

Synthesis of New Nucleoside Analogues from Theobromine

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Abstract

Nucleoside Analogues play important role in different medical drugs, therefore we synthesized new nucleoside analogues using theobromine as a nucleobase, for the first time, by conversion to its mercury salt, [1]. Then coupled with three kinds of sugar, including (glucose, galactose and mannose), after protection [2-4] and activation as bromo sugar [5-7] afforded blocked nucleoside analogues [8-10] which subsequently hydrolyzed to give our synthetic goal the free nucleoside analogues [11-13]. All prepared compounds were identified by FT-IR and some of them with ¹H-NMR spectroscopy. The synthesized compounds (11-13) were screened for their in-vitro antibacterial and antifungal activity. [DOI: [10.22401/ANJS.00.2.03](https://doi.org/10.22401/ANJS.00.2.03)]

Keywords: Nucleoside Analogues, theobromine, mercury salt

1. Introduction

Nucleosides are the key to life as they make up DNA and RNA in Nature. In medicine, many drugs contain synthetic nucleosides or nucleoside analogues for the treatment of disease, especially cancer and viral infection^[1,2]. Also to improve the pharmacologic activity^[3,4]. A variety of functionalities have been introduced into either the sugar moiety^[3,5] or the heterocyclic moiety^[6,7]. Nucleoside analogs represent a potentially important class of antiviral and anticancer agents^[8-10] with antimicrobial and cholinesterase inhibitory activities^[11-14] and are commonly used to treat hepatitis B virus^[15,16], hepatitis C virus^[17], herpes simplex^[18], Human Immunodeficiency Virus (HIV) and neoplasms^[19,20].

Well-established strategies of preparing N-nucleosides include the following: (1) Fischer and Helferich reported that purine nucleosides could be synthesized by coupling purines with acetobromoglucose and applying silver or mercury salts as catalysts^[21,22]; (2) the improved silyl-Hilbert-Johnson reaction^[23,24], the most widely used synthetic method, involves the coupling of per-silylated heterocyclic bases with per-acylated sugars in the presence of Friedel-Crafts catalysts (e.g., SnCl₄ or TMSOTf). This reaction has been the dominant method for the preparation of pyrimidine, purine and other heterocyclic nucleosides.

Theobromine (THB) is one of the major xanthine-like alkaloids, found in cacao plant and a variety of other foodstuffs such as tea leaves, guarana and cola nuts^[25].

Theobromine (from *Theobroma cacao*; theo = god, and broma = food; thus, food of the gods) are the two most abundant methylxanthines in chocolate, both of which have received considerable attention in the food and nutrition fields, in part because of the physiological effects which they elicit^[26,27].

Theobromine can be produced by both synthetically and natural route. For the chemical synthesis of theobromine, it is important to develop a highly stereoselective process, due to the 3,7-dihydro-3,7-dimethyl-1H-purine-2,6-dione are highly stereospecific nature^[28].

2. Experimental Part

2.1 Instruments and Chemicals:

2.1.1 Instruments:

- Melting points were recorded by using hot stage **Gallen Kamp** melting point apparatus and were uncorrected, England.
- Infrared spectra were recorded using Fourier Transform infrared **SHIMADZU** (8300) (FTIR) infrared spectrometer, Japan, KBr disc in the (4000-600) cm⁻¹ spectral range was performed by Baghdad University.
- ¹H-NMR spectra was recorded on near magnetic resonance Bruker, Ulter-shield

(400) MHz in Isfahan University, Iran, CDCl_3 was used as solvent.

- The biological activity were screened in central environmental laboratory in collage of science in university of Baghdad.

2.1.2 Chemicals:

All chemicals used in this study were of the highest purity available which were supplied from BDH, Fluka and Sigma-Alderich chemicals.

2.2 Preparation of Sugar Moiety

2.2.1 Preparation of Beta-glucose penta acetate[2]^[29]

A mixture of glucose (5 g, 27.75 mmol), sodium acetate anhydrous (4 g, 48.76 mmol) and acetic anhydride (25 ml) place in 100 ml round bottomed flask fitted with condenser. the mixture was heated on a water bath until a clear solution is obtained (approximately 30 minutes), the mixture was shake from time to time. then heating continue for 4 hrs. the reaction mixture is pour into (50 ml) of ice water in a beaker. the crystal was filtered at the pump and wash well with water. the product [2] is recrystallize from ethanol.(m.p 131-132), (9.3 g, 85.92 %).

