



ISSN: 0067-2904

GIF: 0.851

Electrochemical Study for the Effect of Simvastatin and Atorvastatin on Coenzyme Q₁₀ by Differential Pulse Polarography Using Mercury Drop Electrode

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Abstracts

Statin drugs act by inhibiting the enzyme HMG-CoA reductase which is responsible for manufacture of cholesterol and the biosynthesis essential energy production Co-factor (Coenzyme Q₁₀). The aim from this research includes study the effect of statin drugs (Simvastatin and Atorvastatin) on the Coenzyme Q₁₀ using differential pulse polarographic technique at a dropping mercury electrode (DME) and in a mixture of [4:1] methanol-phosphate buffer of pH7.0 as supporting an electrolyte. Prior to this, the behaviors of Simvastatin, Atorvastatin and Coenzyme Q₁₀ were studied separately in their solvents. The half-wave potential ($E_{1/2}$) of Co Q₁₀ were -0.31volt and -1.37Volt, -1.33 Volt for simvastatin and atorvastatin respectively. A mixture of Coenzyme Q₁₀ with Simvastatin and Atorvastatin in the same solvent shows a shift in their peak potential (E_p) toward more negative potentials values by (-0.06volt) for Simvastatin and by (-0.09volt) by Atorvastatin.

Keywords: Statin Drugs, Coenzyme Q₁₀, differential pulse polarography.

دراسة كهروكيميائية لتأثير السيمفاستاتين والأتورفاستاتين مع مساعد الانزيم Q₁₀ بواسطة البولاروغرافيا النبضية المشتقة باستخدام قطب الزئبق المتقاطر

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قسم الكيمياء، كلية العلوم، جامعة بغداد، بغداد، العراق.

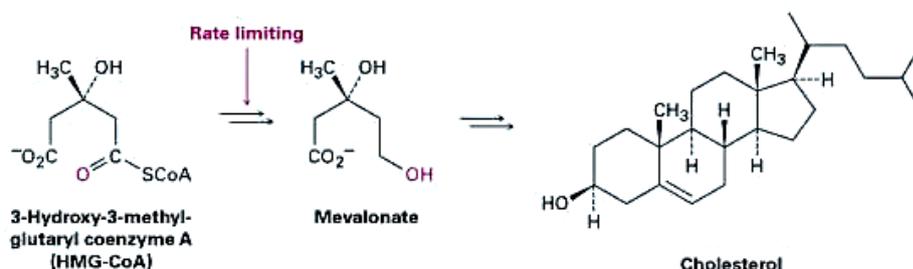
الخلاصة

تعمل ادوية الستاتينات على تثبيط انزيم HMG-CoA reductase وهو الانزيم المسؤول عن تصنيع الكوليسترول والتركيب الحيوي لأنتاج الطاقة Coenzyme Q₁₀. ويتضمن هذا البحث دراسة تأثير ادوية الستاتينات (السيمفاستاتين و الأتورفاستاتين) على مساعد الانزيم Q₁₀ باستخدام تقنية البولاروغرافيا النبضية المشتقة عند قطب الزئبق المتقاطر والمذاب كل منهما في مزيج من الميثانول والمحلول المنظم الفوسفاتي pH7.0 كإلكتروليت ساند بنسبة [1:4]. وقيل دراسة التأثير، درس سلوك كل من ادوية الستاتينات ومساعد الانزيم Q₁₀ بصورة منفصلة في محاليلهما وتم تعيين جهد القمة لمساعد الانزيم وكانت بقيمة -0.31Volt و بقيمة -1.37Volt، -1.33Volt لكل من السيمفاستاتين والأتورفاستاتين. وتبين ان خليط ادوية الستاتينات مع مساعد الانزيم Q₁₀ في نفس المذيب ادى الى ازاحة في قيم جهد القمة لمساعد الانزيم Q₁₀ نحو قيم اكثر سالبية وكانت بمقدار -0.06Volt بالنسبة للسيمفاستاتين و بمقدار -0.09Volt بالنسبة للأتورفاستاتين.

