THIAMINE – INDUCED FORMATION OF THE MONOPYRROLE MOIETY OF PRODIGIOSIN

Sanad B. AL-Arrji*  Zeena F.Ahmed**  Anas G. Abdul kareem***

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ABSTRACT:
Thiamine stimulates the production of a red pigment, which is chromatographically and spectrophotometrically identical to prodigiosin, by growing cultures of *serratia marcescens* mutant 9-3-3. This mutant is blocked in the formation of 2- methyl -3- amyl pyrorol(MAP), the monopyrrole moiety of prodigiosin, but accumulates 4-methoxy-2, 2-bipyrrrole -5- carboxaldehyde (MBC) and can couple this compound with(MAP) to form prodigiosin.

Addition of thiamine caused production of MAP, and as little as 0.02 mg of thiamine / ml in peptone- glycerol medium stimulated production of measurable amounts of prodigiosin. Phosphate saltes and another type of peptone decreased the thiamine- induced formation of prodigiosin ,yeast extract and glycerol enhanced formation of this substance.

Thiamine also enhanced production of prodigiosin by wild – type Strain Nima of *S. marcescens*. The pyrimidine moiety of thiamine was also 10% as effective as the vitamin; the thiazol moiety only 4% , and the two moieties together , 25% . Thiamine did not stimulate production of prodigiosin biosynthesis as strain 9-3-3 . This is not surprising since strain 9-3-3 originated as a result of two mutational events one event may involve thiamine directly , and the other may involve the biosynthesis of MAP.

INTRODUCTION:
Prodigiosin, the red pigment of *serratia marcescens* has been characterized as atripyrpyryl methane although this structure has never been confirmed by synthesis [11].

Prodigiosin is separated into four fraction: one blue, two red, and one orange, the blue fraction has not been reported previously [18]. Its produced by *S marcescens*, pseudomonas magnesiourubra, *Vibrio psychroerythrus* and other bacteria [6] [14].

Microscopic observation of *S marcescens* colonies showed that Prodigiosin pigment was localized in vesicles (extra cellular and cell associated ) or intercellular granules [9] [12].

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this pigment is synthesized in a bifurcated pathway, in which mono-end bipyrrrole precursors are synthesized separately and then coupled to form prodigiosin.

AIM OF THE STUDY:
This study aimed for detection prodigiosin which is produced from bacteria *S marcescens* and the role of thiamine to induced this pigment.

MATERAIL AND METHODS:
ORGANISMS:
Strains of *serratia marcescens* use in this investigation were:
1. Nima, Atypical ,wild-type strain that produces prodigiosin[18].
2. Mutan WF [2 ] [19 ] which produces 2- methyl -3- amyl pyrorol( MAP)but is blocked in synthesis of 4-methoxy-2, 2- bipyrrrole -5- carboxaldehyde (MBC) ,and yet can couple these two moieties to form prodigiosin [3 ] [ 13 ] .
3. Mutan 9-3-3 [2 ] which extract MBC and also produces the enzyme for terminal biosynthesis of prodigiosin but is blocked in synthesis of MAP [ 15 ] .
4. Mutan H – 262 ,which is phenotypically like strain 9-3-3 .

MEDIA:
The organism were maintained in stock on slants containing 2.5 % agar , 0.5% peptone (difco) end either 0.5 % L- prolin or 1.0% glycerol .
Liquid media used for quantitative experiments were :
1. PG , a medium containing 0.5% peptone and 1.0% glycerol in distilled water [ 5 ] .
2. MM , the minimal medium of Bunting [ 5 ] .
3. CM ,a complete medium [ 18 ] made by adding 0.1 % yeast extract (difco) and 0.2% N-Z case peptone to MM medium .
4. SM , synthesis medium [ 7 ] containing distilled water 0.7 % K2HPO4 , 0.3% KH2 PO4 , 0.05% Na3C2H2O7.2H2O , 0.01% MgSO4 , 0.1% (NH4)2SO4 and 0.2% glucose that was autoclaved separately and added aseptically to the other ingredients after they had been autoclaved.
All media were adjusted to { pH 7.2 } before autoclaving .
Thiamine and other test compounds were sterilized by filtration through membrane filters and then added to various media as indicated in the text cultures were grown in 250 ml or liter Erlenmeyer flasks containing 200 ml of liquid media .