2.2.2 Preparation of 1,2:3,4-Di-O-isopropylidene- α -D-galactopyranose[3]^[30a]

Freshly fused, and powdered anhydrous zinc chloride (9.5 g, 70 mmol) was rapidly weighed into a dry (250 ml) Erlenmeyer flask, and (100 ml) of dry acetone was added with stirring at room temperature until zinc chloride was dissolved. Concentrated sulfuric acid (0.32 ml) was rapidly added drop-wise. To the resulting colorless solution finely powdered anhydrous D-galactose (9 g, 50 mmole) was added. The mixture was stirred magnetically for 4 hours. After that a suspension of (16 g) of anhydrous sodium carbonate in (28 ml) of water was added in portions and the mixture was stirred for about one hour. The suspension was filtered, and the precipitate was washed several times with acetone. The filtrate and washings are combined, then the solution was evaporated under reduced pressure. The mixture was extracted 3 times with ether (3 \times 10 ml), dried

with anhydrous sodium sulfate, filtered, and evaporated to dryness under reduced pressure to give product [3] as pale yellow syrup, (9.54 g, 73.37%). Compound [3] showed physical properties according reference above.

2.2.3 Preparation of 2,3:5,6 di-O-isopropylidene- α -D-mannofuranose[4]^[30b]

α -D-Mannose (5 g, 27 mole) was shaken together with a (15) fold amount of anhydrous acetone (75 ml), containing (3.5 ml) of conc. sulfuric acid. After (3-4 h) all the D-mannose was dissolved. The light yellow solution was neutralized with anhydrous sodium carbonate (8 g in 14 ml water) and filtered. Then the filtrate was evaporated under reduced pressure and dissolved in a little amount of dry ether and precipitated with petroleum ether yielding [4] as yellow ppt. (6.3 g, 87.26%).

2.2.4 Preparation of α -Bromo glucose tetra acetate[5]^[30c]

The acetylated glucose [2] (0.5 g, 1.3 mmole) was treated with 50% hydrogen bromide in glacial acetic acid (2 ml) (which was added drop wise at 0 °C . The solution was kept at 0 °C until TLC indicated reaction completion (generally within one hour) then poured into an ice-cold chloroform (17 ml), washed with iced water (3 \times 15 ml) and then with saturated aqueous solution of sodium bicarbonate to remove the remaining acid. After a final washed with iced water (20 ml) the organic layer was dried over anhydrous sodium sulfate and the solvent was evaporated to give colorless syrup (0.44 g, 84.62 %). The product was used directly for the nucleoside synthesis.

2.2.5 Preparation of 1,2:3,4-Di-O-isopropylidene- α -D-galactopyranosyl bromide[6]^[30b]

Hydrogen bromide in glacial acetic acid (5 ml) of (45%) was added to 1,2:3,4-di-O-isopropylidene- α -D-galactopyranose [3] (4 g) then (5 ml) of glacial acetic acid was added. the mixture was stirred for 30 min. at room temperature and then left for 6 hours at room temperature. The mixture left over night at (5 °C) then the mixture was dissolve in

chloroform and neutralized with saturated aqueous sodium bicarbonate solution and extracted with chloroform (4×8 ml), dried over anhydrous sodium sulfate, filtered and evaporated in reduced pressure to give a brown syrup (3.54 g, 71.23%).

2.2.6 Preparation of Bromo 2,3:5,6 di-O-isopropylidene- α -D-mannofuranose [7]^[30b]

Compound [4] was brominated according to the same procedure for compound [3] with some modification. Hydrogen bromide in glacial acetic acid (45%) (2 ml) was added to 2,3:5,6-di-O-isopropylidene- α -D-mannofuranose [4] (1.5 g) then (2 ml) of glacial acetic acid was added. the mixture was stirred for 30 min. at room temperature and then left for 6 hours at room temperature. The mixture left over night at (5 oC) then the mixture was dissolve in chloroform and neutralized with saturated aqueous sodium bicarbonate solution and extracted with chloroform (3×5 ml), dried over anhydrous sodium sulfate, filtered and evaporated in reduced pressure to give a brown syrup (0.95 g, 51.08%).