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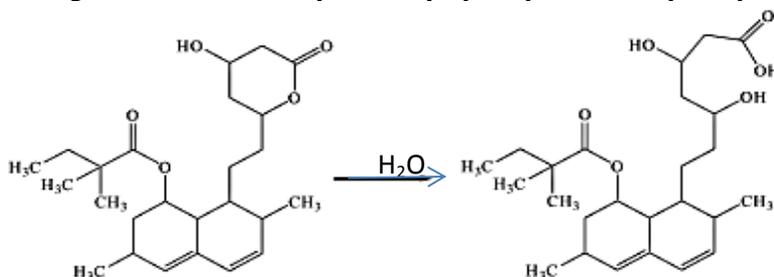
Introduction

Simvastatin and atorvastatin are examples of statin drugs; statins are the most commonly used and the most effective class of drugs for lowering of plasma cholesterol [1]. Statins reduce cholesterol in the blood by block an enzyme in the liver that synthesizes cholesterol and prevent the liver from producing and releasing cholesterol in the blood stream [2]. The principle action of statins is to inhibit cholesterol synthesis by inhibiting 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase, an enzyme that controls the rate of cholesterol synthesis in the liver via the mevalonate pathway, it is an enzyme that catalyzes the conversion of HMG-CoA to mevalonate, which through several further biochemical steps is metabolized to cholesterol which takes place in the liver as shown in scheme -1 [1, 3].



Scheme 1- Mavalonat pathway [4].

Simvastatin (1S,3R,7S,8S,8Ar8-{2-[92R,4R)-4-hydroxy-6-oxooxan-2yl]ethetyl}-3,7-dimethyl 1,2,3,7,8a-hexahydronaphalen-1-yl 2,2-dimethylbutanoate, belongs to the first generation of statins (Type I statins). They are all derived directly or from fungal metabolites. And share a similar structure which contains a (polar head group) and a hydrophobic moiety which includes a decalin ring [5, 6]. The chemical structure of simvastatin contains a β -hydroxy-lactone. Physiologically active form of the drug is the β -hydroxy acid, which is formed by a ring-opening reaction of lactone ring. In vivo prodrug lactone form is enzymatically hydrolyzed to its hydroxyl-acid pharmacophore scheme-2 [7].



Scheme 2- Molecular structures of simvastatin and physiologically simvastatin β -hydroxy acid form.

Atorvastatin is a (bR,dR)-2-(r-fluorophenyl)-b,d-dihydroxy-5-isopropyl-3-phenyl-4(phenylcarbamoyl) - pyrrole-1-hepatanoicacid (1:2) trihydrate, belongs to the type II statin. There are synthetic structures contain a different and large hydrophobic moiety. The synthetic statins have in common a fluorinated phenyl group, and a base structure made up of a five or six member ring wione atom substituted with a nitrogen atom Figure- [6, 8].

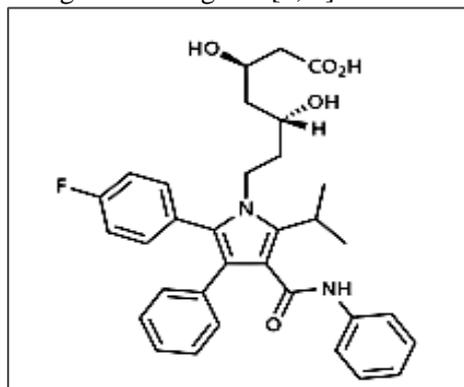


Figure 1-Chemical structure of atorvastatin

Statins produce both therapeutic effects and side effects through the same mechanism, HMG-CoA reductase is at the base of mevalonate pathway, the mevalonate pathway is involved not only in the biosynthesis of cholesterol but also in the biosynthesis of essential co-factor required for energy production Coenzyme Q₁₀, thus statins block the endogenous biosynthesis of both cholesterol and Coenzyme Q₁₀ by inhibiting the enzyme HMG-CoA reductase [9, 10].

