Effect Of Low Level Laser Therapy On Intra Oral Wound Healing

Harith H Kaskos
BDS, MSc (Lect)
Dept. of Oral and Maxillofacial Surgery
College of Dentistry, University of Mosul

Abdull H Al-Hasan
BV, MSc, PhD (Asst Prof)
Department Surgery and Theriogenology
College of Veterinary Medicine, University of Mosul

ABSTRACT

Aim: To evaluate the response of intraoral soft tissue by low level laser light, histologically and biochemically.

Materials and methods: The study was performed on (20) healthy rats. A clean two sided mucoperiosteal flap was raised opposite to the area of lower incisors. The rats were divided into two groups, ten for each. Histological specimens were taken at days 5th, 9th and 11th postoperative period. The dose of laser therapy in each session was 0.75 J/cm². Each rat in each group received this dose daily for 11th days at an interval of 15 minutes. Histopathological analysis were made for both groups detecting number of blood vessels (neovascularization), collagen arrangements and number of fibroblasts. Biochemical analysis of C-reactive protein (CRP) was also conducted. Results: Laser therapy showed enhancement of neovascularization at days 5 and 9 postoperatively. While collagen arrangement showed significant changes at the 5th, 9th and 11th postoperative days. Biochemical analysis showed significant increase of CRP at the 5th postoperative day. This increase was followed by a significant decrease at the 7th postoperative day in comparison with the control group. Conclusion: The present results indicated that low level laser therapy at a dose of 0.75 J/cm² enhanced wound healing.

Key words: Soft laser, wound healing, intra oral

INTRODUCTION

Wound healing can be defined as the summation of a number of processes which follow injury including coagulation, inflammation, matrix synthesis and deposition, angiogenesis, fibroplasias, epithelialization and remodeling. Many methods have been advocated to promote wound healing either biological such as Larva therapy, stem cell therapy, platelet rich plasma and gene therapy, physical such as electrical stimulation, magnetic field warming, miscellaneous therapies such as hyperbaric oxygen, topical oxygen therapy and laser or low level laser therapy (LLLT). Other names for the latter term include cold laser, soft laser, therapeutic laser and low power laser.
Laser energy is generated based on the principle of light amplification of stimulated emission of radiation. Laser has been used in several medical specialties and recently the dental profession has used laser both as a surgical tool and a biomodulating agent. Laser energy is highly coherent, polarized focused and monochromatic concentrated light which in contact with different tissues results in several effects on tissue depending upon the wave length and optical properties of irradiated tissue. LLLT has been used as an important tool for control of inflammatory process. The determination the level of some serum proteins may be useful for the differentiation of inflammatory conditions as well as the clinical conditions showing an increase or decrease in the acute phase response one of which is C-reactive protein (CRP). The magnitude of CRP varies directly with the degree of tissue damage. CRP plasma is usually low, increased quickly at the onset of acute inflammatory process and quickly falls when effective control of the process occurs.

The mechanism of action of LLLT on the cell is briefed as that: When the light photons are applied to the tissue, absorption in the mitochondria and cell membrane by chromophore results in an increase in ATP synthesis (Ca²⁺ and respiratory chain involved) with formation of a low amount ROS of Reactive O₂ species like H₂O₂, DNARNA protein synthesis is increased as well as mitosis and cellular proliferation resulting in rapid tissue healing and pain control. Since cell proliferation is one of the basic manifestations of any living organism, many authors through experimental studies confirmed that LLLT stimulate cell and fibroblast proliferation, increased the mean blood vessel sections on day 7 healing period of third degree burns in a histopathological study on rats. Non-invasive, low-power lasers with an output up to 500 mW have been reported to have stimulatory, anti-inflammatory and analgesic effects.

MATERIALS AND METHODS

Twenty healthy rats of both sex were enrolled in the experiment. The rats were placed in cages and received basal diet and tap water. The weight of each rat ranged from (250-350 g), aged 3-5 months. They were divided randomly into two groups,

1-Control group: The rats in this group were (10) that underwent an oral surgical procedure without postoperative laser irradiation.

2-Experimental group: The rats in this group were (10) that underwent an oral surgical procedure with scheduled laser irradiation.

Anesthesia of animal: The experimental rats were anaesthetized by using a combination of Ketamine and Xylazine at a dose of 80mg/kg and 10mg/kg respectively by intramuscular injection in the thigh muscle. The duration of anesthesia was about 45 minutes.

Surgical procedure: The surgical field sterilized by iodine solution (2%). A two sided intra-oral buccal mucoperiosteal flap was designed opposite to the area of present lower incisors using a Bard parker Scalpel handle (No 3) with a No 15 blade mounted on it, flap was reflected by atissue dissector and repositioned after it was irrigated with sterile normal saline solution 0.9% (Alnasser pharmaceutical chemicals, Co, Egypt) and sutured with 3/0 black silk (Kangyou Medical Instrument Co, China), one stitch in each vertical relaxed incision.