INCUBATION :
All cultures were incubated aerobically at 27 C . liquid cultures were shaken on a new bruns – wick rotary shaker ( model G 26 ) having a displacement of 1 inch & rotating at 198 strokes/min .

PIGMENT INTERMEDIATES AND CHEMICALS:
The supernatant fluid from cultures of strain 9-3-3 grown for 24 hr in PG medium was used as the source of MBC .
Cells were removed by centrifugation , and the supernatant fluid was sterilized by membrane filtration .
cultures of strain WF growing on PG agar were the source of the natural monopyrol ,
the commercial compounds were diluted in 0.05 µ phosphate buffer , pH 6.8 , before sterilization.

ANALYTICAL PROCEDURES:
Protein was determined by the method of Lowry etal .[10 ] with bovine serum albumin as a standard .
The amount of MBC present was determined by first extracting the supernatant fluid with chloroform and determining spectrophotometrically the difference in absorption at 363 and 400 nm [ 16 ] .
This difference was proportional to amount of MBC in the supernatant fluid. Prodigiosin and prodigiosin analogue were measured by extracting pigmented suspensions with acidic methanol and then measuring the difference in absorption of the extract at 534 & 655 nm [ 18 ] .
The same wave length could be used to measure both compounds a difference in absorbancy of 1.0 between the two wave lengths was equivalent to 19.3 µg of prodigiosin or 17.2 µg of prodigiosin analogue 1 mg bacterial protein .
An indirect assay [ 17 ] was used to measure the volatile MAP because as yet no direct assay has been developed for this compound .
The indirect assay utilized the synthrophic interaction of mutant strain WF and 9-3-3 ;
the volatile MAP produced by strain WF or strain 9-3-3 when grown with thiamine, is coupled with the MBC synthesized by strain 9-3-3 to form prodigiosin. Strain WF or strain 9-3-3 was grown on the appropriate test medium in the bottom of a Petridish, and strain 9-3-3 was spread uniformly over the surface of PG agar in the top of the same Petri dish. The amount of prodigiosin formed in the latter culture of strain 9-3-3, after incubation of the closed Petridish at 27°C for 48 hr, was then measured by the procedure given above. One molecule each of MBC and MAP couple to form one molecule of prodigiosin.

RESULTS:

When thiamine was added to PG medium, mutant strain 9-3-3 produced a red pigment (Fig 1).

Measurable amounts of pigment appeared at a thiamine concentration of 0.02 µg/ml of medium. Up to a concentration of 10 to 14 µg of thiamine/ml, the amount of pigment formed increased almost linearity (Fig 1). Higher concentration of the vitamin resulted in only slight increases in pigmentation.

As will be shown later, addition of thiamine to PG medium also enhanced production of prodigiosin by growing cultures of the wild – type strain, Nima.

Addition of thiamine to other media also promoted formation of prodigiosin by strain 9-3-3 (Table 1 & 2). Phosphate salts {Table 1} and N-Z case peptone {Table 2} reduced the amount of prodigiosin formed even in the presence of thiamine. Yeast extract and glycerol enhanced the effects of thiamine {Table 2}.

In medium containing 100 µg of thiamine/ml, 1.5% glycerol, and 0.2% yeast extract, almost 90 µg of prodigiosin/mg of protein was formed a value that represented about 1 to 2% of the dry weight of cells.