Coenzyme Q₁₀ is a fat soluble, synthesized by human body. It was first isolated from beef heart mitochondria in 1957 by Frederick Crane. It is a Quinone with an isoprenoid side chain [11]. The primary function of Coenzyme Q₁₀ is an indispensable cofactor in the electron transport of mitochondria, function as an electron carrier between the enzyme complexes of the respiratory chain. It is also a lipid-soluble antioxidant that scavenges lipid radicals within biological membranes [12]. The aim of this work is the study the effect of statin drugs on the peak potential of Coenzyme Q₁₀ by the application of differential pulse polarographic technique.

Experimental

Chemicals and Reagents

- Simvastatin and Atorvastatin in pure form were supplied by Drug Control Department for Iraq Ministry of Health.
- Phosphate buffer solution pH7.0 (K₂HPO₄·2H₂O and NaHPO₄·H₂O) was supplied from Radiometer Copenhagen.
- Ubiquinone (Coenzyme Q₁₀) from Fluka product of USA.
- Absolute Methanol with purity 99% from the Merck Chemical Company.

Because of the low solubility of statin drugs and CoQ₁₀ in water, polarographic measurements were done in a mixture of Methanol and phosphate buffer.

Simvastatin (5×10⁻⁴M) was prepared by dissolved (0.0209gm) in (80 ml) methanol and (20 ml) phosphate buffer pH7.0. Atorvastatin (5×10⁻⁴M) was prepared by dissolved (0.0279gm) in (80 ml) methanol and (20 ml) phosphate buffer pH7.0.

A stock standard solution of Coenzyme Q₁₀ (3×10⁻⁵M) was prepared by dissolved (0.0259gm) in methanol. A working solution of Coenzyme Q₁₀ (10⁻⁵M) was prepared by dissolving (33.3ml) from stock solution of Coenzyme Q₁₀ and complete to the 100ml by adding methanol and 20 ml of phosphate buffer pH7.0. All solutions were prepared freshly just before use.

Instrumentation

Polarographic Analyzer model 174A was used for polarographic measurement. It was coupled to an (Kompenson X-Y Recorder model C1924) the supplier of the company Siemens. The Polarographic cell composed of three electrodes, the mercury drop electrode, which is normally a cathode of the polarographic cell flows from a glass capillary tube, a saturated calomel reference electrode and the counter electrode (Mercury pool) into which the drop falls, acts as an anode.

Procedure

A mixture of (1:1) proportion of statin drugs with Coenzyme Q₁₀ were placed in the polarographic cell. Then recording the polarographic curve, the solution was deoxygenated by purging with purified nitrogen through the solution for several minutes. The instrumental setting for the DDP was as follows: Potential scan rate 10 mv/SEC, Drop time 0.5, Sec, Modulation Amplitude 50 mV, current Range 0.5 mA. All measurements were carried out at room temperature.

Results and Discussion:

Prior to the study of the interaction of statin drugs with CoQ₁₀, has been studied the electrochemistry reduction and behavior of both statin drugs and Coenzyme Q₁₀ separately, each dissolved in a mixture of methanol and the solution of phosphate buffer pH7 as a supporting electrolyte with ratio 4:1. These values measured against the reference calomel electrode.

Simvastatin gave two reduction waves, the first wave at (-0.4 volt) is an adsorption wave, and the second wave at (-1.37 volt) refers to the reduction wave as shown in Figure-2. That is in good agreement with previous polarographic investigation [13].

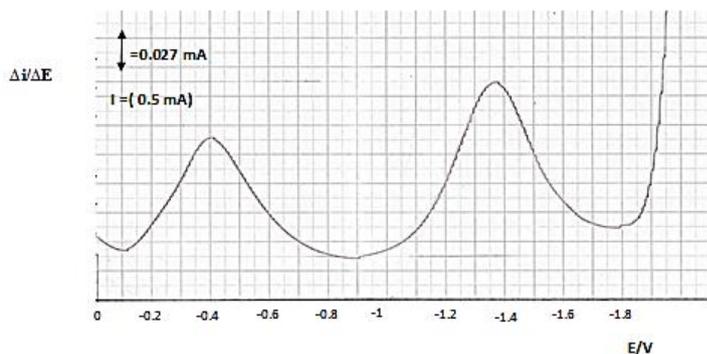


Figure 2-Polargram of (5×10^{-4} M) simvastatin at drop mercury electrode in a mixture of (4:1) methanol and phosphate buffer pH 7.

