

CONCOMITANT OF ANAPLASMOSIS WITH ACID-BASE BALANCE ,BLOOD GAS ANALYSIS , ACUTE PHASE RESPONSE AND HEMOGRAM IN CATTLE

Kamal.M. Alsaad Hussam.MS. Alimam

Department of internal and preventive medicine College of Veterinary Medicine,
University of Mosul, Mosul, Iraq.

Keywords; Anaplasmosis ,Cattle , Blood.

(Received 26May 2013,Accepted 5 June 2013)

ABSTRACT

The objective of the present work was to evaluate Hemogram, acid-base balance, blood gas analysis and acute phase response in cattle infected with anaplasmosis in Mosul,Iraq.The study were conducted on (167), local cattle breed 2-5 years old of both sexes. Eighty local cattle breed out of (167) were clinically infected with *Aanaplasam marginale* since the causative Rickettsia diagnosed by Giemsa stained blood smears and confirmed by indirect Elisa test. (20) clinically normal cattle were served as controls. Results indicated statistically significant decrease ($P<0.05$) in TRBCs, Hb, and PCV values of diseased animals, Macrocytic hypochromic type of anemia was indicated and the percentage of Rickettsemia (Hemoparasitism) ranged between (12-21%) with a mean of (15.57%). Results also indicated a significant increase in TLC as a result of significant increase lymphocytes. *A marginale* were detected in 96.25% of diseased cattle .There were no significant difference encountered in clotting factor indices. Statistically significant decrease were encountered in P_{CO_2} , Oxygen saturation percent (SO_2), bicarbonate ions, Base excess and blood pH in diseased cattle than in controls, However statistically increase in Anionic gap have been detected, Moreover Titritonal metabolic acidosis were registered . Statistically significant difference have been encountered in acute phase response as haptoglobin and fibrinogen values were increased in clinically infected cattle than in controls.

Key words: Anaplasmosis , Cattle, hemogram, Acute phase response, Acid base balance, Blood gas analysis.

INTRODUCTION

Anaplasmosis is an infectious, non contagious, tick born disease of domesticated and wild ruminants. Progressive anemia, emaciation , digestive disturbances, and increase body temperature are the main characteristics of this disease (1,2).

The disease were globally distribution, especially over the tropical and sub tropical regions, Nevertheless it was also recorded in some temperate areas(3). The disease either seen sporadically or as outbreaks leading to a deleterious significant economic losses (4). In Iraq, The disease has wide distribution especially at the north parts (5,6,7,8,9,).

Studies of anaplasmosis in local cattle breed at Mosul, Iraq concerning evaluation of acute phase response, acid –base balance and blood gas analysis are very limited and little information had been provided. Therefore the main objects of this study was to investigate, hematological observation, and some biochemical

changes as well as the effect of anaplasmosis on clotting factors indices in cattle naturally infected with *Anaplasma marginale*.

MATERIALS AND METHODS

The study were conducted on (167), local cattle breed 2-5 years old of both sexes in Mosul , Iraq . Eighty local cattle breed out of (167) were clinically infected with *Aanaplasma marginale* since the causative Rickettsia diagnosed by Giemsa stained blood smears and confirmed by indirect Elisa test ,(Svanova- Sweden). (20) clinically normal cattle served as controls. Clinical examination of all animals had been carried out, and fecal samples were screened for parasitic loud using standard technique.

Blood samples (11 ml) were obtained from each animal via jugular vein-puncture. 2.5 ml of Blood mixed with EDTA used to determine erythrocyte count (TRBs), haemoglobin (HB), packed cell volume (PCV),Mean corpuscular volume (MCV),Mean corpuscular hemoglobin concentration (MCHC), platelets count (Plt), mean platelets volume (MPV), platelets distribution width (PDW), total leukocyte counts (TLC) on an automatic full digital cell counter (Beckman,USA). Clotting time (CT) was also estimated according to (10).