Laser system: The laser system used in this experimental study was made by Space Laser Corporation, Italy as shown in Figure(1), with He-Ne radiation only. Its characteristics are as follows: laser system class IV, He-Ne output power 0.006mW, He-Ne wave length 632.8nm.

Figure (1): Laser device
**Animal irradiation:**  
After completion of the surgical procedure, the experimental animal was placed 30 cm away from the source of laser beam according to the manufacturers instruction with the beam directed perpendicular to the wound site as shown in Figure (2). The first session began directly after finishing the surgical operation and then continued daily till day 11. Each session was for 15 minutes. As the power of He-Ne laser of the device is 0.006mW and the surface area of exposure is 12 cm², so the energy density (Dose) for each session was: 0.006mW*15 minutes*60 sec/ 12 cm² = 0.75 J/cm².

![Figure (2): Animal irradiation 30 cm away from laser beam](image)

**Blood sampling:**  
A blood sample was taken directly from the heart (1 ml) of rats who were sacrificed at experimental days (5 - 9 - 11) respectively for serological analysis of CRP. Biopsy was taken from the intraoral soft tissue that was raised and reflected for histological sectioning. NO systemic Antibi-otic or other medications was given for both experimental and control group which might affect the inflammatory process.

**Materials used for serological test:**  
The CRP kit includes CRP negative control, CRP positive control, CRP latex reagent, Standarded company, Spain plane tube. Disposable plastic syringe 1 ML Alma-dispo, Jordan. Histological evaluation included the following:-  
1- Neovascularization : (It was made by counting the number of blood vessel per square at magnification of 10X. Each field was selected randomly).
2- Fibroblast number(fibroplasia): (It was made by examining 5 locations selected randomly in each slide by counting the number of inflammatory cells within the indicating square at a power field of 10X).
3- Collagen arrangement (it was made by observing the collagen arrangement at 10Xpower field in all over the wound area and as follows based on the scoring system reported in Yu et al)\(^{(2)}\).

\[0=\text{None to sparse amount at edge.} \]
\[1=\text{Thin layer at edge.} \]
\[2=\text{Thin layer across wound.} \]
\[3=\text{Uniformly thick.} \]

The Student T-test was used for statistical comparison between experimental and control groups.

**RESULTS**

**CRP analysis:**  
At day 5, the serum concentration of C-reactive protein (CRP) for the laser treated group increased significantly\( p<0.05 \) as compared to control group. At day 9 post operatively, the CRP concentration decreased significantly\( p<0.001 \) as compared to control .The results are shown in Table (1).

<table>
<thead>
<tr>
<th>Postoperative days</th>
<th>Group tested</th>
<th>Mean mg/l</th>
<th>SE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Day 5</td>
<td>Control</td>
<td>280.000</td>
<td>30.0</td>
</tr>
<tr>
<td></td>
<td>Experimental</td>
<td>180.000</td>
<td>24.0</td>
</tr>
<tr>
<td>Day 9</td>
<td>Control</td>
<td>70.4 **</td>
<td>8.9</td>
</tr>
<tr>
<td></td>
<td>Experimental</td>
<td>320.4</td>
<td>60.5</td>
</tr>
</tbody>
</table>

\*Significant \( p<0.05 \). **very reduced, Significant ; \( p<0.001 \)

**Histopathological analysis:**

1- At day 5 (control group) diffuse extensive mononuclear(lymphocyte, macrophage) visualized along the edge of wound with very small newly (more numbers) formed blood vessels.

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The collagen density and arrangement were poor, little number of fibroblasts spread along with collagen fibers. The results are shown in Figure (3).

2-At day 5 (experimental group)
There was extensive vascularisation, better collagen arrangement in addition to more extensive fibroplasia in comparison to the control wound. The results as shown in Figure (4).

3-At day 9 (control group)
The wound line was filled with dense mature granulation tissue infiltrated by mononuclear inflammatory cells. The results are shown in Figure (5).

4-At day 9 (experimental group):
Similar histopathological changes were seen in the experimental wound at the same period. The results are shown in Figure (6).

5-At day 11 (control group)
Dense and mature granulation tissue was observed. The results are shown in Figure (7).

6-At day 11th (experimental group)
The healing process was more completed than the control group, with more fibrous tissue. The results are shown in Fig(8) Statistical analysis: - are

At day 5* Statistical comparison between the experimental and control group showed that revascularization, collagen arrangement and number of fibroblast were highly significant (P<0.01) Table(2).

Figure (8): Dense granulation tissue with severely increased in fibroblast at 11 postoperative day with laser irradiation.

Table (2): Histological finding at day 5 postoperatively

<table>
<thead>
<tr>
<th>Groups</th>
<th>Mean number of vessel +SE</th>
<th>Mean number of collagen arrangement +SE</th>
<th>Mean number of fibroblast cells +SE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>15.6 + 1.8</td>
<td>1.8 + 0.19</td>
<td>9.89 + 1.66</td>
</tr>
<tr>
<td>Experimental</td>
<td>7.5 + 0.7</td>
<td>0.6 + 0.22</td>
<td>3.79 + 0.59</td>
</tr>
</tbody>
</table>

* Significant p<0.001

Histopathological analysis:
1-At day5 (control group) Diffuse extensive mononuclear (lymphocyte,macrophage) visualized along the edge of wound with very small newly(more numbers) formed blood vessels. The collagen density and arrangement were poor, little number of fibroblasts spread along with collagen fibers . The results are shown in Figure(3).