Atorvastatin also gave two reduction waves, the first wave at (-0.43Volt) is an adsorption wave, the second wave at (-1.33Volt) refers to the reduction wave as shown in Figure 3. That is in good agreement with previous polarographic investigation [8].

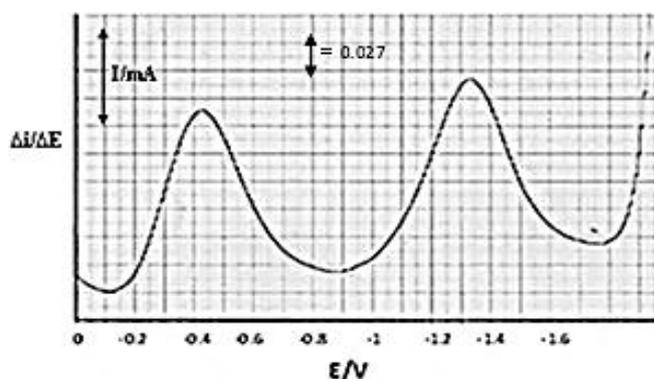


Figure 3-Polargram of (5×10^{-4} M) Atorvastatin at drop mercury electrode in a mixture of (4:1) methanol and phosphate buffer pH 7.0

The dropping mercury electrode may adsorb the electroactive species or the product of the electrode reaction, this adsorption is due to surface forces, the particles of the dissolved substance may be bound to the surface by physical, in this case a separate wave, the adsorption wave is formed [14]. In figure (-4 and -5) these polarograms show two waves. An adsorption prewave is observed if the electrolysis product is adsorbed and causes the electrode reaction to occur more easily [15]. And been provable, it was found that when reducing the height of mercury head, decreased the value of the limiting adsorption current. $I_a = \text{const } h$. The limiting adsorption current dependence on the height of the mercury head [14]. Electrochemistry reduction of Coenzyme Q_{10} were done in (4:1) methanol and in the solution of phosphate buffer pH 7.0 as a supporting electrolyte, it is reduced in two successive one-electron reduction steps which show two waves at a potential value in the range of (-0.31-1.27 volt) as shown in Figure-4. That is in good agreement with previous polarographic investigation [16].

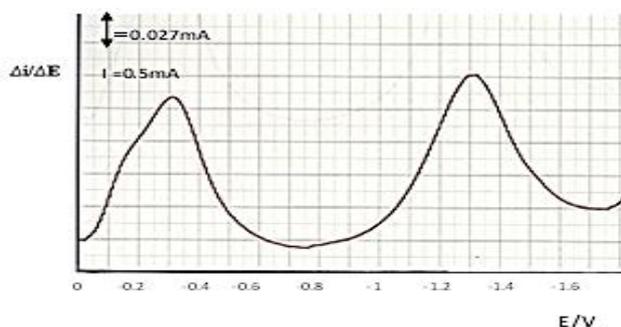
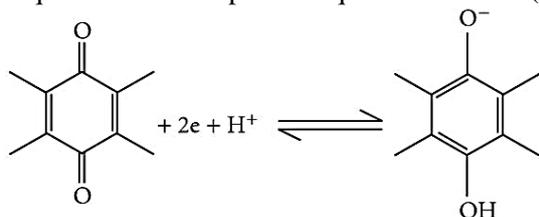


Figure 4-Polarographic waves of 10^{-5} M Coenzyme Q at drop mercury electrode in (4:1) methanol and phosphate buffer pH 7.0

The first wave at -0.31V is completely reversible one proton one electron leads to produce semiquinone $Q^{\cdot-}$ (quinone anion radical) and the second step at -1.3V is irreversible, the reoxidation of semiquinone leads to produce quinone dianion (Q^{2-}) as shown in scheme-3.