2.3 Bis (theobromine-1-yl) mercury (II) [1]^[31]

This compound were prepared according literature⁽³¹⁾ with some modification. Theobromine (1 g, 5.49 mmol) was dissolved in hot water (30 mL) and sodium hydroxide (0.4 g, 5.2 mmol) was added, to vigorously stirred solution of mercuric chloride (0.82 g, 2.6 mmol) in hot ethanol (50 mL). The resulting yellow mixture was stirred at room temperature for 1.5 hours, then allowing it to stand at room temperature for 16 hours. The resulting suspension was cooled down and filtered off, then washed with distilled water until the filtrate was neutral to litmus. After filtration by suction a quantitative yield of [1] was obtained. m.p 326 dec.oC and stored in desiccators.

2.4 General procedure for synthesis of protected nucleoside analogues [8-10]^[32]

The theobromine mercury salt (0.18 g, 0.5 mmol) was finely powdered suspended in (20 ml) sodium-dried *m*-xylene in the presence of celite (1 g) to remove trace of

water the solvent was partially distilled. When the temperature of the mixture was raised to 137 °C, the residual suspension was allowed to cool (below 50 °C). The protected sugar [4-6] (0.97 mmol) in dried *m*-xylene (30 ml) was then added and refluxed with vigorous stirring for 1 h. TLC (chloroform-ether 9:1). Indicated the presence of unreacted trace of compound which was filtered from the hot xylene suspension and washed with dichloro methane (7 mL). The organic layer was washed with (3×5 mL) of 20% aqueous potassium iodide to remove the remaining trace of the mercuric salt, washed with water (3×5 mL) dried over anhydrous sodium sulphate and the solvent was removed to give protected nucleoside analogues [8-10]. Physical properties of these compounds were listed in table (4).

2.5 Hydrolysis of 1-(2',3':4',6'-tetra-O-Acetyl- β -D-glucopyranosyl) theobromine. [11]^[33]

A solution of (0.44 g, 0.862 mmol) of the blocked nucleoside [8] in (20mL) of 0.1M methanolic sodium methoxide was refluxed with stirring for 0.5 hour TLC (DCM: EtOH 8:2) showed that the reaction was complete, the mixture was neutralized with acetic acid (5 drops) and evaporated to dryness, the residue was partitioned between water and chloroform and the aqueous phase was evaporated to dryness in vacume. To give the nucleoside analogues [11] as off white syrup. (0.23 g, 82.14%).

2.6 Hydrolysis of 6-(1',2':3',4'-di-O-Isopropylidene-6-galacto pyranosyl) theobromine. [12]^[34]

For galacto nucleoside [9], a solution of blocked nucleoside (0.71 mmol) in (47 ml) of 50% aqueous acetic acid was refluxed for 15 min. (10 ml) of water was added and extracted with chloroform (3×5 ml), then dried over anhydrous Na₂SO₄, the solvent was evaporated to give the free nucleoside [12] (0.19 g, 82.6 %) as a yellow syrup.

2.7 Hydrolysis of 1-(2',3':5,6- di-O-Isopropylidene- β -D-manno furanosyl) theobromine.[13]^[34]

For manno nucleoside (10), a solution of blocked manno nucleoside (0.8 mmol) in (53 ml) of 50% aqueous acetic acid was refluxed for 15 min. (10 ml) of water was added and extracted with chloroform (3 \times 7 ml), then dried over anhydrous Na₂SO₄, the solvent was evaporated to give the free nucleoside [13] (0.2 g, 77 %) as deep brown syrup.