Table 1: Production of prodigiosin by S. marcescens strain 9-3-3 when grown on various liquid media

<table>
<thead>
<tr>
<th>Medium</th>
<th>Prodigiosin(µg/mg of protein/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.5% peptone &amp; 0.5 % praline</td>
<td>0.21</td>
</tr>
<tr>
<td>0.5% peptone &amp; 0.5 % praline plus thiamine</td>
<td>13.37</td>
</tr>
<tr>
<td>0.5% peptone &amp; 1.0% glycerol</td>
<td>0.37</td>
</tr>
<tr>
<td>0.5% peptone &amp; 1.0% glycerol plus thiamine</td>
<td>17.93</td>
</tr>
<tr>
<td>Minimal medium (MM)</td>
<td>0.12</td>
</tr>
<tr>
<td>MM plus thiamine</td>
<td>2.62</td>
</tr>
<tr>
<td>Complete medium (CM)</td>
<td>0.15</td>
</tr>
<tr>
<td>CM plus thiamine</td>
<td>2.30</td>
</tr>
<tr>
<td>CM with phosphate salts omitted</td>
<td>1.24</td>
</tr>
<tr>
<td>CM with phosphate salts omitted plus thiamine</td>
<td>48.37</td>
</tr>
<tr>
<td>Synthetic medium (SM)</td>
<td>0.23</td>
</tr>
<tr>
<td>SM plus thiamine</td>
<td>1.91</td>
</tr>
<tr>
<td>SM with low phosphate plus 0.1%tris-(hydroxymethyl) mimemethane (Tris)buffer</td>
<td>0.23</td>
</tr>
<tr>
<td>SM with low phate plus 0.1%tris buffer plus thiamine</td>
<td>4.38</td>
</tr>
</tbody>
</table>
Table 2: Effect of complex natural nutrients upon thiamin-induced formation of prodigiosin by S. arcscens strain 9-3-3

| Additions to basal medium                  | Prodigiosin(µg  
| mg of protein/  ml) | %formed  
| compared to  thiamine ) |
|-------------------------------------------|-----------------|
| 0.2% yeast extract                        | 1.35            |
| 0.2% yeast extract plus thiamine          | 6.56            |
| 0.1% yeast extract                        | 1.43            |
| 0.1% yeast extract plus thiamine          | 26.11           |
| 0.1% yeast extract plus 0.2%N-Z case peptone | 1.25            |
| 0.1% yeast extract plus 0.2%N-Z case peptone plus thiamine | 5.92            |
| 0.1% yeast extract plus 0.2%N-Z case peptone plus 1% glycerol with thiamine | 1.40            |
| 0.2%N-Z case peptone                      | 0.42            |
| 0.1%N-Z case peptone                       | 0.42            |
| 0.1%N-Z case peptone plus thiamine            | 0.42            |

since addition of thiamine either to PG medium or to CM medium from which phosphate salts were omitted induced formation of significant amounts of prodigiosin, these media were used in most of our experiment.

The pyrimidine moiety of thiamine 2-methyl - 4- amino - 5- aminomethyl pyrimidine, was about 10% as effective as the vitamin in inducing prodigiosin formation in strain 9-3-3. The Thiazole moiety, 4- methyl – 5 (ß-hydroxyethyl)- thiazole, was evenless effective.

Addition of both moieties to PG medium gave about 25% of the response of thiamine itself. The above data suggested that the addition of thiamine enabled strain 9-3-3 to synthesize MAP.

DISCUSSION:

Thiamine induced formation of pigment in mutant strain 9-3-3 by stimulating production of MAP. Spectral & chromatographic evidence indicated that the pigment was prodigiosin. Thiamine did not cause reversion of strain 9-3-3 to a pigmented state, nor did it favor the selection of a spontaneous, pigmented mutant. When red pigmented cells of strain 9-3-3 were harvested washed, and then inoculated into fresh medium without thiamine, their ability to form prodigiosin disappeared. Thiamine also enhanced production of prodigiosin by wild-type strain nima, however strain H-262, a single-step mutant that is phenotypically like strain 9-3-3, did not form pigment when grown in the presence of thiamine. This was not surprising, since strain 9-3-3 is a spontaneous mutant isolated from another spontaneous mutant, strain 9-3-3 that originated from wild-type strain 274 of S. marcescens. Thus, strain 9-3-3 arose as the result of two mutations. Although both mutational events might affect biosynthesis of MAP, only one might involve thiamine. The vitamin might relieve one block, allowing synthesis of an intermediate that could be used to overcome the second block. Relief of the second block would permit biosynthesis of MAP.

Mutant strain 9-3-3 grew well on a minimal medium containing no thiamine, the organism can synthesize enough of the vitamin to carry out basic metabolic reaction.