Giemsa-stained blood smears were used to evaluated differential leukocyte counts (11), Moreover another 2.5 ml of blood mixed with trisodium citrate was used to determine fibrinogen, prothrombin time (Prt) and activated partial thrompoplastine time (Appt) using commercial kits (Biolabo, France).The remaining (5mL) of blood were used for obtaining serum, Serum haptoglobin concentrations were assayed according to (12). 1 mL of blood mixed with heparin were drained separately from each cattle used to determine P_{CO_2} , P_{O_2} ,Oxygen saturation percent(SO_2), Bicarbonate, Bass access , Anionic gap and Blood pH, Sodium and potassium (Opti-critical care analyzer/ USA) according to (13).Serum chloride values were estimated according to (14).

Statistical analysis The significance of variations between diseased and healthy cattle were statistically analyzed using T-test (SPSS), (15).

RESULTS

Clinically infected cattle show sings of paleness of mucus membranes, fever, loss of appetite, emaciation, rough hair coat, lacrimation with discharging serious ocular discharge, furthermore ticks were detected on different regions of the body.

Anaplasma marginale appears as spherical granules near periphery of infected red blood cells and Rickettsemia ranged between (12-21%) with a mean of (15.57) , Fig.1.

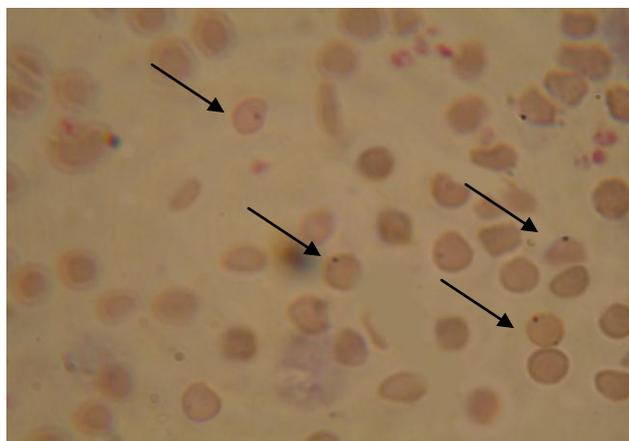


Fig.1: *A. marginale* in blood smear of cow erythrocytes

There was a significant decrease ($P<0.05$) in the mean values of TRBs, Hb and PCV, in diseased cattle infected with Anaplasmosis and anemia was of Macrocytic hypochromic type. Results also indicated significant increase ($p<0.05$) in total leukocytes count as result of significant increase ($p<0.05$) lymphocytes. Table 1.and 2.

Table (1): Blood parameters of cattle infected cattle with Anaplasmosis and controls

Parameters	Control cattle	Infected Cattle
TRBs ($\times 10^6$)	8.34 \pm 0.67	4.83 \pm 0.89 *
Hb (g/dl)	10.87 \pm 0.61	6.23 \pm 0.39 *
PCV (%)	34.36 \pm 1.16	22.75 \pm 1.43 *
MCV/fl	41.19 \pm 1.57	47.12 \pm 1.46*
MCHC /dl	31.63 \pm 0.45	27.38 \pm 5.57*
Parasitemia	-----	15.57 \pm 6.81*

* ($P<0.05$), Values are mean \pm standard

error of mean

Table(2): Total and absolute differential leukocyte count of cattle infected with anaplasmosis and controls

Parameters	Control cattle	Infected cattle
TLC($\times 10^3$)	10.48 \pm 0.35	13.66 \pm 1.75*
N/ <i>absolute</i>	4673.7 \pm 144.16	3982.2 \pm 50.67 *
L/ <i>absolute</i>	4481.7 \pm 143.1	7055 \pm 726.36*
M/ <i>absolute</i>	422.76 \pm 113.66	566 \pm 108.78
E/ <i>absolute</i>	514.66 \pm 204.65	565.21 \pm 53.76
B/ <i>absolute</i>	73.43 \pm 77.33	75 \pm 77.3

* ($P<0.05$), Values are mean \pm standard error of mean

Moreover there was no significant difference were encountered in clotting factors indices among infected and control groups of cattle, Table 3.