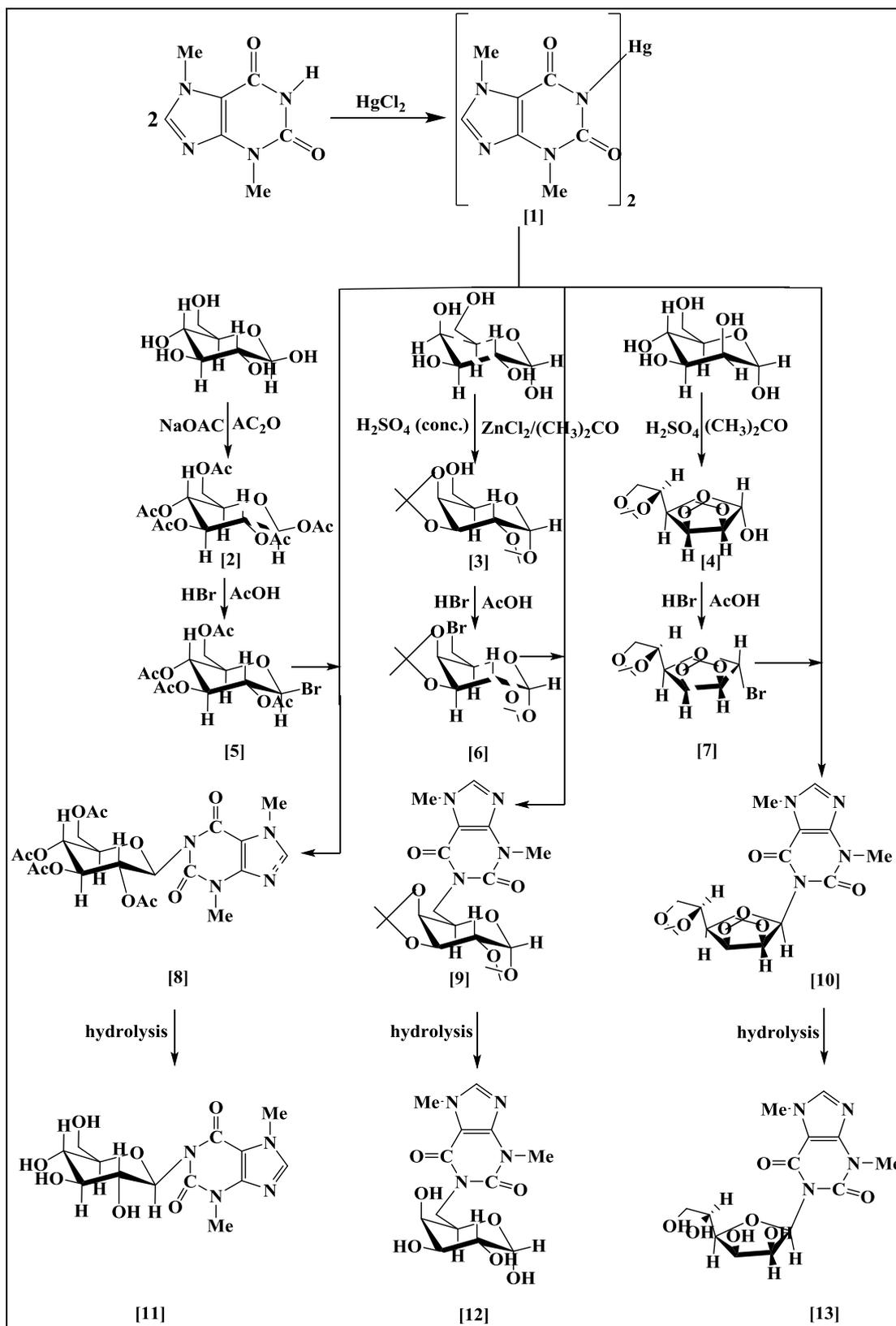
3. Anti-microbial activity test

The inhibition zone of growth of microorganisms was measured against *Staphylococcus aureus* and *Streptococcus* (Gram +ve) and *Escherichia coil* and *Pseudomonas aeruginosa* (Gram -ve) and *Aspergillus Flavus* (fungal) using Cup-plate methods. The petridishes were placed on a flat surface to ensure that the layers of the medium were of uniform thickness. Cylindrical cavities of 6 mm diameter were made on the medium. 50 μ l of test and standard solutions were transferred into cylindrical cavities, the plates were incubated for 24 h. for bacteria and 72 h. for fungal, at 37 °C and the circular inhibition zone was measured.

