

**Detection Of Pseudomonas Aeruginosa Isolated From Burn  
Patients At The Teaching Hospital In The-Qar Province  
And Its Resistance To Antibiotics**

**Hamed Maged Mostafa**

Technical Institute / DhiQar-Shatrah

Southern University / Basra

**Abstract**

The current study included the collection of (110) of Clinical samples *Pseudomonas-aeruginosa* from models bacteria from Sarsary models for burn patients in Thi-Qar Governorate for the period (1/12/2016 until 1/7/2017). The diagnosis of bacteria was based on plant traits, microscopic examination and growth on the differential and standard biochemical tests. The method of spreading the tablets and measuring the inhibition areas around the disk were used to test antibiotic resistance, (16) isolates were shown to be resistant antibiotics resistance to both antibiotics (Levofloxacin 90%, Amikacin 80%, Aztraonam 75%, Ampicillin 95%, Carbenicillin 80%, Ciprofloxacin 90%, Gantamicin 75%, Cftazillin 70%, Tetracycline 55%, Imipenem 30%)

**Key words :-** Pseudomonas aeruginosa , Burn patients, Antibiotics

## الكشف عن بكتريا الزائفة الزنجارية *Pseudomonas aeruginosa* المعزولة من مرضى الحروق في مستشفى التعليمي في محافظة ذي قار ومقاومتها للمضادات الحيوية

همسة ماجد مصطفى

المعهد التقني/ذي قار-الشطرة

الجامعة الجنوبية/البصرة

### الخلاصة

تضمنت الدراسة الحالية جمع (110) عينة لجرثومة الزائفة الزنجارية من نماذج سريرية لمرضى الحروق في محافظة ذي قار للمدة (١/١٢/٢٠١٦ لغاية ١/٧/٢٠١٧). اعتمد تشخيص البكتريا على الصفات الزرعوية والفحص المجهرى والنمو على الاوساط التفرقية والاختبارات الكيموحيوية القياسية استخدمت طريقة الانتشار بالأقراص وقياس مناطق التثبيط حول القرص لاختبار المقاومة المضادات الحيوية أظهرت (١٦) عزلة مقاومة لكل من المضادات الحيوية (Levofloxacin بنسبة 90%, Amikacin بنسبة 80%, Aztraonam بنسبة 75%, Ampicillin بنسبة 95% Carbenicillin بنسبة 80%, Ciprofloxacin بنسبة 90%, Gantamicin بنسبة 75% Cftazillin بنسبة 70% Tetracyclin بنسبة 55%, اما مضاد Imipenem بلغت المقاومة 30%)

الكلمات الدالة:- الزائفة الزنجارية , مرضى الحروق, المضادات الحيوية

## **Introduction**

*Pseudomonas aeruginosa* is a bacillus that is found in single cells or in an air chain (Brooks et al., 2004) and is characterized by its ability to live at (42) ° C and an ideal degree of (37) ° C (Toder, 2004). *Pseudomonas aeruginosa* is a widespread bacteria found in soil and water and is found on the skin surface of humans and animals and on the surface of plants (Cornelis, 2008). *Pseudomonas aeruginosa* is an opportunistic bacterium with low immunity caused by bacteremia, septicemia, skin infection and injury to burns and burns, a cause of the burns that have caused the deaths of many patients with septicemia. The risk of these bacteria due to the widespread in hospitals and the surrounding areas, causing many diseases, which caused (10-120) of hospital-acquired diseases and post-traumatic diseases traction. The burned areas are a sensitive location for the invasion of opportunistic germs due to the loss of natural protection provided by the skin tissue and the weakness of the immune system due to injury. Dead tissue is a suitable medium for growth and reproduction of germs, (Future, 2008). Bacterial resistance to antibiotics is due to several genetic factors. (Storz&Aronis 2000, Wickens& Wade, 2005). Increased resistance to bacterial strains has led to an increase in disease, mortality, and many infections that have been responsive to antibiotics (Blaser et al., 1995), which can no longer be easily overcome. Due to the abuse and excessive use of antibiotics (Nicolao and Budi 2002), the emergence of bacterial strains is not affected by antibiotics, especially hospital patients (Levy, 1999).