Table(3): Clotting factors indices of infected cattle with anaplasmosis and controls

Parameters	Control cattle	Infected cattle
Plt $\times 10^3$	408.23 \pm 50.62	392.21 \pm 70.65
MPV /fl	9.32 \pm 1.26	9.63 \pm 3.22
PDW %	17.52 \pm 2.51	18.76 \pm 4.24
CT / mint	3.22 \pm 0.78	3.87 \pm 2.49
Prt / sec	11.52 \pm 4.51	12.43 \pm 5.57
Aptt /sec	60.82 \pm 3.359	63.51 \pm 7.463

* (P<0.05), Values are mean \pm standard error of mean

A marginale was diagnosed on the basis of Giemsa stained blood smears and was confirmed by Indirect ELISA test and results showed that out of 80 samples tested (96.25%) of cattle were positive, whereas (3.75%) were detected as negative .

Results were also showed significant decrease(p<0.05)in P_{CO_2} , Oxygen saturation percent, Bicarbonate, Base excess and blood pH, however significant increase(p<0.05) in Anion gap were indicated, Moreover Titritonal metabolic acidosis were registered. Table 4.

Table(4): Acid-base balance and blood gas analysis of cattle infected with anaplasmosis and controls

Parameters	Control cattle	Infected cattle
P_{CO_2} /mm Hg	46.91 \pm 1.65	39.61 \pm 3.76 *
P_{O_2} /mm Hg	152.61 \pm 5.61	152.87 \pm 6.81
SO ₂ %	94%	83% *
Bicarbonate mEq/L	26.65 \pm 3.76	20.84 \pm 2.57 *
Base excess /mEq/L	4.72 \pm 1.32	-5.94 \pm 0.43 *
Blood pH	7.53 \pm 0.11	6.93 \pm 0.58 *
Anion gap/ mEq/L	7.22 \pm 2.64	13.48 \pm 2.76 *
Sodium/ mEq/L	137 \pm 4.55	136 \pm 6.28
Potassium /mEq/L	4.77 \pm 0.65	4.64 \pm 0.68
Chloride /mEq/L	94.75 \pm 2.43	95.77 \pm 5.38

* (P<0.05), Values are mean \pm standard error of mean

Furthermore significant increase (p<0.05) in haptoglobin and fibrinogen have been encountered in diseased cattle than in controls, Table 5.

Table(5):Haptoglobin and fibrinogen values of cattle infected with Anaplasmosis and controls

Parameters	Control cattle	Infected cattle
Haptoglobin mg/100mL	0.24 \pm 0.6	0.83 \pm .27*
Fibrinogen mg/100mL	242.28 \pm 36.32	301.11 \pm 43.66*

* (P<0.05), Values are mean \pm standard error of mean

DISCUSSION

Results of hemogram indicated a significant decrease in TRBC, HB, PCV, reflecting Macrocytic hypochromic type of anemia, similar results were recorded by (5, 16,17). The cause of anemia during anaplasmosis may be multifactorial, the direct effect of the Rickettsia to the infected erythrocytes may be incriminated or decrease life span of RBCs and also suppression of hemopoitic system (18,19), Moreover extensive erythrophagocytosis initiated by *A.marginale* to erythrocytes and the anti-erythrocytic auto antibodies changes in bone marrow are an indication to bone marrow depression followed by anemia ,Therefore Hemoglobinuria was an unusual

clinical sign of anaplasmosis, because anemia results from extravascular opsonization and phagocytosis of parasitized erythrocytes by reticuloendothelial cells (20).