Mutant strain 9-3-3 also apparently synthesize only a limited amount of the
pyrimidine component of the vitamin , since additional ( amounts of this moiety do induce formation of some prodigiosin . Addition of the thiazole moiety induce formation of only a limited amount of pigment . Addition of both moieties together does not cause formation of pigment equivalent to that formed by the addition of the whole molecule , thus , the mutant evidently has a limited capacity to couple the two moieties to form thiamine . strain 9-3-3 grows well aerobically , presumably by utilizing cytochrome enzymes . These observations indicate that the mutant synthesizes the pyrrole groups of the porphyrins found in these enzymes . the usual path way for pyrrole biosynthesis involves δ- amino levulinic acid , but this compound is poorly incorporated into prodigiosin [11] [19] . this fact suggests that pyrrole groups of prodigiosin are synthesized by different path way than the pyrrole of the porphyrins. The pyrrole moieties of prodigiosin , MAP & MBC , although they may have common early precursors [13] . our data indicate that thiamine influence synthesis of MAP , but the effect of the vitamin may be indirect and may involve some other pathway , rather than affecting synthesis of MAP directly . the vitamin dose not substitute for MAP because addition of thiamine to suspensions of cells of strain 9-3-3 . phosphate salts inhibit biosynthesis of prodigiosin by wild– type organism[1] [9] , biosynthesis of MBC in strain 9-3-3 [17] and biosynthesis of MAP & MBC in other mutas [5] [13] . phosphate causes a similar inhibition of thiamine- induced pigmentation in strain 9-3-3 . The effect of thiamine and of phosphate salts may be related and may involve a common biosynthetic step. Interestingly , when strain 9-3-3 is grown in the absence of thiamine in media lacking phosphate salts [14] or in PG medium , it produces a purple pigment .

This pigment has different spectral characteristic than prodigiosin and is probably identical to the dipyrolyl dipyrromethene analogue of prodigiosin recently reported [15].

CONCLUSION
1. Serratia marcescens produces a red pigment when grown aerobically at temperatures below 37 C.
2. Mutant strain 9-3-3 is blocked in synthesis of MAP but can synthesis MBC.
3. When thiamine is added to the growth medium , strain 9-3-3 produces a red pigment.
4. Thiamine is probably involved in the regulation of the biosynthesis of MAP.

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الخلاصة :

الثامن يحظى انتاج الصبغة الحمراء البروديجيوزين والتي يمكن قياسها بالكروماتوغرافيا والمطيافية الضوئية
، عن طريق تدفق إلكترود المشتركة المتبقية من الطرف 9-3 في الوسط الزراعي.

هذه الطفرة تخارية تكويين-3- ميل بيرول (MAP) . يشتر热水احاد البيرو الاصبغة البروديجيوزين لكسن
 Diese الطفرة تخارية تكويين-3- ميل بيرول (MAP) . يمكن دمج هذا المركب مع

أضافة الثامن يبيث في انتاج (MAP) تسامح كمية قليلة من الثامن بمقدار 0.02 ملم/هل/مل/مل في وسط بيتون -

MBCQ. يمكح ميترية أثر كلاصية الصبغة البروديجيوزين الم终结ية.

املاح الفوسفات储存ات أخرى من البيتون تنقص من تحقيز الثامن على تكوين صبغة البروديجيوزين، خلاصة

الخياري واللكسيبرول يعزز تكوين هذه المادة.

الثامن يحظى انتاج الصبغة البروديجيوزين من سلالة النوع العالي ولسلالة المشتركة المتبقية من نوع

30% شائر البروينيين للثامن تكوين فقط بنسبة 10% أي يكون تأثيرها كفتيثاء بينما شائر النويزيبل فقط بنسبة 4%

وتشير عما نسبة 25% .

الثامن لإحتضن على البناء الحيوي لإنتاج صبغة البروديجيوزين مثل سلالة 9-3 وهذا لاعتبر مفاجئة حيث أن

السلاة 9-3 هي نتيجة حدث طفرتين واحدة منها تستلزم الثامن مباشرة , والآخر ربما تتضمن البناء

الحيوي (MAP)