## **The aim**

- 1-Isolation of *Pseudomonas aeruginosa* bacteria from burns to compare disease characteristics
- 2-Testing the extent of resistance to isolates studied antibiotic

## **Material and method**

- 1-Collection of samples

A total of 110 samples of burn patients were collected at Al-Hussein Technical Hospital in Thi-Qar province (1/12/2016 - 1/7/2017). Swabs were taken with wound swab for the purpose of isolating and diagnosing *Pseudomonas aeruginosa*. Plantation and Biochemistry (Mucfadge, 2000)

## **2-Inoculating**

Samples were inoculated on the (Blood Agar, Nutrient agar, MacConkey agar) to study the phenotypic characteristics of the colonies and to obtain single colonies for diagnosis.

### 3- Lab Diagnosis

Diagnosis based on phenotypic traits and biochemical tests based on diagnostic sources (Forbes, 2007) and diagnosis using API20 NE.

#### 4- Susceptibility Antibiotic test

All isolates of antibiotics were tested according to the Bauer and Kirby method (1996). In this (12) antimicrobial agents, as shown in Table( 1.1). *Pseudomonas aeruginosa* for each strain (18) hours to the middle surface (MHA) and sprayed well Swab and left a quarter of an hour to dry the surface and distributed on the surface of the pollinated medium with antibiotics.

**Table (1-1) Percentage of antibiotic resistance for isolates**

antibiotic	Percentage of resistance%
Ampicillon	95
Levofloxacin	90
Gantamicin	75
Amikacin	80
Ciproflaxacin	90
Ceftazilim	70
Aztereonam	75
Tetracyclin	55
Carbanem	80
Imipenem	30

### Results and discussion

A total of (110) clinical samples were collected from burn patients in Al Hussein Educational Hospital in The-Qarprovence(1/12/2016 - 1/7/2017). shows that the percentage the rate of infection in females is higher (84.45%) than that of males(17.45%) as shown in table (1-2), may be due to the fact that these bacteria are widespread in the soil and water in addition to being the bacteria spread in hospitals, which cause infections for patients Suffer from a lack of body defenses in addition to patients who have been in hospital for more than a week. On the other hand, these types of opportunistic germs, which need nutrients are simple to grow and it is characterized by resistance to disinfectants and

antibiotics and possess many factors of ferocity.

A total of (16) samples of *Pseudomonas aeruginosa* (19.59), (73) of different types of bacteria were obtained (66.36%), and there was no bacterial infection z (21) with (54% - 14%) without any type of bacteria

Isolation (16) was tested for bacteria *Pseudomonas aeruginosa* were selected for (10) antibiotic antibiotics using the Kirby and Bauer method. Bacteria resistance was determined by the measurement of diameters around the disc (Benson, 2002). The results were compared with the (CLSI, 2012) as shown in Table (1-1).

Bacteria are resistant to Ampicillin (95%) and are not produced by  $\beta$ -lactamase enzymes. This enzyme leads to resistance to the plasmid or chromosome (Cole, 1986). These results were not consistent with (Zine El Abidine, 2015). Gentamycin (75%) was antagonistic to Aminoglycoside, which causes a decrease in membrane permeability. This result is not consistent with the results (Al-Rawi, 1999).

)70% .(This antibody belongs to the cephalosporin group. Resistance may be due to the frequent use of the treatment and the results are consistent with ( Kazem 2010)

The anti-Ciprofloxacin (90%) is a quinolone that is lethal to microorganisms and inhibits the building of DNA or inhibiting the DNA gyrase enzyme as it leads to rapid death (Hardy, 2000). These bacteria have a wide spectrum against negative and gram (Katzung, 2004). These findings were agreed with (Jawad ,2011)

While the resistance percentage of the antibiotic return Amikacin (80%) This antibiotic is due to the aminoglycoside group, which is a killer of microorganisms The bacterial protein is inhibited by binding to the unit (Katzung, 2004). These results were not consistent with (Uribe ,2010)

The isolates showed under study low resistance to counter the direction Imipenem reached (30%) antibiotic half an artificial from the group Carbapenim produced by the bacterium *Streptomyces cattleya*, a broad spectrum against bacteria positive and negative gram which is fixed does not degrade the enzymes  $\beta$ -lactamases produced by most types of bacteria that It is also effective against negative bacteria gram (Buckley et al., 1992) and agreed with these results (Abidin, 2005).