Examination of stained blood smears in the current work revealed that *Anaplasma marginale* appears as spherical bodies, dark red in color, near periphery of infected erythrocytes, these results were similar to those seen by (21,7,22). Leukocytosis which accompanied by increase in the lymphocytes were in agreement with that reported by (23,24). The increase in WBC is due to stimulation of lymphoid tissues and stem cells in the bone marrow by the causative agent and their reactant factors, Moreover Allison and Memkoth (25) added that leukocytosis occur as a result to lymphoid depletion and disorganization with massive lymphocytes. lymphocytosis especially in *Anaplasma* infected cattle agree with that recorded by Aubry and Geale (2) whose stated that lymphocytosis was marked during the formation of antibodies in response to antigen and during anaplasma infection.

There were no significant difference encountered in clotting factors indices in current study, similar results were detected by (26, 7,27).

Results of Indirect Elisa test revealed that (96.25%) of tested cattle were seropositive to *A.marginale*, similar results were also recorded by Coetzee (28) whose stated that Indirect Elisa test may be an alternative for increased and sensitive detection of acute and latent anaplasmosis, Moreover Molloy(29) and Hornok(30) added that ELISA using recombinant antigens which were developed as a more specific method for the serodiagnosis of Anaplasmosis.

For maintaining the pH range differences, Acid-base balance is a critical and required for various enzyme systems to function ideally in the body, However disorders are restricted to an alteration in CO_2 or HCO_3^- with or without a compensatory response (31). In current study results showed decrease blood pH and bicarbonate in infected cattle which indicated Metabolic acidosis, similarly results also mentioned by(32,33). Two types of metabolic acidosis have been mention previously, Secretional metabolic acidosis which caused by loss of bicarbonate rich fluid such as diarrhea or saliva and Titrational metabolic acidosis which caused by the presence of non- CO_2 acids that titrate bicarbonate causing a decreased HCO_3^- (34). Titration type metabolic acidosis were indicated when endogenous or exogenous acids in the plasma will increased (35,36), Titrational metabolic acidosis were detected in the current work.

It have been mentioned before that hypovolemia result in decreased blood perfusion and hypoxia were follow, therefore anaerobic metabolism become a consequence, thereby lactic acid accumulates and Hyperlactemia will result (37,38). Furthermore the negative Base excess were also indicate metabolic acidosis (39).

With Titrational metabolic acidosis, the anion gap is increased which were indicated in the present study, since anion gap is a reflected value based on the principle of electroneutrality which states that the total anions in the body must be equal to the total cations (40,41).

Blood gas analysis of infected cattle were also indicated the tissue hypoxia via decrease level of Percent of Oxygen Saturation which consider as an indicator of the percentage of hemoglobin saturated with oxygen at the time of the measurement(42). Moreover P_{CO_2} reflects the amount of carbon dioxide gas dissolved in the blood, Therefore is an independent measure of the respiratory component of acid-base balance and were decreased in hypoxia and metabolic acidosis, (4,1).

The reactive Inflammatory response to any tissue injury is a mechanism through which the host sets up defense against further injury and starts the healing process (43), The early and immediate set of inflammatory reactions is known as acute phase

response (APR)(44). One of the predominant features of APR is changes in the concentrations of a number of plasma proteins associated with the host response(45). These changes are mainly the result of alterations in acute phase proteins synthesis in the liver (46). Haptoglobin has been one of the acute phase proteins most commonly monitored as a marker of inflammation in cattle (47,48).The function of the APR is to prevent tissue damage, and initiation of APR most commonly starts by the release of inflammatory mediators from tissue macrophages or blood monocyte cells that gather at the site of damage, these inflammatory mediators set off both the local and systemic inflammatory processes(49). The main function of haptoglobin is binding free hemoglobin and the hemoglobin binding property has a bacteriostatic effect, as it limits free iron available for bacteria (44). In current study results showed increase values of haptoglobin in cattle affected with Anaplasmosis ,same results were also mentioned by (50).

In current work increase fibrinogen level were also indicated .Fibrinogen is a plasma protein that considers as an acute phase protein in most species, including cattle, Therefore evaluation of this protein was found to be particularly useful in detecting inflammatory diseases (51).