4. Results and discussion

Theobromine (THB) is a unique natural alkaloid, for there similarity to purine molecule (adenine and guanine) also it has moderate central nervous system (CNS) stimulant. On the other hand nucleoside analogues play important therapeutic agent for cancer, viral and bacterial disease.

For these importance we synthesized new nucleoside analogues, which were synthesized first of all. The designed synthetic rout was started with thebromine as a nucleobase and three kinds of sugar (glucose, galactose and mannose) as a sugar moiety (Schem 1).



Schem 1 Synthetic rout for synthesis of nucleoside analogues.

The base theobromine was converted to its mercury salt [1] to polarized the new covalent band leading to increase the nucleophilicity of nitrogen atom. The (THB) salt was characterized by it's high melting

point m.p > 326 °C and the FT-IR spectrum showed the disappearance of (N–H) band and other bands for theobromine itself. (table 1).

Table 1
The FT-IR spectral data of theobromine and its salt [1] and protected sugar.

Comp. No.	$\nu(\text{O-H})$	$\nu(\text{C-H})$ alkene	$\nu(\text{C=C})$ aromatic	$\nu(\text{C-H})$ aliphatic	$\nu(\text{C=N})$	$\nu(\text{C=O})$	$\nu(\text{C-N})$ aromatic	Other band
Theobromine	–	3033	1548	2952 2885	1595	1695 1670	1226 1335	3448 for $\nu(\text{N-H})$
1	–	3118	1500	2945	1539	1654 1631	1224 1313	–
2	–	–	–	2964	–	1762 (acetyl)	–	1074 for $\nu(\text{C-O-C})$ sugar
3	3465	–	–	2987	–	–	–	Sy.1039- asy.1255 for $\nu(\text{C-O-C})$ sugar
4	3485	–	–	2941	–	–	–	Sy.1072- asy.1251 for $\nu(\text{C-O-C})$ sugar

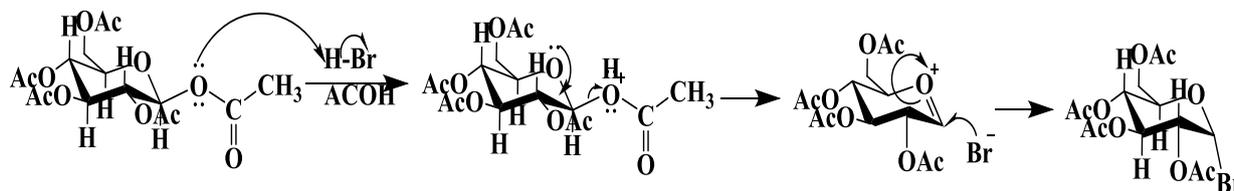
The $^1\text{H-NMR}$ spectrum showed three singlet signals at $\delta(3.52, 4.0$ and $7.54)$ ppm assigned to $(\text{CH}_3\text{-N})$ of pyrimidine, $\text{CH}_3\text{-N}$ of imidazole and proton of imidazole respectively the aromatic proton appeared at 7.54 ppm⁽³⁵⁾ the disappearance of N-H proton gives good indication for salt formation.

The sugar moiety was protected by too different way: glucose was protected as per acetate [2], using acetic anhydride and sodium acetate as a catalyst giving a pyrano conformation. while galactose and mannose protected as a di-*O*-isopropylidene using dry acetone and freshly fused zinc chloride as a catalyst. Galactose formed pyran conformation [3], while mannose formed furan conformation [4]. The protected sugar

were confirmed by their physical properties which were identical with literature⁽³⁰⁾

The FT-IR spectrum of sugar [2] showed appearance of carbonyl band of acetate group at (1751 cm^{-1}) while sugars [3 and 4] showed the stretching band of ether band (C-O-C) at $(1222\text{-}1240\text{ cm}^{-1})$ Table(1).

Glycosyl bromides are commonly synthesized by treating per acetylated sugar [3] with HBr in glacial acetic acid. "The more stable α -anomer usually obtained in high yield"⁽³⁶⁾. The anomeric acetyl was converted to good leaving group by protonation as shown in (Scheme 2) giving 1- bromo sugar [5].



Scheme 2 Mechanism for synthesis of α -Bromo glucose tetra acetate.