Scheme 3-Two- electron one proton reduction of Coenzyme Q_{10} in aqueous buffer.

The interaction of drugs with the enzyme assistant Q_{10} were studied using high concentration of drugs much higher than the assistant enzyme Q_{10} concentration to ensure occurrence of interaction between them and deviation of balance towards the formation of molecular complexes. Due to the significant self-association of the Coenzyme Q_{10} , it was found necessary to study their interaction under dilute concentration, ($< 10^{-5}$ mol/L), in order to avoid any interference with the complication process. The interaction between the Coenzyme Q_{10} and statin drugs was followed: through the recording a polarographic curves for the mixture in period of time, it took three to four hours to obtain an equilibrium state which were noted by the fixed values of $E_{1/2}$ and I_p . The results of a polarographic measurements obtained were shown in table (-1 and -2) and Figure (-5 and -6).

Table 1-peak potential (E_p) measured in Volt, diffusion current (i_d) measured in milliamper for the reduce of mixture CoQ_{10} concentration ($10^{-5}M$) with Simvastatin ratio (1:1) and values change in peak potential ($\Delta E_{1/2}$) to reduce the CoQ_{10} in Phosphate buffer solution pH7.0 at DME.

Substance	E_p Volt	ΔE_p Volt	I_d (mA)	T(min)
Coenzyme Q_{10} ($10^{-5}M$)	-0.31	000	0.12	00
Coenzyme Q_{10} + Simvastatin in phosphate buffer pH7.	-0.33	-0.02	0.08	30
	-0.34	-0.03	0.09	60
	-0.35	-0.04	0.13	90
	-0.36	-0.05	0.14	120
	-0.37	-0.06	0.20	150

Table 2-peak potential (E_p) measured in Volt diffusion current (i_d) measured in milliamp and reduce mixture CoQ_{10} concentration (10^{-5}) with Atorvastatin with ratio 1:1 and values change in the peak potential (ΔE_p) to reduce the CoQ_{10} in Phosphate buffersolution pH7.0.

Substance	E_p Volt	ΔE_p Volt	I_d (mA)	T(min)
Coenzyme Q_{10} ($10^{-5}M$)	0.31	000	0.12	
Coenzyme Q_{10} + Atorvastatin in phosphate buffer pH7.0	0.35	0.04	0.08	30
	0.36	0.05	0.09	60
	0.37	0.06	0.11	90
	0.38	0.07	0.16	120
	0.39	0.08	0.18	150
	0.4	0.09	0.23	180