CONCLUSIONS

Anaplasmosis were affected cattle and exhibited different clinical signs, a significant changes were noticed between diseased and control animals in hemogram ,acid base balance and blood gas analysis. Furthermore acute phase response were also detected in diseased cattle .Therefore The disease is responsible for substantial significant economic losses in endemic areas.

Acknowledgments

This article was supported by the College of Veterinary Medicine, University of Mosul, Iraq

ترابط حدوثية داء الانابلازموسز مع التوازن الحمضي-القاعدي، تحليل غازات الدم، استجابة الطور الحاد والصورة الدموية للأبقار

كمال الدين مهلهل السعد حسام محمد صبحي الامام
فرع الطب الباطني والوقائي , كلية الطب البيطري, جامعة الموصل, الموصل, العراق.

الخلاصة

كان الهدف من هذه الدراسة تقييم الصورة الدموية، التوازن الحمضي القاعدي، تحليل غازات الدم وقياس استجابة الطور الحاد للأبقار المحلية المصابة سريريا بداء الانابلازموسز في الموصل-العراق. إذ فحص (167) حيواناً من الأبقار المحلية بأعمار مختلفة تراوحت بين 2-5 سنوات ومن كلا الجنسين، شملت مجموعة الأبقار المصابة (80) حيواناً أظهرت علامات سريرية لداء الانابلازموسز الحاد وتم تأكيد اصابتها باستخدام مسحات دموية مصبوغة بصيغة الكمزا فضلاً عن استخدام اختبار الاليزا غير المباشر . أما مجموعة حيوانات السيطرة فشملت (20) حيواناً سليماً سريريا. أظهرت نتائج الدراسة انخفاض معنوي للعدد الكلي لكريات الدم الحمر وتركيز خضاب الدم وحجم خلايا الدم المرصوصة بالمقارنة مع مجموعة السيطرة، كما لوحظ ارتفاع معنوي لمعدلات الحجم الكروي وانخفاض معدلات تركيز خضاب الدم الكروي في الأبقار المصابة بداء الانابلازموسز بالمقارنة مع مجموعة السيطرة إذ كان فقر الدم من النوع ذي الكريات كبيرة الحجم قليلة الصباغ، وتراوحت النسبة المئوية للتطفل الدموي بين (12-21%) وبمعدل (15.57%). كما تبين حدوث زيادة ملحوظة في معدلات العدد الكلي لخلايا الدم البيض في الأبقار المصابة بداء الانابلازموسز

وكانت هذه الزيادة ناتجة من زيادة العدد التفريقي للخلايا اللمفية. كما تأكد إن 96.25% من الأبقار المفحوصة بتقنية الاليزا كانت مصابة بالنوع *A. marginale*. لم تظهر نتائج الدراسة أي اختلافات معنوية في معدلات معايير قيم عوامل تخثر الدم في حيوانات الدراسة، كما تناقصت معنوياً معدلات الضغط الجزئي لغاز ثاني اوكسيد الكربون P_{CO_2} ، نسبة تشبع الدم بالاكسجين Oxygen saturation percent (SO_2)، تركيز ايون البيكاربونات، الزيادة القاعدية Base excess، باها الدم Blood pH في الإبقار المصابة بداء الكمثرينات بالمقارنة مع مجموعة السيطرة، في حين لوحظ تزايد معنوي في معدلات فجوة الصاعد Anionic gap. إذ توضح إن نوع الحمض الأيضي هو من النوع حمض ابيضي معياري Titrational metabolic acidosis. وقد سجل الاختلاف في استجابة بروتينات الطور الحاد حيث لوحظ ارتفاع معنوي في معدلات الهابتوكلوبين Haptoglobin ومنشئ الليفين Fibrinogen في الأبقار المصابة بداء الانابلازموسز بالمقارنة مع مجموعة السيطرة.