Table (1-2) percentage of pathogens for patients

Age group	Male	%	Female	%
15-20years	3	17.6	9	9.6
21-25years	5	29.4	20	21.5
26-30years	6	35.29	36	38.7
31-35years	3	17.6	28	30.1
Total	17	15.45	93	84.45

#### المصادر

- 1- سلمان زين العابدين , زهراء عايد احمد. (٢٠١٥). التحري عن الانزيمات البييتالاكتاميزالمستحثه في بكتريا (*pseudomonasauroginosa*) المعزوله من نماذج مرضيه. رسالة ماجستير . كلية العلوم . جامعة كركوك
- 2- الراوي, ندى فاضل (١٩٩٩). دارسه تشخيصيه وفسلجيه لعدد من الاجناس التابعة لمجموعه الجراثيم العضوية غير مخمره . أطروحة دكتوراه , كلية العلوم , جامعة الموصل
- 3- , كاظم , الهام جواد (٢٠١٠). التحري عن انزيمات (AMPC) البييتالاكتاميز في العزلالتسريرية لبكتريا الزنجاريه (*pseudomonasauroginosa*) في مستشفيات مدينة النجف . مجلة جامعة الكوفة . كلية الطب / جامعة الكوفة
- 4- عريبي, سناء مهدي (٢٠١٠). مكافحة جرثومة (*pseudomonasauroginosa*) في إصابات سريريه مختلفة لبعض المضادات الحيوية . مجلة علوم ذي قار . مجلد ح العدد ٤
- 5- ب,ستورت , ليفي (1999).تحديات مقاومة البكتريا للمضادات الحيوية ,مجلة العلوم (15),المجلد31,العدد 10,ص16,الكويت
- ٦- نيكولاوس.ك - بوديس.ن. (2002) . خلفخطوطالعدو،مجلةالعلوم،المجلد 18 ، العدد 4
- 39،مجلةالعلوم،الكويت ص32
- ٧- صبا جاسم جواد (2011) دراسة بكتريولوجي للمصابين بالتهاب الاذن الوسطى ,مجلة العلوم , Vol:7NO:2
- 8- Brooks ,G.F. , Carroll, K.C. , Butel, J.S. and Mores , jawetz ,S.A. , melnick, and Adel bergs. Medical microbiology. 24thed . McQraw-Hill-USA . (2007).
- 9- Todar, K. (2004). Pseudo monas aeruginosa of Wisconsin-madison depart ment of Bacteriology .

- 10- Benson , J.H.(2002). Microbiological Applications : Laboratory manual in General microbiology.8<sup>th</sup>ed.MC Graw Hill.P.145, 165-175
- 11-Henry J. B. (2001). Clinical diagnosis and Management by laboratory methods, 20th Edition volium3, PP 1088, 1106, 1123, Sound ears company
- 12- Collee , J.G.; Fraser , A .G .; Marmion , B.P. and simmons , A. ( 1996) . Mackie & MC cartney practical medical microbiology 14<sup>th</sup>ed . Churchill living stone INC. New York
- 13- Cole. St . and Nicolas ,MH. . B-lactamase resistance mechanisms in gram negative bacteria. Microbiological sciences.(1986).1:334-93
- 14- Blaser M. J, Smith P. D, Ravdin. J. I, Greenberg H. B., Guerrant R. L. (1995). Infections of the gastrointestinal tract, ch.95, Raven press Ltd, New York, p.1499-1523
- 15- Storz G. R., Aronis H. (2000). Chapter 22, Bacterial stress responses, p323-366, ASM press Washington, D. C
- 16- Wickens H., Wade P. (2005). Understanding Antibiotic Resistance, The pharmaceutical journal, Vol. 274, P 501-504
- 17- Bauer, A. W.; Kirby, W. M. M.; Sherris, J. C.; and Truck, M.(1966). Antibiotic susceptibility testing by a standardized single disk method. Am. J. Clin. Pathol., 45: 493 – 496.
- 18- Clinical and Laboratory Standards Institute (CLSI). (2012). Performance standards for antimicrobial susceptibility testing; 22ed. Informational Supplement. 32(3).
- 19- Brown, A . (2007). Bensons Microbiological application laboratory manual in general microbiology . McGraw –Hill Co.INC. USA . P:102 -263.
- 20- MacFaddin, J. F. (2000). Biochemical tests for identification of medical bacteria. 3rd ed. Lippincott Williams and Wilkins, USA.
- 21-T.T.Tan .Future .threat of gram –negative resistance in sing apore .Ann Acad Med sing apore .(2008).37:884-890.
- 22-Cornelis P. (2008). Pseudomonas: Genomics and Molecula Biology, 1st ed .,Caister Academic Press USA
- 23- Itah, A.Y. and Essien, J.P. . Growth Profile and Hydrocarbonoclastic Potential of Microorganisms Isolated from Tarballs in the Bight of Bonny , Nigeria ". WorldJournal of Microbiology and Biotechnology.(2005). 21 (6–7): 1317–22. doi:10.1007/s11274-004-6694-z.