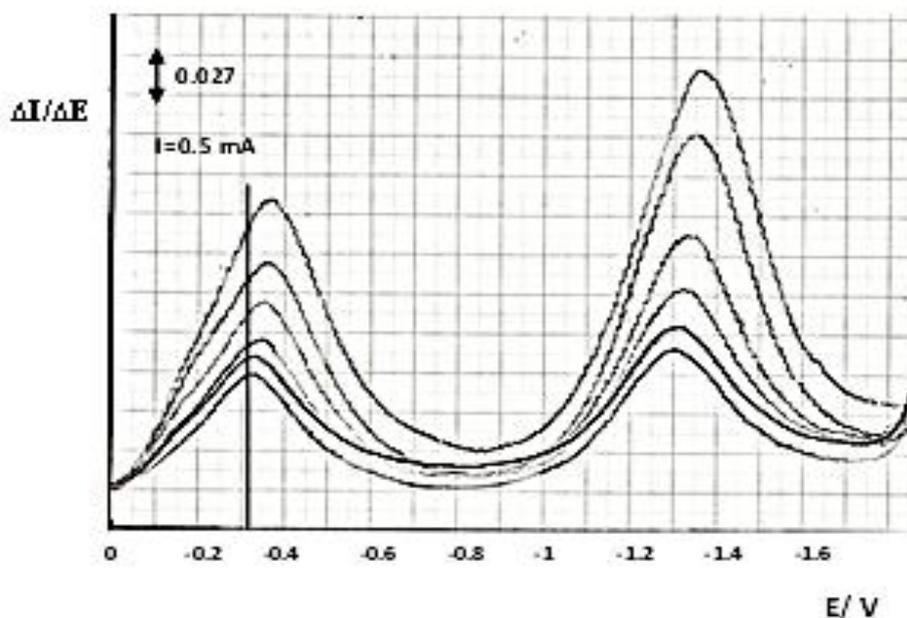


Figure 5-Polarogram for mixture Coenzyme Q₁₀ with simvastatin illustrating the change values peak potential (E_p) and diffusion current for Coenzyme Q₁₀ by adding simvastatin at different time.

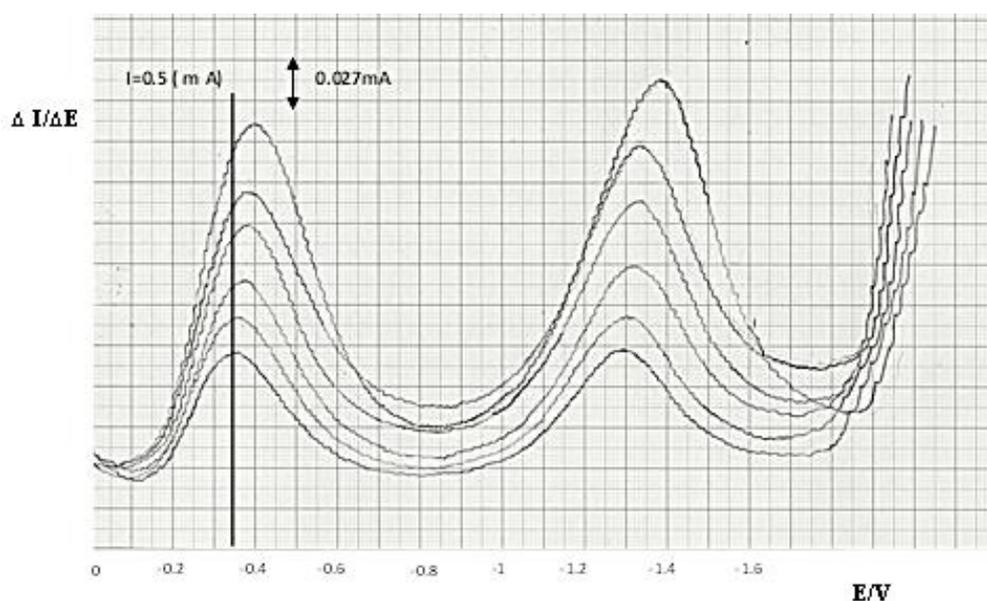


Figure 6-Polarogram for mixture Coenzyme Q₁₀ with atorvastatin illustrating the change values peak potential (E_p) and diffusion current for Coenzyme Q₁₀ by adding simvastatin at different time.

From results a polarographic measurement in table 1 and 2 and figure the following observation were obtained.

1. The addition of statin drugs to Coenzyme Q₁₀ cause a shift in E_p value to a more negative potential due to the formation of molecular complex, this will block the role of Coenzyme Q₁₀ as an electron carrier in the respiratory chain.
2. This interact between the statin drugs and Coenzyme Q₁₀ raise the energy level of the lowest unoccupied molecular orbital of Coenzyme Q₁₀ (LUMO) which lead to an increase in its redact potential of this results agreed with other previous work of Coenzyme Q₁₀ with antimalarial drugs [17].

A number of papers have been published, in which the relation between half-wave potential and the extent of conjugation in the system is treated in terms of quantum mechanics. Thus, it was

found that there is an excellent correlation between the half-wave potentials of hydrocarbons and the energy of their *lowest molecular orbitals* LUMO [14].