REFERENCES

- 1-Radostitis, OM., Gay,CC., Blood,DC and Hinchliff, KW. Veterinary Medicine. A text book of the diseases of cattle, sheep, goats and horses.10th ed, WB Saunders Co. 2007. pp:1456-1460.
- 2-Aubry, P and Geale,DW. A Review of Bovine Anaplasmosis. Transbound. Emerg. Dis. 2011 .58 1–30.
- 3-DE waal ,DT. Anaplasmosis Control and Diagnosis in South Africa. Annals of the New York Academy of Sciences . 2006 . 916 (1) 474-483.
- 4-Smith, BP. Large animal internal medicine, 4th ed., New York, Mosby. 2004.pp: 1017.
- 5-Alsoad,KM.Clinical,hematological and biochemical studies of Anaplasmosis in local cattle breed .MSC ,thesis ,University of Mosul ,Mosul- Iraq . 1990. (In arabic).
- 6-Alsoad,KM., Al-obaidi,QT and Esmaeel,SA.Hematological and biochemical study on the effect of some common blood parasites in native goats in Mosul area. Iraqi .J .Vet. Sci . 2009. 23(1):101-106.
- 7-Alsoad, KM. Clinical, Hematological and Biochemical Studies of Anaplasmosis in Arabian One-Humped Camels (*Camelus dromedaries*).JAVA.2009.8(11) 2106-2109.
- 8-Ameen, KAH., Abdullah, BA and Abdul-Razaq, RA. Seroprevalence of *Babesia bigemina* and *Anaplasma marginale* in domestic animals in Erbil, Iraq. Iraqi .J .Vet. Sci. 2012. 26.supp, III. 109-114.
- 9-Bahzad. H. salih Mustafa. Clinical and Hematological study on ovine anaplasmosis in sulaimani province. Iraq. Bas.J.Vet.Res. 2012.11 (1).58-65.
- 10-Bush, BM. Veterinary laboratory manual. 1st ed., the Gresham press,London. 1975. pp: 113-167.
- 11-Weiss, DJ., Wardrop, KJ. Schalm's Veterinary Hematology ,6th ed(Ames, Wiley- 182 Blackwell). 2010.
- 12-Hiss, S., Mielenz,M., Bruckmaier RM and Sauerwein,H. Haptoglobin concentrations in blood and milk after endotoxin challenge and quantification of mammary Hp mRNA expression. J.Dairy Sci. 2004. 87: 3778–3784.
- 13-Shiroshita, Y., Tanaka,R ., Shibazaki,A and Yamane,Y. Accuracy of a Portable Blood Gas Analyzer Incorporating Optodes for Canine blood. J.Vet.Int.Med. 1999.13(6) 597- 600.
- 14-Katsuhiko,Y. Colorimetric determination of chloride in biological samples by using mercuric nitrate and phenylcarbazone. Bio.Tra.Ele.Res. 2002.85(1):87-94.