Galactose and mannose were brominated at free OH in C_6 and C_1 producing bromo sugar [6] and [7] respectively. using the same reagent as in [5] with different period of reaction. Following the Koenigs- Knorr

method the bromo sugar [5-7] (which was prepared immediately) was subjected to coupled with theobromine mercury salt [1] in dry xylene with appearance of celite in reflux condition afforded the protected nucleoside

[8-10] via S_N2 reaction by displacement of mercury atom by sugar bromide group. The new protected nucleoside was characterized by FT-IR spectroscopy. the spectrum showed the disappearance of (C–Hg) band and appearance of band between (1226-1375) cm^{-1} for (C–N) nucleoside, also showed a stretching bands of (1550-1599 cm^{-1}) and at (1610-1706 cm^{-1}) for (C=N) and imide carbonyl respectively for theobromine moiety.

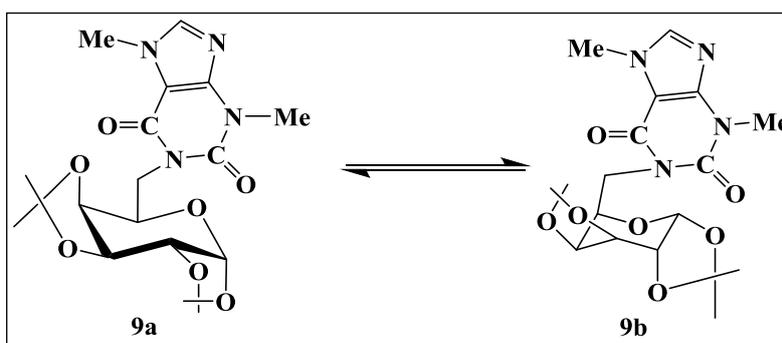
In addition of these band nucleoside [8] showed absorption band at 1751 cm^{-1} for carbonyl of acetyl group in sugar moiety, while nucleoside [9 and 10] showed a stretching band at (1224-1240 cm^{-1}) and at (1047-1049 cm^{-1}) for asymmetric and symmetric (c–o–c) for isopropylidene groups in sugar moiety. Other absorption bands are listed in (table 2).

Table 2
The FT-IR spectral data of protected nucleoside analogues [8-10]

Comp. No.	$\nu(C-H)$ aliphatic	$\nu(C=O)$	$\nu(C=N)$	$\nu(C-N)$ aromatic	Other band
8	2958	1610 1680 1751	1595	1226 1369	1041 for $\nu(c-o-c)$ sugar 1751 for $\nu(c=O)$ acetyl
9	2923 2856	1606 1706	1599	1228 1371	$\nu(c-o-c)$ Sy. 1047 Asy. 1224
10	2983 2943	1610 1666	1550	1230 1375	$\nu(c-o-c)$ Sy. 1049 Asy. 1240

Nucleoside [9] conformed also by 1H -NMR its spectrum showed a complicated spectrum which may be due to flipy of chair conformation that is gives other non isolated

conformer which cause to interpenetrate of the signals for the same proton of two conformer as shown below:



To obtain our target the free nucleoside, the protected group was hydrolyzed using sodium methoxide in methanol and aqueous acetic acid for nucleoside [8] and [9 and 10] respectively, forming new free form [11-13]. The FT-IR spectrum of these nucleoside showed the appearance of stretching band at

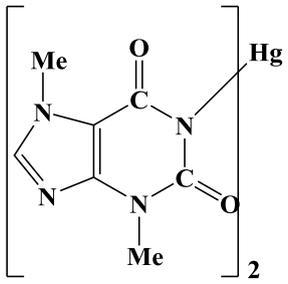
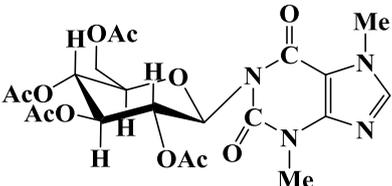
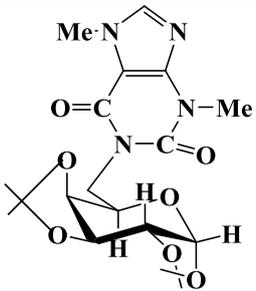
(3427-3390 cm^{-1}) for hydroxyl group and C–H aliphatic of sugar moiety which gives a good indication for formation of free form. Other characteristic bands for theobromine base are listed in table 3.

