly located in the cell wall [19] and it is one of the key enzymes controlling plant growth and development. It takes place in various cellular processes including construction, rigidification and eventual lignifications of cell walls [20].Removal of two major systems for the enzymatic removal of H₂O₂ and peroxidative damage of cell H₂O₂ is therefore a protective mechanism for the preservation of biological membranes when lead and other metals accumulate in the symplast of the cell [21], proline is probably the most widespread, and is considered to be an indicator of
environmental stress [22]. Accumulation of proline has been shown to protect plants against damage by ROS (Reactive oxygen species), acting as a very effective singlet-oxygen quencher, binding to redox-active metal ions and also activating and protecting enzymes such catalase, peroxidase and polyphenol oxidase [23]. POD and Catalase are walls is controlled by the potency of antioxidative peroxidase enzyme system [24]. However, the role that peroxidase plays in metabolism is not clear because of the large number of reactions it catalyzes and the considerable number of isoenzymic species [25]. It was reported that peroxidase had been used for biotransformation of organic molecules [26–28]. Because of its broader catalytic activity, a wide range of chemicals can be modified using POD. Also, it can be used for the applications such as synthesis of various aromatic compounds, removal of phenolic from waste waters and the removal of peroxides from foodstuffs, beverages and industrial wastes [29]. POD is also related to quality of plant commodities, particularly the flavor, in both raw and processed foods. POD activity is also correlated to fruit ripening as shown in a number of cases and it is also involved in enzymatic browning, either or together with polyphenol oxidase activity. A more precise understanding of the implication of POD in these mechanisms is an essential step towards a more efficient control of these undesirable reactions, particularly in heat-processed products, which frequently contain residual peroxidase activity [30–31]. Adaptation of plants to grow in different habitats requires specific abilities that differ among plant [32] and reduces plant performance through damaging plant parts and cell components such as cell membranes, proteins, lipids, pigments and DNA expression [33, 34]. Plants in order to accommodate more tolerance against environmental stresses have developed efficient physiological and biochemical enzymatic response mechanisms such as production of superoxide dismutase, Catalase and peroxidase, and non-enzymatic antioxidant compounds such as phenolic compounds and flavonoids to rid themselves of free radicals [35–42]. Thirty four species of Artemisia (with English names worm wood and sage brush) are the main and most common perennial species in steppic and semi-steppic ecosystems of Iran. Due to their distinctive features, Artemisia plants are highly resistant against extreme environmental conditions and very effective in stabilizing the habitat; have great forage value, are medicinal and exhibit strong antioxidant property via their phenolic compounds and have conservation and aesthetic values. Also both mechanism CAT and POD breakdown free radical release water and oxygen, proline of accumulation in oxidative stress [43, 44].

References


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