- 15-Leech, NL., Barrett, KC. and Morgan, GA. SPSS for intermediate statistics: use and interpretation .1st (ed.), Lawrence Erlbaum Asso.USA. 2007. pp: 20-51.
- 16-Alfonso, J., Medina, R., Fazzino, F., Caballero, H. Clinical and hematological changes in calves infected with *Anaplasma marginale*.Acta. Cient. Venez. 1996. 47(1): 50-57.
- 17-Alsaad,KM.The efficacy of imidocarb,oxytetracyline 20% and diminazine in the treatment of naturally infected cows with anaplasmosis . Iraqi .J .Vet. Sci. 2007b.21 (2) 307-316.(In Arabic).
- 18-Hofmann-lehmann R., Meli ML., Dreher, UM., Gönczi ,E., Deplazes, P., Braun U., Engels, M., Schüpbach, J., Jörgen, K., Thoma, R., Griot, C., stark, KDC., willi, B., Schmidt, J., Kocan ,KM .,Lutz, H. Concurrent infections with vector-borne pathogens associated with fatal haemolytic anemia in a cattle herd in switzerland. J. clin. microbiol. 2004. 42. 3775–3780.
- 19-Nazifi, S., Razavi, SM., Mansourian, M., Nikahval, B., Moghaddam, M. Studies on correlations among parasitaemia and some hemolytic indices in two tropical diseases (theileriosis and anaplasmosis) in Fars province of Iran. Trop. Ani.Heal. Prod. 2008. 40: 47–53.
- 20-Yasini, SP., Khaki ,Z., Rahbari, S., Kazemi, B., Salar Amoli, J., Gharabaghi ,A., Jalali ,SM . Hematologic and Clinical Aspects of Experimental Ovine Anaplasmosis Caused by *Anaplasma ovis* in Iran. Iranian J Parasitol.2012. 7(4) 91-98.
- 21-Rymaszewska, A and Grenda, S. Bacteria of the genus *Anaplasma* – characteristics of *Anaplasma* and their vectors: a review . Veterinarni Med. 2008. 53 (11)573–584.
- 22- Fosgate,GT., Jose, H. Urdaz-Rodr,G., Mark, D., Dunbar, D. Owen ,RG, Arthur, D., Pedro M., Georgina, L. Dobek, A. Rick, A. Diagnostic accuracy of methods for detecting *Anaplasma marginale* infection in lactating dairy cattle of Puerto Rico. J Vet Diagn Invest . 2010.22:192–199
- 23-Yousif, YA., Dimitri,RA., Dwivedi,SK. and Ahmed,NJ. Anaemia due to Anaplasmosis in Iraqi goats. I. Clinical and haemological features under field Conditions. Indian Vet.J. 1983.60: 576-578.
- 24-Omuse, JK. A comperative hematological picture of field cases of East cost fever, Anaplasmosis and babesiosis in bovine around kabete. In: "Tick born disease and their vector"1st ed by Wild J.K, Center for tropical Vet. Med. University of Edinburgh. 1987.pp: 181-187.
- 25-Allison RW, Memkoth JH. Anemia caused by Rickettsia, Mycoplasma, and Protozoa erythrocytes. In, Weiss DJ, Wardrop KJ (Eds): Schalm’s Veterinary Hematology. 6th ed. Blackwell Publishing Ltd,. 2010. pp:199-210.
- 26-Alsaad, KM. Comparative studies on the effect of common blood parasites on the blood picture and blood clotting factors in cattle. Basrah. J. Vet. Res. 2007a. 6: 16-19.
- 27-Pantanowitz, L. Mechanisms of thrombocytopenia in Tick-born diseases. Intrnet . J. Infec . Dis. 2003. 2: 1-7.
- 28-Coetzee JF., Schmidt, PL., Apley, MD., Reinbold, JB. and Kocan, KM. Comparison of the complement fixation test and competitive ELISA for serodiagnosis of *Anaplasma marginale* infection in experimentally infected steers. Am. J. Vet. Res. 2007. 68. 872–878.
- 29-Molloy, JB., Bowles, PM., Knowles, DP., Mcelwain, TF., Bock, RE., Kingston, TG., Blight, GW and Dalglieshr,J. Comparison of a competitive inhibition