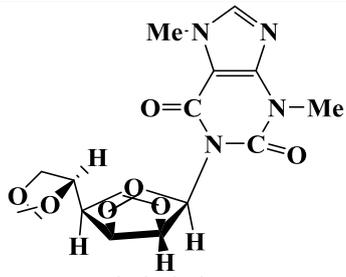
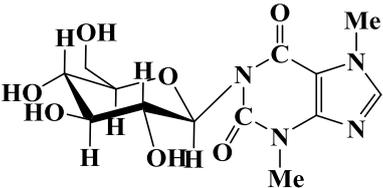
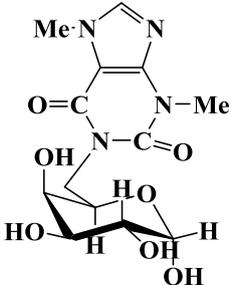
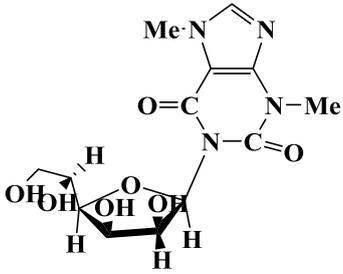
Table 3
The FT-IR spectral data for free nucleoside analogues[11-13]

Comp.No.	$\nu(\text{O-H})$	$\nu(\text{C-H})$ aliphatic	$\nu(\text{C=O})$	$\nu(\text{C=N})$	$\nu(\text{C-N})$ aromatic	$\nu(\text{C-O-C})_{\text{sugar}}$
11	3427	2930	1634 1701	1562	1361	1071 3618
12	3390	2963	1639 1704	1566	1340	1051
13	3402	2979 2941	1605 1668	1580	1229 1373	1080 1230

Physical properties of the prepared compounds (1) and (8-13) and other data are listed in Table 4.

Table 4
Physical properties of the prepared compounds (1) and (8-13)

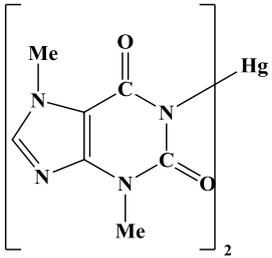
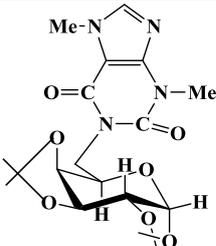
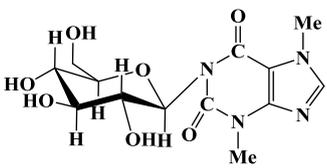
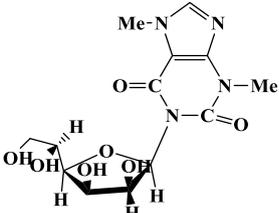
Comp. No.	Compound structure	Molecular formula	M.wt (g/mol)	M.P °C	Colour	Yield %
1	 <p>Bis(theobromine-1-yl)mercury(II)</p>	$\text{C}_7\text{H}_7\text{HgN}_4\text{O}_2$	379.16	326	Yellow	82
8	 <p>1-(2',3':4',6'-tetra-O-Acetyl-β-D-glucopyranosyl) theobromine</p>	$\text{C}_{21}\text{H}_{26}\text{N}_4\text{O}_{11}$	510.16	syrup	Color less	78
9	 <p>1-(1',2':3',4'-di-O-Isopropylidene-6-deoxy-D-galacto pyranosyl) theobromine</p>	$\text{C}_{15}\text{H}_{20}\text{N}_4\text{O}_7$	422.44	syrup	Brown	81

10	 <p>1-(2',3':5',6'- di-O-Isopropylidene-β-D-mannofuranosyl) theobromine</p>	$C_{15}H_{20}N_4O_7$	368.35	Syrup	Yellow	75
11	 <p>1-(β-D-Glucopyranosyl) theobromine</p>	$C_{13}H_{18}N_4O_7$	326.31	syrup	Off white	82.14
12	 <p>1-(β-D-6'-deoxygalactopyranosyl) theobromine</p>	$C_{13}H_{18}N_4O_7$	326.31	Syrup	Yellowish	82.6
13	 <p>1-(β-D-Mannofuranosyl) theobromine</p>	$C_{13}H_{18}N_4O_7$	326.31	Syrup	Deep brown	77