3. The difference in the ability of Simvastatin and Atorvastatin to form complex with Coenzyme Q₁₀ were note also from the difference in ΔE_p cause by their interactions which illustrated in table 1 and 2. This is due to the difference in their functional groups which may participate in the formation of their molecular complexes with Coenzyme Q₁₀.
4. diffusion current of the mixture of Coenzyme Q₁₀ and statin drugs decrease at first due to their interact and the formation of a molecular complex which illustrating that this formation is very fast then id increase again with time may be due to a rearrangement of two molecules to form complex which have a higher diffusion coefficient than the parent molecular.
5. theoretical calculation: the second order perturbation theory and Huckel molecular orbitals were used to calculate the interaction energy of statin drugs and Coenzyme Q₁₀.

This interaction increase the energy level of the lowest unoccupied molecular orbital of Coenzyme Q₁₀.

Kinetic Calculation

Calculated rate constant from first and second order for interaction of Coenzyme Q₁₀ with atorvastatin and simvastatin by tracking the increase in a diffusion current of Coenzyme Q₁₀ with statin drugs with time.

Interaction equation from first order:

I = diffusion current at t .

I_0 = diffusion current at $t = 0$

k = Rate constant

Rate constant (k) has been calculated from mile straight-line equal to $\left(\frac{-k}{2.303}\right)$ the output of the draw (Log i) against time shown in Figure -7, -8.

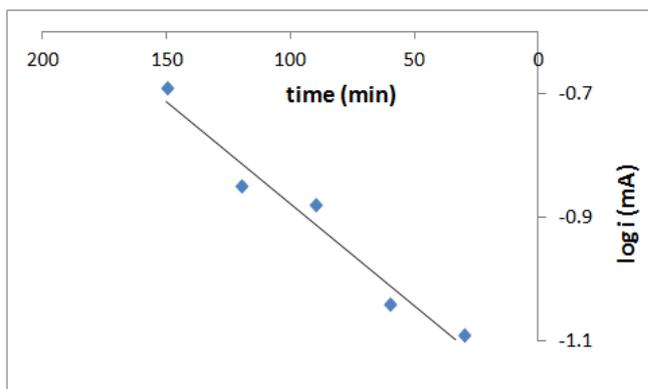


Figure 7-Determenation of rate constant from first order for interaction of Coenzyme Q₁₀ with simvastatin in (1:1) proportion using differential pulse polarography.

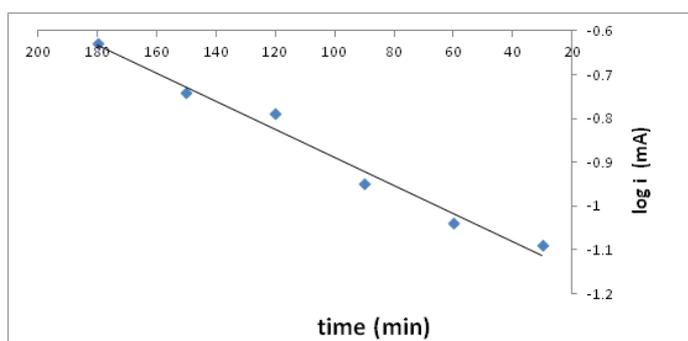


Figure 8-Determination of rate constant from first order for interaction of Coenzyme Q₁₀ with atorvastatin in (1:1) proportion.

Illustrates Table-3 values constant rate interaction from first order for Coenzyme Q₁₀ with simvastatin and atorvastatin in 1:1.

Table 3-Values rate constant from first order for interaction of Coenzyme Q₁₀ (10⁻⁵) with simvastatin and atorvastatin using differential pulse polarography.

Coenzyme Q ₁₀ + ...	K(min ⁻¹)
Simvastatin	7.59 × 10 ⁻³
Atorvastatin	7.36 × 10 ⁻³

The straighter line graph and values of rate constant prove the interaction of coenzyme Q₁₀ with simvastatin and atorvastatin corresponds to conditions kinetics of first order interaction.