- ELISA and the card agglutination test for detection of antibodies to *Anaplasma marginale* and *Anaplasma centrale* in cattle. Aust. Vet. J. 1999. 77. 245–249.
- 30-Hornok, S., Elek, V., de la Fuente, J., Naranjo, V., Farkas, R., Majoros, G., Foldvari, G. First serological and molecular evidence on the endemicity of *Anaplasma ovis* and *A. marginale* in Hungary. Vet Microbiol. 2007.122(3-4):316-322.
- 31-Dibartola, SP. Fluid, Electrolyte and Acid-Base Disorders in Small Animal Practice. 3rd ed. Elsevier, City, 2006.pp : 91-309.
- 32-Allen PC and Kuttler KL. Effect of *Anaplasma marginale* infection upon blood gases and electrolytes in splenectomized calves.J.Parasitol. 1981. 67(6):954-6.
- 33-McHolland ,LE and Caldwell,DR. Pyruvate metabolism by *Anaplasma marginale* in cell free culture .Can.J.Microbiol.1999.45(2)185-189.
- 34-Ayers, P and Warrington, L. Diagnosis and treatment of simple acid-base disorders. Nutr. Clin. Pract. 2008. 23(2):122-127.
- 35-Alexander C., Wiseman, MD and Stuart Linas, MD . Disorders of Potassium and Acid-Base Balance. Ame. J .Kidney Dis., 2005. 45(5):941-949.
- 36-Leisewitz, AL., Jacobson, LS., de Moraes, HS and Reyers,F. The mixed acid based disturbances of severe canine Babesiosis.J.Vet.Inter.Med.2001.15:445-452.
- 37-Sherlock, M., Healy, AM., Doherty,ML. Acid base balance in field cases of bovine babesiosis .Vet. Rec. 2003. 152(22):687-688.
- 38-Prieto-Callejero A., Sanz Marquez ,S., Perez Encinas, M. Linezolid-associated severe metabolic acidosis and anemia .Farm Hosp.2012.36(1):52-53.
- 39-Siggaard-andersen, O and Fogh-andersen, N. Base excess or buffer base (strong ion difference) as measure of a non-respiratory acid-base disturbance. Acta Anaesthesiol. Scand. 1995. 39 (106): 123-128.
- 40-Lolekha, PH and Lolekha, S . "Value of the anion gap in clinical diagnosis and laboratory evaluation". Clinical Chemistry .1983.29 (2): 279–83 .
- 41-Rose,B and Post,T. Clinical Physiology of Acid-Base and Electrolyte Disorders.1^{ed} 2000.pp:442-4445.
- 42-Beall, CB., Almasy, LA., Blangero, J., Williams-blangero, S., Brittenham, GM. Percent of Oxygen Saturation of Arterial Hemoglobin Among Bolivian Aymara at 3,900–4,000 m. Amer. J. Physic. Anthropol. 1999. 108:41–51.
- 43-Eckersall, PD. The time is right for acute phase protein assays. Vet. J. 2004. 168:3-5.
- 44-Cray,C., Zaias, J., Altman,NH. Acute Phase Response in Animals: A Review. Com. Med. J. 2009. 59(6): 517-526.
- 45-Ceron JJ., Martinez-Subiela, S., Ohno, K., Caldin, M. A seven -point plan for acute phase protein interpretation in companion animals. Vet .J . 2008.177(1)6–7.
- 46-Jain,S., Gautam,V and Naseem,S. Acute-phase proteins: As diagnostic tool. J Pharma. Bio. Sci . 2011. 3(1): 118–127.
- 47-Gruys, E., Toussaint, MJ., Niewold, TA., Koopmans, SJ., van Dijk, E., Meloen, RH. Monitoring health by values of acute phase proteins. Acta Histochem. 2006.108:229–232.
- 48-Ganheim, C., Alenius, S., Persson Waller, K. Acute phase proteins as indicators of calf herd health. Vet .J. 2007.173:645–651.

- 49-Eckersall, PD and Bell, R. Acute phase proteins: Biomarkers of infection and inflammation in veterinary medicine. *Vet. J.*2010. 185(1):23-27.
- 50-Nazifi, S ., Razavi, SM ., Kaviani, F ., Rakhshandehroo, E . Acute phase response in cattle infected with *Anaplasma marginale*.: *Vet. Microbiol.* 2011. 155(2-4):267-271.
- 51-van Wuijckhuise-Sjouke ,LA. Plasma fibrinogen as a parameter of the presence and severity of inflammation in horses and cattle. *Tijdschr Diergeneeskd.* 1984.109(21)869.