Nucleoside [11] and [13] were conformed by 1H -NMR. The spectrum of nucleoside [11] showed two singlet signals at δ (1.28 and 2.09) ppm for two CH_3 group linked with nitrogen in six and five membered ring respectively. The (O-H) group appeared at 2.38 ppm as a singlet. The sugar protons appeared at (3.4-3.9; 4.3-4.35; 4.42-4.48; 4.61-4.67; 4.87-4.91 and 5.9-5.97) ppm as multiplet, triplet doublet, doublet and quartete assigned to H'_5 , H'_4, H'_3 , H'_2 , H'_1 and H'_6 and H''_6 respectively while the C-H aromatic protone appeared at

the downfield at 7.54 ppm as a singlet. (table 4). Nucleoside [13] showed the singlet signal at 2.03 ppm belong to hydroxyl group the two methyl group of theobromine moiety appeared as a singlet at (3.4 and 3.91) ppm other singlet appeared at downfield 7.53 ppm for C-H theobromine, the sugar protons appeared as multiplet, doublet, multiplet, double-doublet, doublet and doublet for H'_5 , H'_4, H'_6 , H''_6 , H'_3 , H'_2 and H'_1 in the region between (5.08-6.97) ppm. All these data are listed in (table 5).

Table 5
¹H-NMR – Spectral data for compound (1,9,11,13)

No.	Compounds	δ ppm
1		3.52(s,3H,CH ₃ N-Pyrimidine); 4.0, (s,3H,CH ₃ ,N-imidazole); 7.54(S,1H, C-H imidazole).
9		2.01-2.45(s,18H,6CH ₃); 4.03-4.5(m,1H,H' ₅); 4.62-4.72 (t,2H,H' ₆ ,H'' ₆);4.98-5.8 (m,4H,H' ₄ ,H' ₃ ,H' ₂ ,H' ₁); 7.7 (s,1H,C-H imidazole).
11		1.28(s,3H,CH ₃ N- Pyrimidine); 2.09(s,3H, CH ₃ N- imidazole); 2.38(s,4H, 4OH); 3.4-3.9(m,1H,H' ₅); 4.3-4.35(t,1H,H' ₄); 4.42-4.48(d,1H,H' ₃); 4.61-4.67(d,1H,H' ₂); 4.87-4.91(d,1H,H' ₁); 5.9-5.97(9,2H,H' ₆ ,H'' ₆); 7.54(S,1H,C-H imidazole).
13		2.03(s, 4 OH); 3.46(s, 3H, CH ₃ Pyrimidine); 3.91(s,3H,CH ₃ imidazole); 5.08-5.28 (m,1H,H' ₅); 5.83-5.85(d,1H,H' ₄); 6.02-6.11(m,2H,H' ₆ ,H'' ₆); 6.55-6.59(d,1H,H' ₃); 6.65-6.68(d,1H,H' ₂); 6.95-6.97(d,1H,H' ₁); 7.53(s,1H,C-H imidazole).

Biological activity

Microbiological Test: The synthesized compounds [11-13] were screened in vitro against four types of bacteria including, *staphylococcus aureus* and *streptococcus* as gram positive and *E.coil* and *pseudomonas auroginosa* as gram negative,also screened antifungal activity against *Aspergillus Flavus*. The obtained results are listed in table (5)

Showed that all synthesized compound have good activity against the four types of bacteria, while inactive against all types of fungi used for this test. This results indicate that the synthesized compounds is specific as antibacterial. This results was coincide with literature^[37].

Table 6
Inhibition Zones of compounds (11-13)

Comp. No.	<i>Escherichia coil</i>	<i>Pseudomonas auroginosa</i>	<i>Staphylococcus aureus</i>	<i>Streptococcus</i>	<i>Aspergillus Flavus</i>
Control	–	–	–	–	–
11	9	10	12	10	–
12	12	8	11	9	–
13	10	11	12	8	–

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