Calculated rate constant interaction from second order for Coenzyme Q₁₀ with atorvastatin and simvastatin from equation as followed:

$$t = \frac{x}{ta(a - x)}$$

The relationship between time (min) and $\left(\frac{x}{a-x}\right)$ equivalent to $\left(\frac{id-i}{i}\right)$ was plotted as shown in Figure - 9,-10.

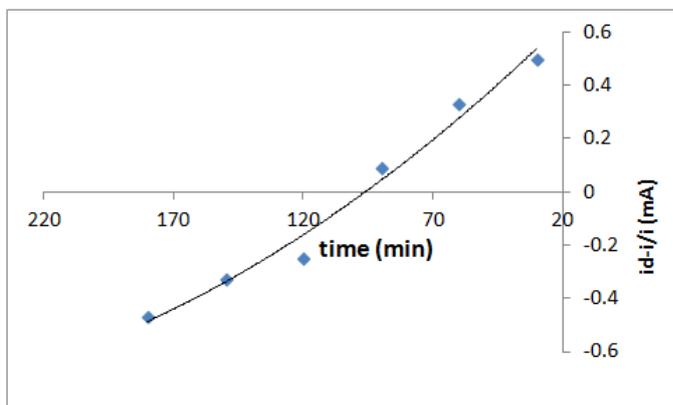


Figure 9-The application of the equation interaction from second order for interaction Coenzyme Q₁₀ with simvastatin.

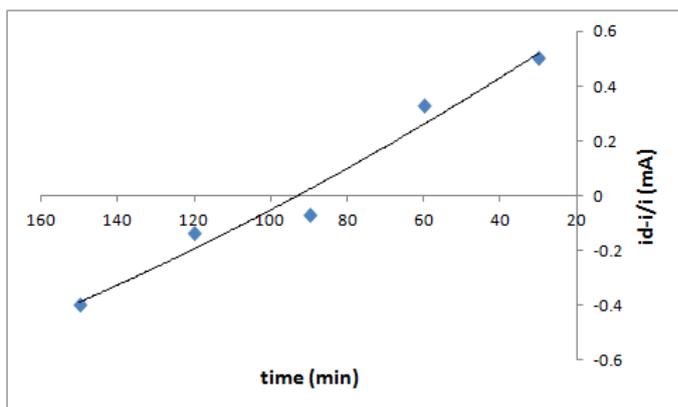


Figure 10-The application of the equation Interaction from second order for interaction Coenzyme Q₁₀ with atorvastatin.

Figure -9 and -10 were shown the relationship non-line and no compatible with kinetics second order. Was calculated free energy of activation from the following the relationship

$$K_1 = \frac{kT}{h} e^{-\Delta G^\ddagger/RT}$$

K= rate constant (min^{-1})

k= Boltzmann's constant $1.38 \times 10^{-23} \text{J.k}^{-1}$

h= Planck's constant $1.38 \times 10^{-34} \text{JS}$.

R= Constant gas $8.3 \text{ mol}^{-1} \text{k}^{-1} \text{J}$

ΔG = free energy of activation $\text{mol}^{-1} \text{.KJ}$.

Results have been installed in Table-4.

Table 4-Values free energy of activation for interaction of Coenzyme Q₁₀ with simvastatin and atorvastatin.

Coenzyme Q10 +....	$\Delta G (\text{KJ.mol}^{-1})$
Simvastatin	41.23
Atorvastatin	41.67

Conclusions

The addition of statin drugs to Coenzyme Q₁₀ cause a shift in *peak potential* (E_p) of Coenzyme Q₁₀ toward more negative value as a result of interaction. The shift was attributed to the formation of *molecular complexes*, thus cut off the flow of electrons and led to disabling to work within the respiratory chain as a carrier electrons. Simvastatin and Atorvastatin differ in their ability to form complexes with CoQ₁₀, it was found that Atorvastatin show a higher shift in E_p values than Simvastatin because of their structure characteristics, Atorvastatin have additional hydrogen binding and polar interactions. Also observed an increase in diffusion current of Coenzyme Q₁₀ due to high diffusion of the interacting molecules.

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