Table (3): Histological finding at day 9 postoperatively

<table>
<thead>
<tr>
<th>Groups</th>
<th>Mean number of vessel +SE</th>
<th>Mean number of collagen arrangement +SE</th>
<th>Mean number of fibroblast cells +SE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>10.8 + 1.0</td>
<td>2.2 + 0.2**</td>
<td>6.9 + 0.69*</td>
</tr>
<tr>
<td>Experimental</td>
<td>7.0 + 0.3</td>
<td>1.6 + 0.3</td>
<td>4.0 + 0.29</td>
</tr>
</tbody>
</table>

* Significant p<0.05, ** Non Significant p>0.05

2-At day 5 (experimental group)
There was extensive vascularisation ,better collagen arrangement in addition to more extensive fibroplasia in comparison to the control wound. The results as shown in Figure (4).
### Table (4): Histological finding at day 11 postoperatively

<table>
<thead>
<tr>
<th>Groups</th>
<th>Mean number of vessel +SE</th>
<th>Mean number of collagen arrangement +SE</th>
<th>Mean number of fibroblast cells +SE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>7.3 +0.4**</td>
<td>2.7 + 0.2**</td>
<td>9.89 + 1.66*</td>
</tr>
<tr>
<td>Experimental</td>
<td>5.9 + 0.4</td>
<td>1.9 + 0.3</td>
<td>3.4 + 0.7</td>
</tr>
</tbody>
</table>

* Significant p<0.05, ** Non Significant p>0.05

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3-At day 9 (control group)

The wound line was filled with dense mature granulation tissue infiltrated by mononuclear inflammatory cells. The results are shown in Figure (5).

4-At day 9 (experimental group):

Similar histopathological changes were seen in the experimental wound at the same period. The results are shown in Figure (6).

5-At day 11 (control group)

Dense and mature granulation tissue was observed. The results are shown in Figure (7).

6-At day 11th (experimental group)

The healing process was more completed than the control group, with more fibrous tissue. The results are shown in Figure (8).

**Statistical analysis:**

At day 5:

Statistical comparison between the experimental and control group showed that revascularization, collagen arrangement and number of fibroblast were highly significant (P<0.01) Table (2).

At day 9:

Statistical comparison showed a significant increase in neo vascularisation (P<0.05) but with no significant differences in collagen arrangement between experimental and control group. A significant increase in the number of fibroblast in the experimental group was observed with the control group (P<0.001). This is shown in Table (3).

At day 11:

Statistical comparison showed a highly significant reduction in the number of inflammatory cell infiltration and a significant increase in the number of fibroblast in the experimental group compared with the control group. There were no significant (P>0.05) differences in the number of blood vessels and collagen in the experimental wound compared with the control one this is shown in Table (4).

**DISCUSSION**

Low level laser therapy (LLLT) is recognized as an effective therapeutic method in improving tissue healing by enhancing collagen synthesis as that suggested in many studies in vivo and vitro by promoting the transformation of fibroblasts to myofibroblast (21). In the present study, serological and histopathological changes were conducted at subsequent days (5, 9, and 11 days) postoperatively to evaluate the inflammatory phase and proliferative phase of wound healing. CRP (C-reactive protein) level was highly significant at day 5 postoperatively. This only means that laser light doesn't exacerbate the inflammatory process (prostaglandine E2 is an important chemical mediator in inflammatory process) but rather condenses the time frame from onset to resolution through an acceleration of the process. A active LLLT reduced prostaglandine E2 concentration as compared with the concentration before the treatment (22).

**Evaluation of neo-vascularisation:**

The significant increase of neo-vascularisation at days 5 and 9 postoperatively was due to an increase in the production of many angiogenic factors like that released from macrophages. These factors have been detected after laser irradiation (23).

**Evaluation of fibroblast cells number (fibroplasia):**

There was significant increase in fibroblast numbers at days 5, 9, and 11. The fibroblast became the prominent cell type at day 5 in clean non infected wounds (24). This may explain that as significant increase at days 5 and 9 postoperatively was noticed and only significant at day 11 where granulation tissue progressed to cellular fibrous tissue. The re-
searcher found that LLLT can increase the proliferative rate of human fibroblast(25).

Evaluation of collagen arrangement:
The better and highly significant collagen arrangement at day 5 postoperative period in the experimental group but with not significance at day 9 and 11 subsequent days came in agreement with researchers (26,27,28)

CONCLUSION
The study concluded that LLLT when used at a dose of 0.75 J/cm² at adjusted time intervals enhanced wound healing by primary intention.

REFERENCES
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